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**(54) UTILITY OF SNP MARKERS ASSOCIATED WITH MAJOR SOYBEAN PLANT MATURITY AND GROWTH HABIT GENOMIC REGIONS**

NUTZEN VON SNP-MARKERN IN ZUSAMMENHANG MIT GENOMISCHEN BEREICHEN FÜR REIFE UND WACHSTUMSGEWOHNHEIT WICHTIGER SOJABOHNNENARTEN

UTILITÉ DE MARQUEURS SNP ASSOCIÉS AVEC DES RÉGIONS MAJEURES DU PORT ET DE LA MATURITÉ DE PLANTES DE SOJA

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**Description**

## Field of the Invention

5 [0001] The invention includes methods for screening plants and seeds from the genus *Glycine* with markers associated with genomic regions that are related to the plant maturity and plant growth habit of *Glycine* plants.

## Background of the Invention

10 [0002] The soybean, *Glycine max* (L.) Merril, is a major economic crop worldwide and is a primary source of vegetable oil and protein (Sinclair and Backman, Compendium of Soybean Diseases, 3rd Ed. APS Press, St. Paul, MN, p. 106. (1989)). The growing demand for low cholesterol and high fiber diets has also increased importance of soybean as a health food.

15 [0003] Soybean varieties grown in the United States have a narrow genetic base. Six introductions, 'Mandarin,' 'Manchu,' 'mandarin' (Ottawa), "Richland," 'AK' (Harrow), and 'Mukden,' contributed nearly 70% of the germplasm represented in 136 cultivar releases. The genetic base of cultivated soybean could be widened through the use of exotic species. In addition, exotic species may possess such key traits as disease and stress resistance. At present, the traits of many exotic species are inaccessible in part due to limitations with crossing soybean plants from extremely different maturity groups. Most soybean variety development crosses are made between parents within 10 maturity days of each other. 20 If the parents differ greatly in maturity, the progeny plants segregate widely for maturity. In order for breeders to obtain and select for soybean plants of the desired maturity group, they must produce and maintain a large number of progeny plants, the practice of which is cost prohibitive.

25 [0004] Plant maturity and yield are closely associated in soybean. An increase of one day in maturity may be equivalent to a ~0.7 bu/A increase in yield. Conversely, a decrease in maturity is often penalized with a ~0.7 bu/A decrease in yield. The correlation of plant maturity and yield confounds the evaluation of potential quantitative trait loci (QTLs) and candidate genes associated with yield. The ability to genetically fix maturity within a soybean plant would be helpful and assist in elucidating traits associated with yield.

30 [0005] Soybean plants are short day plants, therefore flowering is initiated by short days due to a decrease in photoperiod (Garner & Allard, J. Agric. Res. 18, 553-606 (1920)). Consequently, photoperiod (day length) and temperature response of the soybean plant determine areas of plant adaptation. Due to photoperiod sensitivity, soybean genotypes are often grown in narrow zones of latitude to optimize yield. Northern soybean varieties, in contrast to Southern varieties, initiate flowering with longer days. Northern varieties planted south of their adaptation zone exhibit accelerated flowering, limited plant growth and reduced yield. Southern soybean varieties planted north of their adaptation zone will have delayed flowering with a potential for frost damage that may reduce yield.

35 [0006] Soybean plant varieties are classified based on bands of adaptation that are determined by latitude and day length. In North America, soybeans are categorized into 13 maturity groups with the designations ranging from maturity groups 000, 00, 0, and I through X. The earliest maturity group 000 soybeans are adapted to the north (45° latitude), while the latest maturity group X soybeans are adapted to regions near the equator. Soybean plants in maturity groups 000 to IV have indeterminate plant structure, while soybean plants in maturity groups V through X have determinate plant structure. Determinate varieties cease vegetative growth after the main stem terminates in a cluster of mature pods. Indeterminate varieties develop leaves and flowers simultaneously throughout a portion of their reproductive period, with one to three pods at the terminal apex. Early maturity varieties (000 to III) are adapted to northern latitudes with the maturity designation increasing in southern latitudes. The maturity group is determined by the maturity date. Plants are considered mature when 95% of the pods have reached their mature color. The maturity date is typically described as a measurement of days after August 31<sup>st</sup> in the northern hemisphere.

40 [0007] US 2006/288444 suggests the use of SNPs to identify soy bean plants with a certain trait value for maturity.

45 [0008] There is a need in the art of plant breeding to identify genomic regions associated with the maturity group of a soybean plant. At present, soybean breeders are limited to crossing plants within similar maturity groups. In addition, a number of traits, like oil levels, are influenced by latitude and maturity growing region. Therefore, there is a need for a rapid, cost-efficient method to pre-select for maturity group of soybean plants. The present invention includes a method for screening and selecting a soybean plant for a preferred plant maturity using single nucleotide polymorphism (SNP) technology.

## Brief Description of Figures

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[0009]

**Figure 1:** Influence of maturity group on percent oil in commercial soybeans.

**Figure 2:** Correlation of stearidonic acid (SDA) levels and GLA (gamma-linolenic acid) and latitude for mature soybean seeds. The soybean plants are transgenic and engineered to produce SDA and GLA.

**Figure 3:** Correlation of stearidonic acid (SDA) levels and latitude for mature soybean seeds over three trials. The soybean plants are transgenic and engineered to produce SDA.

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### Summary of the Invention

**[0010]** The present invention relates to a method of screening and selecting a soybean plant or soybean seed for maturity group association comprising:

- 10 (a) assaying genomic nucleic acids of said soybean plant or soybean seed for the presence of a genomic maturity marker genetically linked to a genomic region, wherein said genomic region is associated with a plant maturity group, said genomic maturity marker is within 10 cM, or within 10,000 kilobases of any of SEQ ID NOs: 143-213;  
 15 (b) determining whether said genomic maturity marker is homozygous or heterozygous;  
 (c) and selecting said soybean plant or seed based on said determination.

### Brief Description of Nucleic Acid Sequences

**[0011]**

- 20 SEQ ID NO: 1 is a forward PCR primer for the amplification of SEQ ID NO: 143.  
 SEQ ID NO: 2 is a reverse PCR primer for the amplification of SEQ ID NO: 143.  
 SEQ ID NO: 3 is a forward PCR primer for the amplification of SEQ ID NO: 144.  
 SEQ ID NO: 4 is a reverse PCR primer for the amplification of SEQ ID NO: 144.  
 25 SEQ ID NO: 5 is a forward PCR primer for the amplification of SEQ ID NO: 145.  
 SEQ ID NO: 6 is a reverse PCR primer for the amplification of SEQ ID NO: 145.  
 SEQ ID NO: 7 is a forward PCR primer for the amplification of SEQ ID NO: 146.  
 SEQ ID NO: 8 is a reverse PCR primer for the amplification of SEQ ID NO: 146.  
 30 SEQ ID NO: 9 is a forward PCR primer for the amplification of SEQ ID NO: 147.  
 SEQ ID NO: 10 is a reverse PCR primer for the amplification of SEQ ID NO: 147.  
 SEQ ID NO: 11 is a forward PCR primer for the amplification of SEQ ID NO: 148.  
 SEQ ID NO: 12 is a reverse PCR primer for the amplification of SEQ ID NO: 148.  
 35 SEQ ID NO: 13 is a forward PCR primer for the amplification of SEQ ID NO: 149.  
 SEQ ID NO: 14 is a reverse PCR primer for the amplification of SEQ ID NO: 149.  
 SEQ ID NO: 15 is a forward PCR primer for the amplification of SEQ ID NO: 150.  
 SEQ ID NO: 16 is a reverse PCR primer for the amplification of SEQ ID NO: 150.  
 40 SEQ ID NO: 17 is a forward PCR primer for the amplification of SEQ ID NO: 151.  
 SEQ ID NO: 18 is a reverse PCR primer for the amplification of SEQ ID NO: 151.  
 SEQ ID NO: 19 is a forward PCR primer for the amplification of SEQ ID NO: 152.  
 SEQ ID NO: 20 is a reverse PCR primer for the amplification of SEQ ID NO: 152.  
 45 SEQ ID NO: 21 is a forward PCR primer for the amplification of SEQ ID NO: 153.  
 SEQ ID NO: 22 is a reverse PCR primer for the amplification of SEQ ID NO: 153.  
 SEQ ID NO: 23 is a forward PCR primer for the amplification of SEQ ID NO: 154.  
 SEQ ID NO: 24 is a reverse PCR primer for the amplification of SEQ ID NO: 154.  
 50 SEQ ID NO: 25 is a forward PCR primer for the amplification of SEQ ID NO: 155.  
 SEQ ID NO: 26 is a reverse PCR primer for the amplification of SEQ ID NO: 155.  
 SEQ ID NO: 27 is a forward PCR primer for the amplification of SEQ ID NO: 156.  
 SEQ ID NO: 28 is a reverse PCR primer for the amplification of SEQ ID NO: 156.  
 SEQ ID NO: 29 is a forward PCR primer for the amplification of SEQ ID NO: 157.  
 55 SEQ ID NO: 30 is a reverse PCR primer for the amplification of SEQ ID NO: 157.  
 SEQ ID NO: 31 is a forward PCR primer for the amplification of SEQ ID NO: 158.  
 SEQ ID NO: 32 is a reverse PCR primer for the amplification of SEQ ID NO: 158.  
 SEQ ID NO: 33 is a forward PCR primer for the amplification of SEQ ID NO: 159.  
 SEQ ID NO: 34 is a reverse PCR primer for the amplification of SEQ ID NO: 159.  
 SEQ ID NO: 35 is a forward PCR primer for the amplification of SEQ ID NO: 160.  
 SEQ ID NO: 36 is a reverse PCR primer for the amplification of SEQ ID NO: 160.  
 SEQ ID NO: 37 is a forward PCR primer for the amplification of SEQ ID NO: 161.  
 SEQ ID NO: 38 is a reverse PCR primer for the amplification of SEQ ID NO: 161.



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SEQ ID NO: 329 is a probe for the detection of the SNP of SEQ ID NO: 200.  
 SEQ ID NO: 330 is a probe for the detection of the SNP of SEQ ID NO: 201.  
 SEQ ID NO: 331 is a probe for the detection of the SNP of SEQ ID NO: 201.  
 SEQ ID NO: 332 is a probe for the detection of the SNP of SEQ ID NO: 202.  
 5 SEQ ID NO: 333 is a probe for the detection of the SNP of SEQ ID NO: 202.  
 SEQ ID NO: 334 is a probe for the detection of the SNP of SEQ ID NO: 203.  
 SEQ ID NO: 335 is a probe for the detection of the SNP of SEQ ID NO: 203.  
 SEQ ID NO: 336 is a probe for the detection of the SNP of SEQ ID NO: 204.  
 SEQ ID NO: 337 is a probe for the detection of the SNP of SEQ ID NO: 204.  
 10 SEQ ID NO: 338 is a probe for the detection of the SNP of SEQ ID NO: 205.  
 SEQ ID NO: 339 is a probe for the detection of the SNP of SEQ ID NO: 205.  
 SEQ ID NO: 340 is a probe for the detection of the SNP of SEQ ID NO: 206.  
 SEQ ID NO: 341 is a probe for the detection of the SNP of SEQ ID NO: 206.  
 SEQ ID NO: 342 is a probe for the detection of the SNP of SEQ ID NO: 207.  
 15 SEQ ID NO: 343 is a probe for the detection of the SNP of SEQ ID NO: 207.  
 SEQ ID NO: 344 is a probe for the detection of the SNP of SEQ ID NO: 208.  
 SEQ ID NO: 345 is a probe for the detection of the SNP of SEQ ID NO: 208.  
 SEQ ID NO: 346 is a probe for the detection of the SNP of SEQ ID NO: 209.  
 SEQ ID NO: 347 is a probe for the detection of the SNP of SEQ ID NO: 209.  
 20 SEQ ID NO: 348 is a probe for the detection of the SNP of SEQ ID NO: 210.  
 SEQ ID NO: 349 is a probe for the detection of the SNP of SEQ ID NO: 210.  
 SEQ ID NO: 350 is a probe for the detection of the SNP of SEQ ID NO: 211.  
 SEQ ID NO: 351 is a probe for the detection of the SNP of SEQ ID NO: 211.  
 SEQ ID NO: 352 is a probe for the detection of the SNP of SEQ ID NO: 212.  
 25 SEQ ID NO: 353 is a probe for the detection of the SNP of SEQ ID NO: 212.  
 SEQ ID NO: 354 is a probe for the detection of the SNP of SEQ ID NO: 213.  
 SEQ ID NO: 355 is a probe for the detection of the SNP of SEQ ID NO: 213.

## Definitions

- 30 [0012] A "maturity group value" can be any indicative number, symbol, or combination of both that provides an indication of when a plant will mature.
- [0013] A "dominant maturity allele" is an allele that, when present either in single copy (heterozygous) or two copies (homozygous), affects the maturity of the plant.
- 35 [0014] A "recessive maturity allele" is an allele that, when present in one copy (heterozygous), does not affect the maturity of a plant.
- [0015] As used herein, determinate growth habit refers to ceasing of vegetative growth after the main stem terminates in a cluster of mature pods.
- 40 [0016] As used herein, indeterminate growth habit refers to the development of leaves and flowers simultaneously throughout a portion of their reproductive period, with one to three pods at the terminal apex.
- [0017] As used herein, an allelic combination is the combination of alleles present at more than one characterized location or loci. An example of an allelic combination is allelic combination 10, which is homozygous dominant at maturity genomic region 1; homozygous recessive at maturity genomic region 2; and homozygous dominant at maturity genomic region 3.
- 45 [0018] As used herein, "line" refers to a group of individual plants from the similar parentage with similar traits. An "elite line" is any line that has resulted from breeding and selection for superior agronomic performance. Additionally, an elite line is sufficiently homogenous and homozygous to be used for commercial production. Elite lines may be used in the further breeding efforts to develop new elite lines. An elite plant is any plant from an elite line.
- 50 [0019] As used herein, "a trait" refers to an observable and/or measurable characteristic of an organism, such as a trait of a plant, *for example*, tolerance to an herbicide, insect and microbe. A trait can be conventional and transgenic. Non-limiting examples of traits include herbicide tolerance, increased yield, insect control, fungal disease resistance, virus resistance, nematode resistance, bacterial disease resistance, mycoplasma disease resistance, altered oils production, high oil production, high protein production, germination and seedling growth control, enhanced animal and human nutrition, low raffinose, environmental stress resistant, increased digestibility, industrial enzymes, pharmaceutical proteins, peptides and small molecules, improved processing traits, improved flavor, nitrogen fixation, hybrid seed production, reduced allergenicity, biopolymers, and biofuels.
- 55 [0020] As used herein, "a transgene" refers to a foreign gene that is placed into an organism by the process of plant transformation. In certain aspects, a soybean plant provided by the invention may comprise one or more transgene(s).

[0021] As used herein, "altered" means increased or decreased at maturity. In this aspect, a mature seed as defined by a seed that is harvested in the field for commercial agricultural practices, such as sale for feed. In an aspect, a soybean plants are selected for preferred geographies for expression of at least one phenotypic trait. The phenotypic trait includes altered levels of a substance or a molecule, such as proteins, oils, or gamma linolenic acid. "Altered" can include any

5 relative increase or decrease of function or production of a gene product of interest, in an aspect up to and including complete elimination of function or production of that gene product. When levels of a gene product are compared, such a comparison is preferably carried out between organisms with a similar genetic background. Preferably, a similar genetic background is a background where the organisms being compared share 50% or greater, more preferably 75% or greater, and, even more preferably 90% or greater sequence identity of nuclear genetic material. In another aspect, a similar genetic background is a background where the plants are isogenic except for one or more of the disclosed markers.

10 [0022] As used herein, a "cultivar" is a race or variety of a plant that has been created or selected intentionally and maintained through cultivation.

15 [0023] As used herein, the term "tissue culture" indicates a composition comprising isolated cells of the same or a different type or a collection of such cells organized into parts of a plant.

#### Detailed Description of the Invention

[0024] Determination of the maturity group value of a soybean plant or seed is important in selecting where a soybean plant should be grown. An aspect of the present invention provides for a method of establishing where a plant or seed should be grown. A suitable region of a soybean plant or seed can be established. Establishment of a region can include selection of a suitable maturity belt region. Maturity belts range in the United States from 000 in the extreme northern U.S. to VIII in the southern Gulf Coast states. The present invention can also be used to determine other maturity belts including IX and X. The present invention can further be utilized to determine whether a plant is suitable for one, two, or more maturity belts or regions.

20 [0025] A suitable geographic region can be selected using a method of the present invention. In addition to maturity belts, other geographic regions that can be selected include maturity group 0 regions, such as and without limitation, Western Maine, North Dakota, Central Montana, Northwestern Oregon; maturity group 1 regions, such as and without limitation, northern Wisconsin, South Dakota; maturity group 2 regions, such as and without limitation, Vermont, Southern Massachusetts, Northern Connecticut, New York, Central Florida, Michigan, Northern Illinois, Southern Wisconsin, Iowa, Nebraska, Colorado, Central California; maturity group 3 regions, such as and without limitation, Western New Hampshire, Pennsylvania, Ohio, Indiana, Southern Illinois, Northern Missouri, Kansas, Southeast Wyoming, Colorado; maturity group 4 regions, such as and without limitation, Maryland, Northern Virginia, Kentucky, Western West Virginia, Central Missouri, Texas, Western Oklahoma; maturity group 5 regions, such as and without limitation, Central Virginia, North Carolina, Central and Western North Carolina, Mississippi, Louisiana, Tennessee; maturity group 6 regions, such as and without limitation, North Carolina, Eastern South Carolina; and maturity group 7 regions, such as and without limitation, Georgia, and Alabama. In another aspect, a disclosed seed can be sent to a geographic region that is desirable to optimize a trait, such as yield.

25 [0026] The present invention also provides methods of selecting a suitable geographic region and methods for determining the maturity group of a soybean plant or seed by genotypic analysis. One aspect of the present invention includes a method of establishing where a soybean plant should be grown by obtaining DNA from the soybean plant; and determining if an allele within maturity genomic region 1 is homozygous or heterozygous using marker SEQ ID NO: 151.

30 [0027] The present invention allows the determination of allelic combinations. Allelic combinations can be any combination of alleles. In one aspect, it can be a combination of 2, 3, 4, 5, 6, 7, or 8 pairs of alleles that occupy a genetic locus. In another aspect, the alleles can be located within 2, 3, 4, 5, 6, 7, or 8 or more maturity genomic regions. Such 35 maturity regions can be selected from maturity genomic region 1, maturity genomic region 2, maturity genomic region 3, maturity genomic region 4, maturity genomic region 5, maturity genomic region 6, maturity genomic region 7, or maturity genomic region 8.

40 [0028] Alleles at any combination of maturity regions can be determined individually or in combination. One illustrative combination is a combination of more than one pair of alleles at maturity regions 1, 2, and 3. Another illustrative combination is a combination of more than one pair of alleles at maturity regions 1 and 2. "Allelic combinations" is intended to include, without limitation, any of homozygous dominant, homozygous recessive, and heterozygous alternatives at a particular locus.

45 [0029] Determination of an allele or the combination of alleles at a locus or loci can be carried out by any appropriate methodology. In an aspect, various assays can be used, such as a Taq-Man® assay, Real Time PCR, and nucleic acid sequencing, and simple sequence repeat mapping, to detect the genotype. In an aspect of the present invention, the assay includes a disclosed nucleic acid molecule. Nucleic acids include deoxynucleic acids (DNA) and ribonucleic acids (RNA) and functionally equivalent analogues thereof.

50 [0030] Nucleic acids for use in the present invention can be obtained from a plant, such as from a plant part which

includes a leaf, vascular tissue, flower, pod, seed, root, stem, or a portion of any.

[0031] In one aspect, nucleic acids are obtained from a plant or plant part using a non-destructive method. In an aspect, the plant part is a seed. In an aspect, the nucleic acids are obtained from a seed in a non-destructive manner, which provides for a seed that is viable. For example, DNA can be obtained from a seed by chipping the seed with a sharp knife at a part furthest away from the 'eye' or by pricking carefully with a needle to puncture the seed. Any method that will obtain DNA for analysis or allow *in situ* analysis of the DNA can be used provided that the plant or plant part retains the ability to grow. If DNA is taken from a seed and the seed is still viable, the method can be considered non-destructive. Exemplary methods to sample seeds without affecting the germination viability of the seeds are detailed in US Patent Application Publication 2006/0042527. In an aspect, seeds are sampled by feeding the seeds individually to a sampling station, removing a sample from the seed in the sampling station, conveying the sample to a compartment in a sample tray and conveying the seed to a corresponding compartment in a seed tray.

[0032] In an aspect, the maturity genomic region associated with plant maturity and plant growth habit is introduced or selected within the genus *Glycine*. The genus *Glycine* includes the wild perennial soybeans and have a wide array of genetic diversity. For example, the cultivated soybean (*Glycine max* (L.) Merr.) and its wild annual progenitor (*Glycine soja* (Sieb. and Zucc.)) belong to the subgenus *Soja*, contain  $2n = 40$  chromosomes, are cross-compatible, usually produce vigorous fertile F<sub>1</sub> hybrids, and carry similar genomes. Crosses between cultivated *Glycine* species and wild perennial *Glycine* species have variable success among accessions. The present invention further provides that the selected plant is from the group consisting of members of the genus *Glycine*, more specifically from the group consisting of *Glycine arenaria*, *Glycine argyrea*, *Glycine canescens*, *Glycine clandestina*, *Glycine curvata*, *Glycine cyrtoloba*, *Glycine falcata*, *Glycine latifolia*, *Glycine latrobeana*, *Glycine max*, *Glycine microphylla*, *Glycine pescadrensis*, *Glycine pindanica*, *Glycine rubiginosa*, *Glycine soja*, *Glycine sp.*, *Glycine stenophita*, *Glycine tabacina*, and *Glycine tomentella*. In an aspect the plant is selected from an elite *Glycine max* line.

[0033] The present invention also provides a soybean plant selected for a desired plant maturity by screening for a maturity marker in the soybean plant or seed, the selection comprising assaying genomic nucleic acids for the presence of a marker molecule that is genetically linked to a genomic region associated with a plant maturity in the soybean plant, where the genomic region is also located on a linkage group associated with a soybean plant of a preferred plant maturity. The disclosed methods include determining if a locus contains a polymorphism, or is homozygous or heterozygous at a maturity region selected from maturity genomic region 1, maturity genomic region 2, maturity genomic region 3, maturity genomic region 4, maturity genomic region 5, maturity genomic region 6, maturity genomic region 7, and/or maturity genomic region 8 by detecting a polymorphism within a nucleic acid molecule comprising a sequence or fragment thereof selected from the group consisting of SEQ ID NOs: 143-174, or complements thereof. The present invention includes the identification of alleles at eight maturity group regions. These regions are termed maturity genomic regions 1 through 8.

[0034] The state of homozygosity or heterozygosity and dominance or recessivity of maturity genomic region 1 can be monitored by assaying for an allele of 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, or 13 or more genetic markers selected from the group consisting of NS0093385, NS0093976, NS0096829, NS0097798, NS0098982, NS00995929, NS0099746, NS0103749, NS0123747, NS0124601, NS0125408, NS0128378, and NS0135390. SNP marker DNA sequences for region 1 include those presented as SEQ ID NO: 143 through SEQ ID NO: 155 and can be amplified using the primers indicated as SEQ ID NO: 1 through SEQ ID NO: 26 with probes indicated as SEQ ID NO: 214 through SEQ ID NO: 239. In another aspect, a maturity genomic region 1 is a region associated with SEQ ID NOs: 143-149, 154-155. In another aspect, a maturity genomic region 1 is a region associated with SEQ ID NO: 149 or SEQ ID NO: 151 or both. In an aspect, maturity genomic region 1 can span 1 centiMorgan (cM), 5 cM, 10 cM, 15 cM, 20 cM, or 30 cM either side of SEQ ID NO: 149 or SEQ ID NO: 151.

[0035] An aspect of the method of the present invention includes a method of determining if a soybean seed will grow into a soybean plant having a maturity group of III-VI by determining a homozygous or heterozygous marker within the soybean seed using a marker with the nucleic acid sequence of SEQ ID NO: 151. In a preferred aspect, the homozygous marker can be recessive or dominant. In another preferred aspect, the maturity of the plant is delayed where the marker is homozygous dominant.

[0036] Another aspect of the method of the present invention includes a method of determining if a soybean seed will grow into a soybean plant having a maturity group between 0.0 - III.0 comprising determining if an 11-basepair insertion within the nucleic acid sequence of SEQ ID NO: 149 exists in the soybean seed.

[0037] The state of homozygosity or heterozygosity and dominance or recessivity of maturity genomic region 2 may be monitored by assaying for an allele of 1, 2, 3, 4, 5, or 6 or more genetic markers including those selected from the group consisting of NS0118907, NS0122182, NS0126989, NS097952, NS0123506 and NS0095677. SNP marker DNA sequences for region 2 include those presented as SEQ ID NO: 156 through SEQ ID NO: 161 and can be amplified using the primers indicated as SEQ ID NO: 27 through SEQ ID NO: 38 with probes indicated as SEQ ID NO: 240 through SEQ ID NO: 251. In another aspect, a maturity genomic region 2 is a region associated with SEQ ID NO: 158. In another aspect, a maturity genomic region 2 is a region associated with SEQ ID NOs: 156-161. In an aspect, maturity genomic region 2 can span 1 cM, 5 cM, 10 cM, 15cM, 20 cM, or 30 cM either side of SEQ ID NO: 158.

**[0038]** The state of homozygosity or heterozygosity and dominance or recessivity of maturity genomic region 3 may be monitored by assaying for an allele of 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, or 13 or more genetic markers including those selected from the group consisting of NS0098853, NS0092561, NS0093197, NS0094891, NS0096225, NS0103853, NS0113929, NS0115535, NS0121511, NS0136544, NS0119569, NS0123708, and NS0114317. SNP

5 marker DNA sequences for region 3 including those presented as SEQ ID NO: 162 through SEQ ID NO: 174 and can be amplified using the primers indicated as SEQ ID NO: 39 through SEQ ID NO: 64 with probes indicated as SEQ ID NO: 252 through SEQ ID NO: 277. In another aspect, a maturity genomic region 3 is a region associated with SEQ ID NOS: 164, 167, 171-174. In another aspect, a maturity genomic region 3 is a region associated with SEQ ID NO: 169. In an aspect, maturity genomic region 3 can span 1 cM, 5 cM, 10 cM, 15cM, 20 cM, or 30 cM either side of SEQ ID NO: 169.

10 **[0039]** The state of homozygosity or heterozygosity and dominance or recessivity of maturity genomic region 4 may be monitored by assaying for an allele of 1, 2, 3, 4, 5, or 6 or more genetic markers including those selected from the group consisting of NS0092743, NS0098176, NS0100078, NS0137415, NS0095530, and NS0129004. SNP marker DNA sequences for region 4 are presented as SEQ ID NO: 175 through SEQ ID NO: 180 and can be amplified using the primers indicated as SEQ ID NO: 65 through SEQ ID NO: 76 with probes indicated as SEQ ID NO: 278-289. In

15 another aspect, a maturity genomic region 4 is a region associated with SEQ ID NO: 178. In an aspect, maturity genomic region 4 can span 1 cM, 5 cM, 10 cM, 15cM, 20 cM, or 30 cM either side of SEQ ID NO: 178. An aspect of the method of the present invention includes a method of detecting maturity genomic region 4 by detecting an allele using a marker selected from any of SEQ ID NO: 175-180.

20 **[0040]** The state of homozygosity or heterozygosity and dominance or recessivity of maturity genomic region 5 may be monitored by assaying for an allele of 1, 2, 3, 4, 5, 6, 7, 8, or 9 or more genetic markers including those selected from the group consisting of NS0120015, NS0113878, NS0101863, NS0115066, NS0123168, NS0119165, NS0123724, NS0103446, and NS0099024. SNP marker DNA sequences for region 5 including those presented as SEQ ID NO: 181 through SEQ ID NO: 189 and can be amplified using the primers indicated as SEQ ID NO: 77 through SEQ ID NO: 94 with probes indicated as SEQ ID NO: 290 through SEQ ID NO: 307. In another aspect, a maturity genomic region 5 is a region associated with SEQ ID NO: 187. In an aspect, maturity genomic region 5 can span 1 cM, 5 cM, 10 cM, 15cM, 20 cM, or 30 cM either side of SEQ ID NO: 187. An aspect of the method of the present invention includes a method of detecting maturity genomic region 5 by detecting an allele using a marker selected from any of SEQ ID NO: 181-189.

25 **[0041]** The state of homozygosity or heterozygosity and dominance or recessivity of maturity genomic region 6 may be monitored by assaying for an allele of 1, 2, 3, 4, 5, 6, or 7 or more genetic markers including those selected from the group consisting of NS0116125, NS0125770, NS0103755, NS0125713, NS0124590, NS0119281, and NS0102717. SNP marker DNA sequences for region 6 including those presented as SEQ ID NO: 190 through SEQ ID NO: 196 and can be amplified using the primers indicated as SEQ ID NO: 95 through SEQ ID NO: 108 with probes indicated as SEQ ID NO: 308 through SEQ ID NO: 321. In another aspect, a maturity genomic region 6 is a region associated with SEQ ID NO: 192. In an aspect, maturity genomic region 6 can span 1 cM, 5 cM, 10 cM, 15cM, 20 cM, or 30 cM either side of SEQ ID NO: 192. An aspect of the method of the present invention includes a method of detecting maturity genomic region 6 by detecting an allele using a marker selected from any of SEQ ID NO: 190-196.

30 **[0042]** The state of homozygosity or heterozygosity and dominance or recessivity of maturity genomic region 7 may be monitored by assaying for an allele of 1, 2, 3, 4, 5, 6, or 7 or more genetic markers including those selected from the group consisting of NS0095211, NS0099531, NS0099417, NS0097307, NS0103004, NS0102630, and NS0102915. SNP DNA sequences for region 7 including those presented as SEQ ID NO: 197 through SEQ ID NO: 203 and can be amplified using the primers indicated as SEQ ID NO: 109 through SEQ ID NO: 122 with probes indicated as SEQ ID NO: 322 through SEQ ID NO: 335. In another aspect, a maturity genomic region 7 is a region associated with SEQ ID NO: 202. In an aspect, maturity genomic region 7 can span 1 cM, 5 cM, 10 cM, 15cM, 20 cM, or 30 cM either side of SEQ ID NO: 202. An aspect of the method of the present invention includes a method of detecting maturity genomic region 7 by detecting an allele using a marker selected from any of SEQ ID NO: 197-203.

35 **[0043]** The state of homozygosity or heterozygosity and dominance or recessivity of maturity genomic region 8 may be monitored by assaying for an allele of 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 or more genetic markers including those selected from the group consisting of N0102362, NS0100652, NS0117716, NS0119574, NS0127728, NS0099639, NS0103255, NS0119106, NS0101020, and NS0101779. SNP DNA sequences for region 8 including those presented as SEQ ID NO: 204 through SEQ ID NO: 213 and can be amplified using the primers indicated as SEQ ID NO: 123 through SEQ ID NO: 142 with probes indicated as SEQ ID NO: 336 through SEQ ID NO: 355. In another aspect, a maturity genomic region 8 is a region associated with SEQ ID NO: 204. In an aspect, maturity genomic region 8 can span 1 cM, 5 cM, 10 cM, 15cM, 20 cM, or 30 cM either side of SEQ ID NO: 204. An aspect of the method of the present invention includes a method of detecting maturity genomic region 8 by detecting an allele using a marker selected from any of SEQ ID NO: 204-213.

40 **[0044]** The disclosed nucleic acid molecules or fragments thereof are capable of specifically hybridizing to other nucleic acid molecules, also included in the present invention, under certain circumstances. In an aspect, the nucleic acid molecules may contain any of SEQ ID NO: 143-213, complements thereof and fragments of any. In another aspect, the

nucleic acid molecules include nucleic acid molecules that hybridize, for example, under high or low stringency, substantially homologous sequences, or that have both to these molecules. As used herein, two nucleic acid molecules are capable of specifically hybridizing to one another if the two molecules are capable of forming an anti-parallel, double-stranded nucleic acid structure. A nucleic acid molecule is the "complement" of another nucleic acid molecule if they exhibit complete complementarity. As used herein, molecules exhibit "complete complementarity" when every nucleotide of one of the molecules is complementary to a nucleotide of the other. Two molecules are "minimally complementary" if they can hybridize to one another with sufficient stability to permit them to remain annealed to one another under at least conventional "low-stringency" conditions. Similarly, the molecules are "complementary" if they can hybridize to one another with sufficient stability to permit them to remain annealed to one another under conventional "high-stringency" conditions. Conventional stringency conditions are described by Sambrook et al., In: Molecular Cloning, A Laboratory Manual, 2nd Edition, Cold Spring Harbor Press, Cold Spring Harbor, New York (1989)), and by Haymes et al., In: Nucleic Acid Hybridization, A Practical Approach, IRL Press, Washington, DC (1985).

**[0045]** Departures from complete complementarity are therefore permissible, as long as such departures do not completely preclude the capacity of the molecules to form a double-stranded structure. In order for a nucleic acid molecule to serve as a primer or probe it need only be sufficiently complementary in sequence to be able to form a stable double-stranded structure under the particular solvent and salt concentrations employed.

**[0046]** As used herein, a substantially homologous sequence is a nucleic acid sequence that will specifically hybridize to the complement of the nucleic acid sequence to which it is being compared under high stringency conditions. The disclosed nucleic-acid probes and primers can hybridize under stringent conditions to a target DNA sequence. The term "stringent hybridization conditions" is defined as conditions under which a probe or primer hybridizes specifically with a target sequence(s) and not with non-target sequences, as can be determined empirically. The term "stringent conditions" is functionally defined with regard to the hybridization of a nucleic-acid probe to a target nucleic acid (i.e., to a particular nucleic-acid sequence of interest) by the specific hybridization procedure discussed in Sambrook et al., 1989, at 9.52-9.55. See also, Sambrook et al., 1989 at 9.47-9.52, 9.56-9.58; Kanehisa, Nucl. Acids Res. 12:203-213, 1984; and Wetmur and Davidson, J. Mol. Biol. 31:349-370, 1968. Appropriate stringency conditions that promote DNA hybridization are, for example, 6.0 x sodium chloride/sodium citrate (SSC) at about 45°C, followed by a wash of 2.0 x SSC at 50°C, are known to those skilled in the art or can be found in Current Protocols in Molecular Biology, John Wiley & Sons, N.Y., 1989, 6.3.1-6.3.6. For example, the salt concentration in the wash step can be selected from a low stringency of about 2.0 x SSC at 50°C to a high stringency of about 0.2 x SSC at 50°C. In addition, the temperature in the wash step can be increased from low stringency conditions at room temperature, about 22°C, to high stringency conditions at about 65°C. Both temperature and salt may be varied, or either the temperature or the salt concentration may be held constant while the other variable is changed.

**[0047]** For example, hybridization using DNA or RNA probes or primers can be performed at 65°C in 6x SSC, 0.5% SDS, 5x Denhardt's, 100 µg/mL nonspecific DNA (e.g., sonicated salmon sperm DNA) with washing at 0.5x SSC, 0.5% SDS at 65°C, for high stringency.

**[0048]** It is contemplated that lower stringency hybridization conditions such as lower hybridization and/or washing temperatures can be used to identify related sequences having a lower degree of sequence similarity if specificity of binding of the probe or primer to target sequence(s) is preserved. Accordingly, the disclosed nucleotide sequences can be used for their ability to selectively form duplex molecules with complementary stretches of DNA, RNA, or cDNA fragments. Detection of DNA segments via hybridization is well-known to those of skill in the art, and thus depending on the application envisioned, one will desire to employ varying hybridization conditions to achieve varying degrees of selectivity of probe towards target sequence and the method of choice will depend on the desired results.

**[0049]** As used herein, an agent, be it a naturally occurring molecule or otherwise may be "substantially purified", if desired, referring to a molecule separated from substantially all other molecules normally associated with it in its native state. More preferably a substantially purified molecule is the predominant species present in a preparation. A substantially purified molecule may be greater than 60% free, preferably 75% free, more preferably 90% free, and most preferably 95% free from the other molecules (exclusive of solvent) present in the natural mixture. The term "substantially purified" is not intended to encompass molecules present in their native state. The agents will preferably be "biologically active" with respect to either a structural attribute, such as the capacity of a nucleic acid to hybridize to another nucleic acid molecule, or the ability of a protein to be bound by an antibody (or to compete with another molecule for such binding). Alternatively, such an attribute may be catalytic, and thus involve the capacity of the agent to mediate a chemical reaction or response.

**[0050]** The agents may also be recombinant. As used herein, the term recombinant means any agent (e.g. DNA, peptide), that is, or results, however indirect, from human manipulation of a nucleic acid molecule.

**[0051]** The agents may be labeled with reagents that facilitate detection of the agent (e.g. fluorescent labels (Prober et al., Science 238:336-340 (1987); European Patent No. 144914), chemical labels (U.S. Patent no. 4,582,789; U.S. Patent No. 4,563,417), modified bases (European Patent No. 119448)).

**[0052]** In an aspect, an agent will specifically hybridize to one or more of the nucleic acid molecules set forth in SEQ

ID NO: 143 through SEQ ID NO: 213 or complements thereof or fragments of either under moderately stringent conditions, for example at about 2.0 x SSC and about 65°C. In an aspect, a nucleic acid will specifically hybridize to one or more of the nucleic acid molecules set forth in SEQ ID NO: 143 through SEQ ID NO: 213 or complements or fragments of either under high stringency conditions.

**[0053]** Agents include genetic markers. Examples of such markers include nucleic acid molecules comprising nucleic acid sequences selected from the group consisting of SEQ ID NOs: 143-213. Examples of public marker databases include, for example: Soybase, an Agricultural Research Service, and United States Department of Agriculture. Other genetic markers are disclosed within.

**[0054]** Agents include fragment nucleic acid molecules. Fragments can contain significant portions of, or indeed most of, SEQ ID NOs: 143-213. In an aspect, the fragments are between 100 and 200 consecutive residues, 150 and 300 consecutive residues, 50 and 150 consecutive residues, or 20 and 50 consecutive residues long of a nucleic molecule. In another aspect, the fragment comprises at least 50, 100, 200, 300, 400, or 500 consecutive residues of SEQ ID NOs: 143-213. In an aspect, a fragment nucleic acid molecule is capable of selectively hybridizing to SEQ ID NOs: 143-213.

**[0055]** In one aspect, a preferred marker nucleic acid molecule has the nucleic acid sequence set forth in SEQ ID NO: 143 through SEQ ID NO: 213 or complements thereof or fragments of either. In another aspect, a preferred marker nucleic acid molecule shares between 80% and 100% or 90% and 100% sequence identity with the nucleic acid sequence set forth in SEQ ID NO: 143 through SEQ ID NO: 213 or complement thereof or fragments of either. In a further aspect, a preferred marker nucleic acid molecule shares between 95% and 100% sequence identity with the sequence set forth in SEQ ID NO: 143 through SEQ ID NO: 213 or complement thereof or fragments of either. In an aspect, a preferred marker nucleic acid molecule shares between 98% and 100% sequence identity with the nucleic acid sequence set forth in SEQ ID NO: 143 through SEQ ID NO: 213 or complement thereof or fragments of either.

**[0056]** The percent identity is preferably determined using the "Best Fit" or "Gap" program of the Sequence Analysis Software Package™ (Version 10; Genetics Computer Group, Inc., University of Wisconsin Biotechnology Center, Madison, WI). "Gap" utilizes the algorithm of Needleman and Wunsch to find the alignment of two sequences that maximizes the number of matches and minimizes the number of gaps. "BestFit" performs an optimal alignment of the best segment of similarity between two sequences and inserts gaps to maximize the number of matches using the local homology algorithm of Smith and Waterman. The percent identity calculations may also be performed using the Megalign program of the LASERGENE bioinformatics computing suite (default parameters, DNASTAR Inc., Madison, Wisconsin). The percent identity is most preferably determined using the "Best Fit" program using default parameters.

**[0057]** The present invention further discloses one or more single nucleotide polymorphism (SNP) markers. The detection of polymorphic sites in a sample of DNA, RNA, or cDNA may be facilitated through the use of nucleic acid amplification methods. Such methods include those that specifically increase the concentration of polynucleotides that span the polymorphic site, or include that site and sequences located either distal or proximal to it. Such amplified molecules can be readily detected by gel electrophoresis or other means.

**[0058]** A method of achieving such amplification employs the polymerase chain reaction (PCR) (Mullis et al. 1986 Cold Spring Harbor Symp. Quant. Biol. 51:263-273; European Patent Nos. 50,424; 84,796; 258,017; 237,362; 201,184; U.S. Patent Nos. 4,683,202; 4,582,788; and 4,683,194), using primer pairs that are capable of hybridizing to the proximal sequences that define a polymorphism in its double-stranded form.

**[0059]** Alleles that associate with plant maturity can be determined based on linkage analysis of plants and nucleic acid molecules. A number of molecular genetic maps of *Glycine* have been reported (Mansur et al., Crop Sci. 36: 1327-1336 (1996); Shoemaker et al., Genetics 144: 329-338 (1996); Shoemaker et al., Crop Science 32: 1091-1098 (1992), Shoemaker et al., Crop Science 35: 436-446 (1995); Tinley and Rafalski, J. Cell Biochem. Suppl. 14E: 291 (1990); Cregan et al., Crop Science 39:1464-1490 (1999)). *Glycine max*, *Glycine soja* and *Glycine max* x. *Glycine soja* share linkage groups (Shoemaker et al., Genetics 144: 329-338 (1996)). A linkage group (LG) is a set of genes that tend to be inherited together from generation to generation. As used herein, reference to the linkage groups (LG), D1b; C2; O; L; and I and of *Glycine max* refers to the linkage group that corresponds to linkage groups, D1b, C2, O, L; and I from the genetic map of *Glycine max* (Mansur et al., Crop Science. 36: 1327-1336 (1996)); Cregan et al., Crop Science 39: 1464-1490 (1999), and Soybase, Agricultural Research Service, United States Department of Agriculture.

**[0060]** Genome-wide surveys revealed SNP markers associated with maturity genomic region 1 are located on linkage group (LG) C2, maturity genomic region 2 is located on LG O, maturity genomic region 3 is located on LG L, maturity genomic region 4 is located on LG I, maturity genomic region 5 is located on LG L, maturity genomic region 6 is located on LG D1b+W, maturity genomic region 7 is located on LG G, and maturity genomic region 8 is located on LG M.

**[0061]** In an aspect, the present invention can be used to identify additional markers associated with maturity genomic regions 1-8. Disclosed is a maturity marker within 1 cM, 5 cM, 10 cM, 15 cM, or 30 cM of SEQ ID NO: 143 - 213. Similarly, one or more markers mapped within 1, 5, 10, 20 and 30 cM or less from the marker molecules can be used for the selection or introgression of the region associated with maturity and/or plant growth habit. The present invention includes a maturity marker that is linked with SEQ ID NO: 143 - 213 and delays maturity. The present invention includes a substantially purified nucleic acid molecule comprising a maturity marker within 5 kilobases, 10 kilobases, 20 kilobases,

30 kilobases, 100 kilobases, 500 kilobases, 1,000 kilobases, 10,000 kilobases, 25,000 kilobases, or 50,000 kilobases of a marker selected from the group consisting of SEQ ID NO: 143 - 213. The present invention includes a maturity marker within 5 kilobases, 10 kilobases, 20 kilobases, 30 kilobases, 100 kilobases, 500 kilobases, 1,000 kilobases, 10,000 kilobases, 25,000 kilobases, or 50,000 kilobases of any of SEQ ID NO: 143 - 213 that cosegregates with any of SEQ ID NO: 143 - 213. Similarly, one or more markers mapped within 5 kilobases, 10 kilobases, 20 kilobases, 30 kilobases, 100 kilobases, 500 kilobases, 1,000 kilobases, 10,000 kilobases, 25,000 kilobases, or 50,000 kilobases or less from the marker molecules can be used for the selection or introgression of the region associated with maturity and/or plant growth habit.

**[0062]** A maturity genomic region is a physical region of a plant chromosome that has been associated with determining a plant's maturity date. A plant is considered mature when 95% of its pods have reached their mature color. In one aspect, the maturity date of a plant is the number of days after August 31<sup>st</sup> in the northern hemisphere. Alleles of maturity genomic regions 1-8 can influence the maturity date of a plant.

**[0063]** In one aspect, the maturity date of a plant can determine the maturity group of a plant. Herein, relative maturity refers to a soybean plant maturity group subdividing a maturity group into tenths, *for example* III.5. Relative maturity provides a more exact description of plant maturity. The number following the decimal point refers to the relative earliness or lateness with a maturity group, *for example*, IV.2 is an early group IV variety and IV.9 is a late group IV.

**[0064]** In another aspect, maturity group can be determined by reference to a commercialized strain for a maturity group. For example, a commercialized strain with a known maturity group is grown in an experiment with a new soybean line and the relative maturity of the new soybean line is ascertained by counting the number of days after Aug 31<sup>st</sup> and comparing to the commercialized strain. Maturity group refers to an industry division of groups of varieties based on a range in latitudes which the plant is best adapted and most productive. Soybean varieties are classified into 13 recognized maturity groups with the designations ranging from maturity groups 000, 00, 0, and I through X, where 000 represents the earliest maturing variety and X represents the latest maturing variety. The maturity groups have corresponding maturity belts.

**[0065]** Soybean plants in maturity groups 000 to IV have an indeterminate plant habit, while soybean plants in maturity groups V through X have a determinate plant habit. Early maturity varieties (000 to III) are adapted to northern latitudes with longer day lengths with the maturity designation increasing in southern latitudes with shorter day lengths.

**[0066]** An increase in maturity can correlate with an increase in yield or other traits such as oil concentration. The correlation of plant maturity and other traits confounds the evaluation of potential markers and candidate genes associated with other traits such as yield. Identification of genomic regions associated with plant maturity, but not with another trait, can allow breeders to genetically fix plant maturity within a soybean plant and separately elucidate other traits, such as those associated with yield.

**[0067]** Disclosed is a method of establishing where a soybean plant or soybean seed should be grown by determining the allelic combination of a soybean plant or soybean seed by obtaining DNA from a soybean plant or soybean seed; determining if alleles at a locus within maturity genomic region 1 are homozygous or heterozygous; determining if alleles at a locus within maturity genomic region 2 are homozygous or heterozygous; determining if alleles at a locus within maturity genomic region 3 are homozygous or heterozygous; determining the allelic combination of the alleles within maturity genomic regions 1, 2, and 3; and assigning a maturity group value to the soybean plant or soybean seed. In a preferred aspect, determining if alleles at a locus are homozygous or heterozygous includes detecting a polymorphism with a nucleic acid molecule having a sequence of any of SEQ ID NOs: 143-174, or complements thereof.

**[0068]** Further disclosed is a method of establishing where a soybean plant or soybean seed should be grown by determining the allelic combination of a soybean plant or soybean seed by obtaining DNA from a soybean plant or soybean seed; determining if alleles at a locus within maturity genomic region 1 are homozygous or heterozygous; determining if alleles at a locus within maturity genomic region 2 are homozygous or heterozygous; determining if alleles at a locus within maturity genomic region 3 are homozygous or heterozygous; determining if alleles at a locus within maturity genomic region 4 are homozygous or heterozygous; determining the allelic combination of the alleles within maturity genomic regions 1, 2, 3 and 4; and assigning a maturity group value to the soybean plant or soybean seed.

**[0069]** Further disclosed is a method of providing information about the maturity of a soybean plant or soybean seed by obtaining DNA from the soybean seed or soybean plant and determining the allelic profile at a locus of genomic region 4.

**[0070]** The present invention also includes a method of establishing where a soybean plant or soybean seed should be grown by determining the allelic combination of a soybean plant or soybean seed by obtaining DNA from a soybean plant or soybean seed; determining if an allele within maturity genomic region 1 is homozygous or heterozygous; determining if an allele within maturity genomic region 2 is homozygous or heterozygous; determining if an allele within maturity genomic region 3 is homozygous or heterozygous; and determining the allelic combination of the alleles within maturity genomic regions 1, 2, and 3.

**[0071]** In a preferred aspect, the soybean plant or soybean seed is homozygous for the alleles within maturity genomics regions 1, 2, and 3. In a preferred aspect, the homozygous alleles are either dominant or recessive. In another aspect, the soybean plant or soybean seed is homozygous for the alleles within maturity genomics regions 1 and 2. In a preferred

aspect, the homozygous alleles are either dominant or recessive. In another aspect, the soybean plant or soybean seed is homozygous for the alleles within maturity genomic regions 2 and 3. In a preferred aspect, the homozygous alleles are either dominant or recessive. In another aspect, the soybean plant or soybean seed is heterozygous for the alleles within maturity genomic regions 1, 2, and 3. In another aspect, the soybean plant or soybean seed is heterozygous for the alleles within maturity genomic regions 1 and 2. In another aspect, the soybean plant or soybean seed is heterozygous for the alleles within maturity genomic regions 2 and 3. In a preferred aspect, the allelic combination is allelic combination 10, allelic combination 11, allelic combination 12, allelic combination 13, allelic combination 14, allelic combination 15, allelic combination 16, allelic combination 17, allelic combination 18, and allelic combination 19.

**[0072]** An aspect of the method of the present invention the method is suitable for establishing where a soybean plant or soybean seed should be grown by determining the allelic combination of a soybean plant by obtaining DNA from a soybean plant or soybean seed; determining if an allele within maturity genomic region 1 is homozygous or heterozygous; determining if an allele within maturity genomic region 2 is homozygous or heterozygous; determining the allelic combination of the alleles within maturity genomic regions 1 and 2; and assigning a maturity growth value to the soybean plant or soybean seed. In a preferred aspect, determining whether an allele is homozygous or heterozygous includes detecting a polymorphism from any of SEQ ID NOs: 143-161. In a preferred aspect, the allelic combination is allelic combination 1, allelic combination 2, allelic combination 3, allelic combination 4, allelic combination 5, allelic combination 6, allelic combination 7, allelic combination 8, and allelic combination 9. In a preferred aspect, the soybean plant or soybean seed is obtained from a cross of an early maturity group parent soybean plant and a mid-maturity parent soybean plant. In a preferred aspect, the early maturity group parent soybean plant is between 00.0 - 1.0 and the mid-maturity parent soybean plant is between III.0-IV.9

**[0073]** An aspect includes a method to determine if a soybean plant has a maturity group of 0.0-III.9 by determining if an allele within maturity genomic region 1 is homozygous or heterozygous; determining if an allele within maturity genomic region 2 is homozygous or heterozygous; and assigning a maturity group value for the soybean plant between 0.0-III.9. In a preferred aspect, maturity in the soybean plant is reached at least 5 days before a soybean plant that is homozygous dominant within maturity genomic region 1, homozygous dominant within maturity genomic region 2 and is grown under the same environmental conditions.

**[0074]** Another aspect includes a method to determine if the maturity of a soybean plant is in a 00.0 - III.0 maturity group by determining if an allele within maturity genomic region 1 is homozygous or heterozygous; determining if an allele within maturity genomic region 2 is homozygous or heterozygous; and assigning a maturity group value for the soybean plant between 00.0 - III.0. In a preferred aspect, a selected soybean seed is homozygous recessive at maturity genomic region 1 and homozygous recessive at maturity genomic region 2 and has a maturity group between 0.5 - II.0. In a preferred aspect, a soybean seed is selected that is homozygous recessive at maturity genomic region 1 and heterozygous dominant at maturity genomic region 2 and has a maturity group between 1.5 - II.9.

**[0075]** Also disclosed is a method where the maturity group of a progeny plant is predicted by whether an allele in maturity genomic region 1 is homozygous dominant, homozygous recessive, or heterozygous and whether an allele in maturity genomic region 2 is homozygous dominant, homozygous recessive, or heterozygous. In an aspect, if the maturity group of a plant is between 0 and II, the maturity group can be identified by determining the allelic combination of maturity genomic regions 1 and 2 in a plant or seed. See, for example, Table 9.

**[0076]** In an alternate aspect, if the maturity group of a plant is between III and V, the maturity group can be identified by determining the allelic combination of maturity genomic regions 1, 2 and 3 in a plant or seed. See, for example, Table 9. In an aspect, if the maturity group of a plant is between IV and V, the maturity group can be identified by determining the allelic combination of maturity genomic regions 1, 2 and 3 in a plant or seed. See, for example, Table 9.

**[0077]** In another aspect, the maturity group of the parent plants is known. In an aspect, the maturity groups of the parent plants are different by more than 10 days, between 10 days - 20 days, between 10 days - 30 days, more than 2 maturity groups, less than 2 maturity groups, between maturity groups 000 and VI. In an aspect, the maturity group of a progeny plant resulting from a cross with at least one parent having a maturity group of 0-II is identified by determining the allelic combination of maturity genomic regions 1 and 2. In another aspect, the maturity group of a progeny plant resulting from a cross with parent plants having a maturity group of III, IV, V, or III-V is identified by determining the allelic combination of maturity genomic regions 1, 2 and 3.

**[0078]** In an aspect, more dominant alleles at a locus in a maturity group region correlate with a delay in maturity. In another aspect, an increase in the number of dominant alleles correlates with a delay in maturity.

**[0079]** In an aspect, parent plants with a difference in maturity group greater than 1.5, 2, 2.5, 3, 3.5 are crossed and their maturity group identified by determining the allelic combination. In an aspect, parent plants with a difference in maturity group between 1 and 3, between 1 and 4, between 2 and 3, between 2 and 5, between 2 and 6, between 2 and 7 are crossed and their maturity group identified by determining the allelic combination of the progeny. In an aspect, parent plants with a difference in maturity group greater than 1.5, 2, 2.5, 3, 3.5 are crossed and their maturity group identified by determining the allelic combination.

**[0080]** In an aspect, a progeny plant has a maturity group earlier than one parent by 5, 10, or 15 days. In another

aspect a progeny plant has a maturity group later than one parent plant by 5, 10, or 15 days. In an aspect, a progeny plant has a maturity group earlier than both parents by 5, 10, or 15 days. In another aspect, a progeny plant has a maturity group later than both parent plants by 5, 10, or 15 days.

**[0081]** In an aspect, an early parent of maturity group 0.1 is crossed with a later maturity parent plant that is a 1.9, and the progeny plants with allelic combination 1 are 0.1 - 0.5 maturity. In another aspect, an early parent with maturity of 0.9 is crossed with a plant having 3.5 maturity, and the plants having allelic combination 1 are maturity group 1.0 - 1.5.

**[0082]** In an aspect, the maturity group of a progeny seed is determined from a cross between a very early maturity parent plant with a later maturity parent plant. In an aspect, the very early maturity parent plant is a maturity group 00.0 - 0.9 and the later maturity parent plant is a maturity group III.5 - IV.5. In an aspect, the very early maturity parent plant is a maturity group 00 and the later maturity parent plant is a maturity group III or IV. In an aspect, DNA can be obtained from plants or plant parts such as seeds in the F<sub>1</sub>, F<sub>2</sub>, F<sub>3</sub>, F<sub>4</sub> or later populations. In an aspect, one or more plants or plant parts are genotyped for alleles in genomic regions 1 and 2. In an aspect, the alleles are determined using the SNP markers NS0128378 (genomic maturity region 1) and NS0118907 (genomic maturity region 2).

**[0083]** In an aspect, the plants are phenotyped for maturity by counting the number of days after August 31<sup>st</sup> until a plant matures. In an aspect, a plant is considered mature when 95% of the pods are brown. In an aspect, when alleles from markers associated with maturity genomic regions 1 and 2 are homozygous recessive, the progeny plant will reach maturity 15, 14, 12, 11, 10, 9, or 8 days sooner than the maturity group if the alleles from markers associated with maturity genomic regions 1 and 2 are homozygous dominant. In an aspect, if an allele from a marker associated with maturity genomic region 1 is homozygous dominant and an allele from a marker associated with maturity genomic region 2 is heterozygous, then the progeny plant will reach maturity between 1 day, 1-2 days, 2-3 days, 2-4 days, or 3-5 days earlier than if the alleles from markers associated with maturity genomic regions 1 and 2 are homozygous dominant.

**[0084]** In another aspect, multiple seeds can be selected or bulked. Multiple seeds may include greater than or equal to 2, 3, 4, 5, 6, 10, 50, 100, 500, 1000, 5,000, 10,000 or more seeds. One or multiple seeds can be distributed to a geographic region suitable for growth of one or multiple plants. In this aspect, seeds selected can be distributed or shipped to an appropriate region.

**[0085]** The present invention also provides multiple soybean seeds in which greater than 50%, 60%, 70%, 80%, 90%, 95%, or 99% of the seeds will grow into plants where the variation in maturity group is within one maturity group, not more than 2 groups or 20 days after August 31<sup>st</sup>, not more than 1 group or 10 days after August 31<sup>st</sup>, not more than 0.9 group or nine days after August 31<sup>st</sup>, not more than 5 days after August 31<sup>st</sup> or 0.5 group, or with a maturity group between 0.0 - II.0, 000.0 - III.9. The multiple soybean seeds can grow into soybean plants having indeterminate soybean plant habit or having determinate soybean plant habit. One aspect includes a method to select a soybean seed based on indeterminate or determinate growth habit comprising determining if maturity genomic region 3 is homozygous or heterozygous. In one aspect, 85% of the multiple soybean seeds can reach maturity within 10 days, 5 days, 3 days of each other. In another aspect, 95% of the multiple soybean seeds can reach maturity within 10 days, 5 days, 3 days of each other.

**[0086]** In another aspect the method of the present invention is suitable to isolate indeterminate-early maturity soybean seeds by obtaining DNA from the soybean seed using a non-destructive method; determining if an allele within maturity genomic region 1 is homozygous or heterozygous; and determining if an allele within maturity genomic region 2 is homozygous or heterozygous.

**[0087]** Such multiple seeds may be in a container. The container of soybean seeds can contain any number, weight, or volume of seeds. For example, a container can contain at least, or greater than, about 10, 25, 50, 100, 200, 300, 400, 500, 600, 700, 80, 90, 1000, 1500, 2000, 2500, 3000, 3500, 4000, 5000, 7500, or 10,000 or more seeds. In another aspect, a container can contain about, or greater than about, 1 gram, 5 grams, 10 grams, 15 grams, 20 grams, 25 grams, 50 grams, 100 grams, 250 grams, 500 grams, or 1000 grams of seeds. Alternatively, the container can contain at least, or greater than, about 0 ounces, 1 ounce, 5 ounces, 10 ounces, 1 pound, 2 pounds, 3 pounds, 4 pounds, 5 pounds, 10 pounds, 15 pounds, 20 pounds, 25 pounds, 30 pounds, 40 pounds, 50 pounds, 60 pounds, 70 pounds, 80 pounds, 100 pounds, 200 pounds, 300 pounds, 500 pounds, or 1000 pounds or more seeds.

**[0088]** Containers of soybean seeds can be any container available in the art. For example, a container can be a box, a bag, a can, a packet, a pouch, a tape roll, a pail, or a tube.

**[0089]** In another aspect, the seeds contained in the containers of soybean seeds can be treated or untreated soybean seeds. In one aspect, the seeds can be treated to improve germination, for example, by priming the seeds, or by disinfection to protect against seed-born pathogens. In another aspect, seeds can be coated with any available coating to improve, for example, plantability, seed emergence, and protection against seed-born pathogens. Seed coating can be any form of seed coating including, but not limited to, pelleting, film coating, and encrustments. One aspect includes a method of distributing a soybean plant based on maturity group by obtaining DNA from a soybean plant; determining if an allele within maturity genomic region 1 is homozygous or heterozygous; determining if an allele within maturity genomic region 2 is homozygous or heterozygous; determining if an allele within maturity genomic region 3 is homozygous or heterozygous; and assigning a maturity growth value to the soybean plant; and shipping the soybean plant to a

preferred geographic region.

**[0090]** A plant may also comprise a gene that confers resistance to insect, pest, viral or bacterial attack. Such a gene may be a transgene. For example, a gene conferring resistance to a pest, such as soybean cyst nematode was described in US Patent Nos. 7,154,021.

**[0091]** Transgenes may also be used to alter protein metabolism. For example, U.S. Patent No. 5,545,545 describes lysine-insensitive maize dihydronic acid synthase (DHPS), which is substantially resistant to concentrations of L-lysine which otherwise inhibit the activity of native DHPS. Similarly, EP 0640141 describes sequences encoding lysine-insensitive aspartokinase (AK) capable of causing a higher than normal production of threonine, as well as a subfragment encoding antisense lysine ketoglutarate reductase for increasing lysine.

**[0092]** In another aspect, a transgene may be employed that alters plant carbohydrate metabolism. For example, fructokinase genes are known for use in metabolic engineering of fructokinase gene expression in transgenic plants and their fruit (see U.S. Patent No. 6,031,154). A further example of transgenes that may be used are genes that alter grain yield. For example, U.S. Patent No. 6,486,383 describes modification of starch content in plants with subunit proteins of adenosine diphosphoglucose pyrophosphorylase ("ADPG PPase"). In EP0797673 transgenic plants are discussed in which the introduction and expression of particular DNA molecules results in the formation of easily mobilized phosphate pools outside the vacuole and an enhanced biomass production and/or altered flowering behavior. Still further known are genes for altering plant maturity. U.S. Patent No. 6,774,284 describes DNA encoding a plant lipase and methods of use thereof for controlling senescence in plants. U.S. Patent No. 6,140,085 discusses FCA genes for altering flowering characteristics, particularly timing of flowering. U.S. Patent No. 5,637,785 discusses genetically modified plants having modulated flower development such as having early floral meristem development and comprising a structural gene encoding the LEAFY protein in its genome.

**[0093]** In another aspect, the present invention provides methods and compositions for the preferred deployment of conventional and transgenic traits related to fatty acid synthesis and oil content. Using present invention, breeders can tailor trait integration to geographies for preferred trait expression, whether the trait is conventional (for example, a mutation) or transgenic. For example, a transgene may be employed that alters plant oil biosynthesis and oil composition. In particular, linoleic acid (LA) (18:2, Δ9, 12) is produced from oleic acid (18:1, Δ9) by a Δ12-desaturase (encoded by FAD2) while alpha linolenic acid (ALA) (18:3, Δ9, 12, 15) is produced from LA by a Δ15-desaturase (encoded by FAD3). Moreover, stearidonic acid (SDA) (18:4, Δ6, 9, 12, 15) and gamma linolenic acid (GLA) (18:3, Δ6, 9, 12) are polyunsaturated fatty acids (PUFAs) produced from LA and ALA by a Δ6-desaturase. Various genes encoding desaturases have been described. For example, U.S. Pat. No. 5,952,544 describes nucleic acid fragments isolated and cloned from *Brassica napus* that encode fatty acid desaturase enzymes. Expression of the *B. napus* Δ15-desaturase of the '544 patent resulted in accumulation of ALA. U.S. Pat. Publication 2006/0156435 describes the expression of fungal Δ15-desaturases to increase omega-3 fatty acid profiles in plants. WO05/021761 discusses genetically engineered plants which produce both SDA and GLA as a result of expressing a Δ6-desaturase and a Δ15-desaturase. Long chain PUFAs such as EPA and DHA can be produced in plants as disclosed in US Pat. Publication 2004/0172682.

**[0094]** Inhibition of the endogenous soy FAD2 gene through use of transgenes that inhibit the expression of FAD2 has been shown to confer a desirable mid-oleic acid (18:1) phenotype (i.e. soybean seed comprising about 50% and 75% oleic acid by weight). Transgenes and transgenic plants that provide for inhibition of the endogenous FAD2 gene expression and a mid-oleic phenotype are disclosed in U.S. Patent 7,067,722. In contrast, wild type soybean plants that lack FAD2 inhibiting transgenes typically produce seed with oleic acid compositions of less than 20%. Inhibition of the endogenous FAD3 gene gene through use of transgenes that inhibit the expression of FAD3 has been shown to confer a desirable linolenic acid (18:3) phenotype. A "FATB" or "palmitoyl-ACP thioesterase" gene encodes an enzyme (FATB) capable of catalyzing the hydrolytic cleavage of the carbon-sulfur thioester bond in the panthothene prosthetic group of palmitoyl-ACP as its preferred reaction. Hydrolysis of other fatty acid-ACP thioesters may also be catalyzed by this enzyme. Representative FATB-1 sequences include, without limitation, those set forth in U.S Pat. Publication 2004/0006792 and U.S. Patent Nos. 5,955,329; 5,723,761; 5,955,650; and 6,331,664. When the amount of FATB is decreased in a plant cell, a decreased amount of saturated fatty acids such as palmitate and stearate may be provided. Thus, a decrease in expression of FATB may result in an increased proportion of unsaturated fatty acids such as oleic acid (18:1). The simultaneous suppression of FAD2, FAD3, and FATB expression thereby results in driving the FAS pathway toward an overall increase in mono-unsaturated fatty acids of 18-carbon length, such as oleic acid (C18:1). See U.S. Patent No. 5,955,650.

**[0095]** In an aspect, the present invention provides methods and compositions for the preferred deployment of conventional and transgenic traits related to fatty acid synthesis and oil content. Soybean seed oil levels are highly impacted by environment. Oil concentration increases with decreasing latitude, therefore, soybeans in maturity groups 00-I generally have lower oil levels than later maturing soybeans (Yaklich et al. 2002. Crop Sci 42:1504-1515). The decrease in oil concentrations is attributed to lower temperatures and shorter growing season (Piper and Boote 1999 J. Am. Oil Chem. Soc. 76:1233-124). In addition, soybeans cultivated under drought stress tend to produce seeds with decreased protein and increased oil (Specht et al. 2001 Crop Sci 41:493-509). Using present invention, breeders can tailor trait

integration to geographies for preferred trait expression, whether the trait is conventional (for example, a mutation) or transgenic.

**[0096]** Genes for altering plant morphological characteristics are also known and may be used in accordance with the invention. U.S. Patent No. 6,184,440 discusses genetically engineered plants which display altered structure or morphology as a result of expressing a cell wall modulation transgene. Examples of cell wall modulation transgenes include a cellulose binding domain, a cellulose binding protein, or a cell wall modifying protein or enzyme such as endoxyloglucan transferase, xyloglucan endo-transglycosylase, an expansin, cellulose synthase, or a novel isolated endo-1,4- $\beta$ -glucanase.

**[0097]** Methods for introduction of a transgene, for instance to soybean, are well known in the art and include biological and physical plant transformation protocols. See, for example, Miki *et al.* (1990), Clemente *et al.* (Clemente et al., Crop Sci., 40:797-803, 2000), and U.S. Patent 7,002,058. A further aspect of the invention relates to tissue cultures of a soybean variety of the invention. Exemplary types of tissue cultures are protoplasts, calli and plant cells that are intact in plants or parts of plants. Plant parts include, but not limited to, embryos, pollen, flowers, leaves, roots, root tips, anthers, vascular tissue, pod, stem, seed, or a portion thereof, or a cell isolated from the plant. In an aspect, the tissue culture comprises plant parts such as embryos, protoplasts, meristematic cells, pollen, leaves or anthers. In these ways, plants or parts thereof can be grown in culture and regenerated. Exemplary procedures for preparing tissue cultures of regenerable soybean cells and regenerating soybean plants therefrom, are disclosed in U.S. Patent Nos. 4,992,375; 5,015,580; 5,024,944, and 5,416,011. An important ability of a tissue culture is the capability to regenerate fertile plants. For transformation to be efficient and successful, DNA must be introduced into cells that give rise to plants or germ-line tissue.

In particular, methods for the regeneration of *Glycine max* plants from various tissue types and methods for the tissue culture of *Glycine max* are known in the art (See, for example, Widholm *et al.*, In Vitro Selection and Culture-induced Variation in Soybean, In Soybean: Genetics, Molecular Biology and Biotechnology, Eds. Verma and Shoemaker, CAB International, Wallingford, Oxon, England (1996). Regeneration techniques for plants such as *Glycine max* can use as the starting material a variety of tissue or cell types. With *Glycine max* in particular, regeneration processes have been developed that begin with certain differentiated tissue types such as meristems, Cartha *et al.*, Can. J. Bot. 59:1671-1679 (1981), hypocotyl sections, Cameya *et al.*, Plant Science Letters 21: 289-294 (1981), and stem node segments, Saka *et al.*, Plant Science Letters, 19: 193-201 (1980); Cheng *et al.*, Plant Science Letters, 19: 91-99 (1980). Regeneration of whole sexually mature *Glycine max* plants from somatic embryos generated from explants of immature *Glycine max* embryos has been reported (Ranch *et al.*, In Vitro Cellular & Developmental Biology 21: 653-658 (1985)). Regeneration of mature *Glycine max* plants from tissue culture by organogenesis and embryogenesis has also been reported (Barwale *et al.*, Planta 167: 473-481 (1986); Wright *et al.*, Plant Cell Reports 5: 150-154 (1986)). Once a transgene is introduced into a variety it may readily be transferred by crossing. By using backcrossing, essentially all of the desired morphological and physiological characteristics of a variety are recovered in addition to the locus transferred into the variety via the backcrossing technique. Backcrossing methods can be used with the present invention to improve or introduce a characteristic into a plant (Poehlman and Sleper, In: Breeding Field Crops, Iowa State University Press, Ames, 1995; Fehr, Principles of Cultivar Development Vol. 1, pp. 2-3 (1987)). Disclosed is a method of soybean plant breeding by crossing at least two different parent soybean plants, where the parent soybean plants differ in plant maturity by over 10 days, 10 days - 20 days, 10 days - 30 days; obtaining a progeny seed from the cross; genotyping a progeny seed of the cross with a genetic marker; and selecting a soybean seed possessing a genotype for preferred maturity. Also disclosed is a method of soybean plant breeding by assaying a soybean plant for the presence of a marker sequences selected from SEQ ID NO: 143 through SEQ ID NO: 213; and associating the soybean plant with a maturity group. The present invention also includes a method of soybean plant breeding comprising crossing a parent soybean plant having a desired trait with a second parent soybean plant, where the parent soybean plants differ in soybean plant maturity by over 10 days, 10 days - 20 days, 10 days - 30 days, by crossing a parent soybean plant comprising a desired trait with a second parent soybean plant; obtaining progeny soybean seed from the cross; screening a progeny soybean seed for the trait; screening a progeny soybean seed for a desired maturity group using a marker selected from the group consisting of SEQ ID NO: 143 through SEQ ID NO: 213 to determine the desired geographical growing region; and selecting a progeny soybean seed containing the desired trait and desired soybean plant maturity.

**[0098]** The method of soybean plant breeding includes crossing at least two different parent soybean plants; obtaining a progeny soybean plant from the cross; nondestructive genotyping a progeny soybean plant or soybean seed of the cross with a genetic marker characterizing a maturity genomic region; and selecting a soybean plant possessing a genotype for a desired maturity group. In a preferred aspect, the maturity phenotype of the progeny soybean plant or soybean seed is unknown. In another preferred aspect, the progeny is grown under conditions that are unsuitable for determining maturity of the soybean plant. In another preferred aspect, the parent soybean plants differ in soybean plant maturity by over 5 days, over 10 days, 10 days - 20 days, 10 days - 30 days. herein a maturity phenotype of at least one of the two different parent soybean plants is unknown. In a preferred aspect, the maturity phenotype of both of the two different parent soybean plants is unknown. In a preferred aspect, the progeny soybean plant is not photoperiod sensitive. In another preferred aspect, at least one parent soybean plant is not photoperiod sensitive. In a preferred

aspect, both parent soybean plants are not photoperiod sensitive. In a preferred aspect, the maturity genomic region is characterized by a dominant allele identified in Table 6. In a preferred aspect, the maturity genomic region is characterized by a recessive allele identified in Table 6.

**[0099]** In an aspect, at least one or both parent soybean plant are an elite variety. In an aspect, a progeny soybean plant is an exotic soybean plant or one or both parent soybean plants are exotic soybean plants.

**[0100]** An aspect includes a method of selecting a soybean plant for germplasm improvement by determining a maturity group by crossing at least two different parent soybean plants; obtaining a progeny soybean plant from the cross; nondestructive genotyping a progeny soybean plant or soybean seed of the cross with a genetic marker characterizing a maturity genomic region; and selecting a soybean plant possessing a genotype for a desired maturity group; and incorporating the selected soybean plant into a use selected from the group consisting of using the soybean plant for breeding, advancement of the soybean plant through self-fertilization, trait integration, use of soybean plant or parts thereof for transformation, and use of soybean plants or parts thereof for mutagenesis.

**[0101]** Another aspect of the method of the present invention includes a method of co-selecting a soybean plant for expression of a non-maturity phenotypic trait and a maturity trait by crossing at least two different parent soybean plants; obtaining a progeny soybean plant from the cross; nondestructive genotyping a progeny soybean plant or soybean seed of the cross with a genetic marker characterizing a maturity genomic region; and selecting a soybean plant possessing a genotype for a desired maturity group; and to determine the desired geography for the progeny soybean plant growth, and a method for determining the non-maturity phenotype.

**[0102]** In a preferred aspect, the method for detecting the non-maturity phenotype is a genotypic or phenotypic method. In a preferred aspect, the non-maturity phenotypic trait is any of herbicide tolerance, increased yield, insect control, fungal disease resistance, virus resistance, nematode resistance, bacterial disease resistance, mycoplasma disease resistance, altered oils production, high oil production, high protein production, germination and seedling growth control, enhanced animal and human nutrition, low raffinose, environmental stress resistant, increased digestibility, industrial enzymes, pharmaceutical proteins, peptides and small molecules, improved processing traits, improved flavor, nitrogen fixation, hybrid soybean seed production, reduced allergenicity, biopolymers, and biofuels.

**[0103]** In another preferred aspect, a phenotypic trait is any of altered protein and oil composition, altered levels of a molecule selected from the group consisting of protein, oil, linolenic acid, stearic acid, palmitic acid, oleic acid, linoleic acid, stearidonic acid, alpha-linolenic acid, gamma linolenic acid, docosahexaenoic acid, eicosapentaenoic acid, docosapentaenoic acid, and combinations thereof.

**[0104]** In one aspect, plants can be used in activities related to germplasm improvement, non-limiting examples of which include using the plant for breeding, advancement of the plant through self-fertilization, trait integration, use of plant or parts thereof for transformation, and use of plants or parts thereof for mutagenesis. Non-limiting examples of breeding decisions include progeny selection, parent selection, and recurrent selection for at least one haplotype. In another aspect, breeding decisions relating to development of plants for commercial release comprise advancing plants for testing, advancing plants for purity, purification of sublines during development, variety development, and hybrid development. In yet other aspects, breeding decisions and germplasm improvement activities comprise transgenic event selection, making breeding crosses, testing and advancing a plant through self-fertilization, using plants or parts thereof for transformation, using plants or parts thereof for candidates for expression constructs, and using plants or parts thereof for mutagenesis. The choice of breeding method depends on the mode of plant reproduction, the heritability of the trait (s) being improved, and the type of cultivar used commercially (e.g., F<sub>1</sub> hybrid cultivar, pureline cultivar).

**[0105]** Descriptions of breeding methods that are commonly used for soybeans can be found in one of several reference books (e.g. Fehr, Principles of Cultivar Development Vol. 1, pp. 2-3 (1987)). In one aspect disclosed is a method of soybean plant breeding by assaying a soybean plant for the presence of a marker sequences selected from the group consisting of SEQ ID NO: 143 through SEQ ID NO: 213; and associating the soybean plant with a maturity group.

**[0106]** In another aspect disclosed is a method of soybean plant breeding comprising crossing a parent soybean plant having a desired trait with a second parent soybean plant, wherein the parent soybean plants differ in soybean plant maturity by over 5 days, over 10 days, 10 days - 20 days, or 10 days - 30 days, by crossing a parent soybean plant comprising a desired trait with a second parent soybean plant; obtaining progeny soybean seed from the cross; screening a progeny soybean seed for the trait; screening a progeny soybean seed for a desired maturity group using a marker selected from the group consisting of SEQ ID NO: 143 through SEQ ID NO: 213 to determine the desired geographical growing region; and selecting a progeny soybean seed containing the desired trait and desired soybean plant maturity. In a preferred aspect, the desired trait is transgenic.

**[0107]** An aspect includes a method of soybean plant breeding by crossing at least two different parent soybean plants, wherein the parent soybean plants differ in soybean plant maturity by over 5 days, over 10 days, 10 days - 20 days, or 10 days - 30 days; obtaining a progeny soybean seed from the cross; genotyping a progeny soybean seed of the cross with a genetic marker; and selecting a soybean seed possessing a genotype for preferred maturity.

**[0108]** Another aspect includes a method of screening soybean seeds based on soybean plant maturity group by obtaining DNA from a soybean seed; determining if an allele within maturity genomic region 1 is homozygous or heter-

ozygous; determining if an allele within maturity genomic region 2 is homozygous or heterozygous; determining if an allele within maturity genomic region 3 is homozygous or heterozygous; and assigning a maturity growth value to the soybean seed.

[0109] One aspect is a method of introgressing an allele into a soybean plant by crossing at least two different parent soybean plants; obtaining a progeny soybean plant from the cross; screening the progeny soybean plant of the cross for the allele; obtaining DNA from a soybean seed of the progeny soybean plant using a non-destructive method; and selecting a soybean seed, wherein the soybean seed comprises the allele and a nucleic acid sequence selected from the group consisting of SEQ ID NOs: 143-213. In a preferred aspect, the selected soybean seed further has a second sequence selected from the group consisting of SEQ ID NOs: 143-213. In another preferred aspect, the allele is selected from any or both of SCN resistance and root rot resistance. Another aspect includes a method of introducing a desired trait into a soybean plant by crossing at least two different parent soybean plants, wherein at least one parent soybean plant has a desired trait; obtaining a progeny soybean seed from the cross; obtaining DNA from a soybean seed of the progeny soybean plant using a non-destructive method; assaying the progeny soybean seed of the cross for evidence of the desired trait; and selecting the soybean seed with the desired trait and a desired maturity group. In a preferred aspect, the desired trait is transgenic.

[0110] A further aspect includes a method of introgressing an allele into a soybean plant by crossing at least two different parent soybean plants; obtaining a progeny soybean plant from the cross; obtaining DNA from a soybean seed of the progeny soybean plant using a non-destructive method; and selecting a soybean seed with the allele and a nucleic acid sequence selected from the group consisting of SEQ ID NOs: 143-174.

[0111] Another aspect includes a method of soybean plant breeding by crossing at least two different parent soybean plants, wherein the parent soybean plants differ in soybean plant maturity by over 10 days; obtaining progeny soybean seed from the cross; genotyping the progeny soybean seed of the cross with a genetic marker selected from the group consisting of SEQ ID NOs: 143-213; and selecting a soybean seed with a desired maturity group. A further aspect includes a soybean plant comprising within its genome an introgressed haplotype associated with maturity, wherein the introgression is facilitated by at least one of the markers from SEQ ID NO: 143-213 or of the markers 143-162.

[0112] Having now generally described the invention, the same will be more readily understood through reference to the following examples.

### Examples

Example 1: Discovery of molecular markers associated with genomic regions affecting plant maturity

[0113] Soybean is a short day plant, therefore flowering is initiated by short days due to a decrease in photoperiod (Garner & Allard, J. Agric. Res. 18, 553 - 606 (1920)). Consequently, photoperiod (day length) and temperature response of the soybean plant determines areas of plant adaptation. Due to photoperiod sensitivity, soybean genotypes are grown to narrow zones of latitude to optimize yield. Northern soybean varieties, in contrast to Southern varieties, initiate flowering with longer days. Northern varieties planted south of their adaptation zone exhibit accelerated flowering, limited plant growth and reduced yield. Southern soybean varieties planted north of their adaptation zone will have delayed flowering with a potential for frost damage that may reduce yield. Most soybean variety development crosses are made between parents within 10 maturity days of each other. If the parents differ greatly in maturity, progeny plants segregate widely for maturity. In order for breeders to obtain and select for soybean plants of a desire maturity group, they must produce and maintain a large number of progeny plants, the practice of which is cost prohibitive. Identification of genomic regions associated with plant maturity facilitated crosses between parents outside 10 maturity days of each other without maintain a large number of progeny plants.

[0114] To identify genomic regions associated with plant maturity, 258 soybean lines (129 pairs of differing maturity groups) are genotyped with one thousand, four hundred single nucleotide polymorphism (SNP) markers, distributed across the 20 linkage groups of the soybean genetic linkage map. In addition, 258 soybean lines are phenotyped for yield and plant maturity. Associations between SNP marker genotype and plant maturity phenotype are then evaluated. This was done in multiple environments (Tables 2-3).

Table 1: Initial identification of maturity genomic regions via marker assisted breeding

Region	Marker	SEQ ID NO:	Effect ( $\Delta d$ )	P- value
1	NS0125408	148	-0.05071	0.009068
1	NS0098982	155	1.242281	0.01081
2	NS0123506	156	-0.57638	0.021863
3	NS0093197	164	1.274868	1.92E-09

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(continued)

	<b>Region</b>	<b>Marker</b>	<b>SEQ ID NO:</b>	<b>Effect (<math>\Delta d</math>)</b>	<b>P- value</b>
5	3	NS0136544	171	1.162352	1.33E-10
	3	NS0119569	172	-1.87063	3.79E-15
	3	NS0114317	174	1.419675	3.01E-08
	5	NS0123168	188	-0.21704	0.025498
	6	NS0103755	190	-0.02572	0.011701
10	7	NS0095211	199	-0.09176	2.99E-07
	7	NS0097307	200	-0.09023	6.66E-07
	7	NS0102630	202	-0.08407	2.26E-06
	7	NS0102915	203	-0.08226	5.19E-06
	8	NS0100652	206	1.75824	3.92E-06
15	8	NS0119574	207	0.446757	0.045212
	8	NS0101020	212	0.829784	0.000462

Table 2: Estimated effect in days of maturity genomic regions

	<b>Region</b>	<b>Marker</b>	<b>SEQ ID NO:</b>	<b>Est. effect on plant maturity (<math>\Delta d</math>)</b>	<b>Effect (<math>\Delta d</math>)</b>	<b>P-value</b>
20	1	NS0124601	143	4.7	0.309636	0.156883
	1	NS0096829	145	4.8	0.444689	0.022932
	1	NS0099746	146	4.7	0.315142	0.191492
	1	NS0123747	147	4.9	0.714394	0.011568
	1	NS0125408	148	4.8	0.538569	0.015846
25	1	NS0128378	149	4.9	0.757069	0.01699
	1	NS0093976	154	5.1	0.989792	0.061019
	1	NS0098982	155	5.2	1.242281	0.01081
	2	NS0123506	156	4.1	0.911763	0.007307
	2	NS0097952	157	5.6	4.069668	5.06E-30
30	2	NS0118907	158	6.3	5.477999	1.01E-33
	2	NS0126989	160	4.6	1.994585	0.000191
	2	NS0095677	161	3.8	0.473053	0.10136
	3	NS0093197	164	5.2	1.274868	1.92E-09
	3	NS0103853	167	6	2.937938	3.78E-09
35	3	NS0136544	171	6.4	3.765493	3.23E-11
	3	NS0119569	172	5.8	2.409513	1.72E-21
	3	NS0123708	173	6	2.876505	3.44E-26
	3	NS0114317	174	5.9	2.627908	1.69E-22
	4	NS0098176	176	4.3	1.068684	6.45E-12
40	4	NS0100078	177	4	0.479955	0.073839
	4	NS0095530	179	4.5	1.364994	2.50E-09
	4	NS0129004	180	4.5	1.48424	8.04E-08
	5	NS0099024	181	3.4	0.732455	0.112193
	5	NS0101863	182	3.3	0.434912	0.078906
45	5	NS0103446	183	3.1	0.181809	0.058299
	5	NS0123168	188	3.2	0.217041	0.025498
	6	NS0103755	190	1.2	0.609071	0.140857
	6	NS0116125	191	0.9	0.456086	0.152892
	6	NS0125713	192	1.1	0.566084	0.036335
50	6	NS0125770	193	0.8	0.414212	0.009099
	6	NS0119281	194	1.6	0.797885	0.038077
	6	NS0124590	195	1.4	0.706375	0.000889

(continued)

	<b>Region</b>	<b>Marker</b>	<b>SEQ ID NO:</b>	<b>Est. effect on plant maturity (<math>\Delta d</math>)</b>	<b>Effect (<math>\Delta d</math>)</b>	<b>P-value</b>
5	6	NS0102717	196	1.5	0.749548	0.000246
	7	NS0099531	197	1.3	0.636575	0.000701
	7	NS0099417	198	2.4	1.181523	0.015954
	7	NS0095211	199	1.7	0.835736	0.099501
	7	NS0097307	200	0.2	0.090232	6.66E-07
	7	NS0102630	202	2.1	1.029761	0.046938
10	7	NS0102915	203	2.5	1.231387	4.37E-09
	8	NS0102362	204	4.8	2.23831	1.23E-09
	8	NS0117716	205	4.3	1.171503	9.09E-06
	8	NS0100652	206	4.6	1.75824	3.92E-06
	8	NS0119574	207	4.3	1.195594	4.79E-05
	8	NS0127728	208	4.5	1.630904	3.33E-07
15	8	NS0099639	209	4.2	1.037891	0.015656
	8	NS0103255	210	4.2	0.975115	0.001037
	8	NS0119106	211	4.3	1.18298	0.023909
	8	NS0101020	212	4.1	0.829784	0.000462
	8	NS0101779	213	4.2	1.000886	0.000563

[0115] The approximate locations of informative markers indicating a state of dominance or recessivity of genomic regions 1, 2, 3, 4, 5, 6, 7, and 8 are determined based upon a survey of polymorphisms among a panel of 258 soybean lines (Table 3 and 4). One factor in choosing these informative markers is based on which marker has the largest effect or is associated with the largest delay in maturity such that it is indicative of the maturity phenotype. Another factor in choosing these informative markers is based on the lowest P value, such that the marker does not get lost in the event of recombination. The markers with lower P value are more likely to be consistently associated with the maturity phenotype across different soybean populations (different parents, different pedigrees). Markers with strong association and predictive of introgression of the genomic region are listed in Table 5. For NS0128378, the SNP is actually an 11-bp indel, where "D" represents the deletion (\*\*\*\*\*\*) and "I" represents the insertion (TTCGAAGATT).

Table 3: Position of SNP markers associated with regions 1, 2, 3, 4, 5, 6, 7 and 8.

	<b>Region</b>	<b>LG</b>	<b>Position (cM)</b>	<b>Polymorphism position on Consensus Sequence</b>		
				<b>Marker</b>	<b>Sequence</b>	<b>SEQ ID NO:</b>
40	1	C2	113.7	NS0124601	884	143
	1	C2	121.9	NS0103749	96	144
	1	C2	121.9	NS0096829	225	145
	1	C2	121.9	NS0099746	330	146
	1	C2	121.9	NS0123747	56	147
	1	C2	121.9	NS0125408	133	148
50	1	C2	121.9	NS0128378	212	149
	1	C2	129.3	NS0135390	108	150
	1	C2	123	NS0099529	243	151
	1	C2	124.3	NS0097798	325	152
	1	C2	129.4	NS0093385	109	153
	1	C2	134.7	NS0093976	242	154
55	1	C2	134.7	NS0098982	383	155
	2	O	125.4	NS0123506	126	156
	2	O	127.7	NS0097952	420	157

(continued)

	Region	LG	Position (cM)	Marker	Polymorphism position on Consensus	
					Sequence	SEQ ID NO:
5	2	O	134.9	NS0118907	450	158
	2	O	151.4	NS0122182	104	159
	2	O	150.8	NS0126989	251	160
10	2	O	158.5	NS0095677	202	161
	3	L	99.4	NS0098853	82	162
	3	L	111.5	NS0092561	190	163
	3	L	99.4	NS0093197	225	164
15	3	L	100.4	NS0094891	83	165
	3	L	99.4	NS0096225	471	166
	3	L	136.2	NS0103853	341	167
20	3	L	114.2	NS0113929	685	168
	3	L	114.2	NS0115535	433	169
	3	L	113.6	NS0121511	512	170
	3	L	132.9	NS0136544	208	171
25	3	L	143.1	NS0119569	262	172
	3	L	145.8	NS0123708	530	173
	3	L	155.9	NS0114317	331	174
30	4	I	48.3	NS0092743	217	175
	4	I	49.6	NS0098176	92	176
	4	I	66.4	NS0100078	1412	177
	4	I	58.3	NS0137415	231	178
35	4	I	33.4	NS0095530	327	179
	4	I	32.3	NS0129004	1014	180
	5	L	40.1	NS0099024	69	181
40	5	L	35.7	NS0101863	381	182
	5	L	40.1	NS0103446	69	183
	5	L	35.9	NS0113878	375	184
	5	L	36.8	NS0115066	298	185
45	5	L	36.9	NS0119165	181	186
	5	L	36.8	NS0120015	449	187
	5	L	36	NS0123168	75	188
50	5	L	38.8	NS0123724	42	189
	6	D1b+W	172.5	NS0103755	45	190
	6	D1b+W	164.1	NS0116125	409	191
	6	D1b+W	176.3	NS0125713	392	192
55	6	D1b+W	165.4	NS0125770	1074	193
	6	D1b+W	134.8	NS0119281	596	194

(continued)

	Region	LG	Position (cM)	Polymorphism position on Consensus		SEQ ID NO:
				Marker	Sequence	
5	6	D1b+W	157.6	NS0124590	1092	195
	6	D1b+W	177.2	NS0102717	402	196
	7	G	111.5	NS0099531	287	197
10	7	G	122.1	NS0099417	408	198
	7	G	125.7	NS0095211	251	199
	7	G	125.7	NS0097307	426	200
15	7	G	130.4	NS0103004	430	201
	7	G	132.1	NS0102630	186	202
	7	G	131.2	NS0102915	193	203
20	8	M	37.7	NS0102362	74	204
	8	M	42.2	NS0117716	74	205
	8	M	44.2	NS0100652	247	206
	8	M	44.2	NS0119574	367	207
25	8	M	42.8	NS0127728	650	208
	8	M	48.8	NS0099639	362	209
	8	M	64.8	NS0103255	289	210
	8	M	64.8	NS0119106	417	211
30	8	M	67.1	NS0101020	238	212
	8	M	67.1	NS0101779	147	213

[0116] Allele-specific fluorescence-resonance-energy-transfer (FRET) probes are used in Real-Time PCR assays. Two FRET probes bearing different fluorescent reporter dyes are used, where a unique dye is incorporated into an oligonucleotide that can anneal with high specificity to only one of the two alleles. The reporter dyes are 2'-chloro-7'-phenyl-1,4-dichloro-6-carboxyfluorescein (VIC) and 6-carboxyfluorescein phosphoramidite (FAM).

Table 4: Listing of SNP markers associated with regions 1, 2, 3, 4, 5, 6, 7 and 8.

	Region	Marker	SEQ ID NO:	SEQ ID NO: Forward Primer	SEQ ID NO: Reverse Primer	SEQ ID NO: FAM Probe	FAM Allele	SEQ ID NO: VIC probe	VIC allele
40	1	NS0124601	143	1	2	214	T	215	G
45	1	NS0103749	144	3	4	216	G	217	A
	1	NS0096829	145	5	6	218	C	219	A
50	1	NS0099746	146	7	8	220	G	221	A
	1	NS0123747	147	9	10	222	T	223	A
	1	NS0125408	148	11	12	224	T	225	C
55	1	NS0128378	149	13	14	226	TTCGAAGATT	227	*****
	1	NS0135390	150	15	16	228	T	229	G
	1	NS0099529	151	17	18	230	T	231	A
	1	NS0097798	152	19	20	232	G	233	A

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(continued)

Region	Marker	SEQ ID NO:	SEQ ID NO: Forward Primer	SEQ ID NO: Reverse Primer	SEQID NO: FAM Probe	FAM Allele	SEQ ID NO: VIC probe	VIC allele
5	1 NS0093385	153	21	22	234	T	235	C
10	1 NS0093976	154	23	24	236	G	237	C
15	1 NS0098982	155	25	26	238	C	239	*
20	2 NS0123506	156	27	28	240	T	241	G
25	2 NS0097952	157	29	30	242	G	243	A
30	2 NS0118907	158	31	32	244	C	245	A
35	2 NS0122182	159	33	34	246	T	247	C
40	2 NS0126989	160	35	36	248	T	249	A
45	2 NS0095677	161	37	38	250	T	251	C
50	3 NS0098853	162	39	40	252	AG	253	**
55	3 NS0092561	163	41	42	254	T	255	C
60	3 NS0093197	164	43	44	256	G	257	A
65	3 NS0094891	165	45	46	258	T	259	G
70	3 NS0096225	166	47	48	260	C	261	A
75	3 NS0103853	167	49	50	262	T	263	C
80	3 NS0113929	168	51	52	264	G	265	C
85	3 NS0115535	169	53	54	266	T	267	G
90	3 NS0121511	170	55	56	268	T	269	C
95	3 NS0136544	171	57	58	270	T	271	C
100	3 NS0119569	172	59	60	272	T	273	A
105	3 NS0123708	173	61	62	274	G	275	A
110	3 NS0114317	174	63	64	276	G	277	A
115	4 NS0092743	175	65	66	278	AGAA	279	****
120	4 NS0098176	176	67	68	280	T	281	C
125	4 NS0100078	177	69	70	282	T	283	G
130	4 NS0137415	178	71	72	284	T	285	C
135	4 NS0095530	179	73	74	286	T	287	A
140	4 NS0129004	180	75	76	288	G	289	A
145	5 NS0099024	181	77	78	290	G	291	A
150	5 NS0101863	182	79	80	292	G	293	A
155	5 NS0103446	183	81	82	294	G	295	A
160	5 NS0113878	184	83	84	296	G	297	A
165	5 NS0115066	185	85	86	298	T	299	A
170	5 NS0119165	186	87	88	300	G	301	A
175	5 NS0120015	187	89	90	302	G	303	C
180	5 NS0123168	188	91	92	304	T	305	C

(continued)

Region	Marker	SEQ ID NO:	SEQ ID NO: Forward Primer	SEQ ID NO: Reverse Primer	SEQID NO: FAM Probe	FAM Allele	SEQ ID NO: VIC probe	VIC allele
5	NS0123724	189	93	94	306	G	307	A
6	NS0103755	190	95	96	308	T	309	A
10	NS0116125	191	97	98	310	T	311	C
6	NS0125713	192	99	100	312	G	313	A
6	NS0125770	193	101	102	314	G	315	A
15	NS0119281	194	103	104	316	G	317	A
6	NS0124590	195	105	106	318	T	319	C
6	NS0102717	196	107	108	320	G	321	A
20	NS0099531	197	109	110	322	AA	323	**
7	NS0099417	198	111	112	324	G	325	C
7	NS0095211	199	113	114	326	T	327	C
7	NS0097307	200	115	116	328	G	329	C
25	NS0103004	201	117	118	330	G	331	A
7	NS0102630	202	119	120	332	C	333	A
7	NS0102915	203	121	122	334	G	335	A
30	NS0102362	204	123	124	336	T	337	C
8	NS0117716	205	125	126	338	ACTT	339	****
8	NS0100652	206	127	128	340	T	341	A
8	NS0119574	207	129	130	342	G	343	A
35	NS0127728	208	131	132	344	G	345	A
8	NS0099639	209	133	134	346	T	347	C
8	NS0103255	210	135	136	348	T	349	C
40	NS0119106	211	137	138	350	G	351	A
8	NS0101020	212	139	140	352	G	353	C
45	NS0101779	213	141	142	354	G	355	C

Table 5: Most predictive markers for genomic regions associated with plant maturity and/or growth habit of soybean plants

Region	Marker	SEQ ID NO:	Rec. Allele	Dom. Allele	
1	NS0099529	151	A	T	
50	1	NS0128378	149	***** TTCGAAGATT	
2	NS0118907	158	A	C	
3	NS0115535	169	T	G	
4	NS0137415	178	C	T	
55	5	NS0120015	187	C	G
6	NS0125713	192	A	G	

(continued)

Region	Marker	SEQ ID NO:	Rec. Allele	Dom. Allele
7	NS0102630	202	C	A
8	NS0102362	204	C	T

[0117] SNP markers associated with region 1 include SEQ ID NO: 143 through SEQ ID NO: 155. All of these SNP makers for region 1 map to a region on linkage group C2. Table 4 lists sequences for PCR amplification primers, indicated as SEQ ID NO: 1 through SEQ ID NO: 26, and probes indicated as SEQ ID NO: 214 through SEQ ID NO: 239.

[0118] SNP markers associated with region 2 include SEQ ID NO: 156 through SEQ ID NO: 161. All of these SNP makers for region 2 map to a region on linkage group O. Table 4 lists sequences for PCR amplification primers, indicated as SEQ ID NO: 27 through SEQ ID NO: 38, and probes indicated as SEQ ID NO: 240 through SEQ ID NO: 251.

[0119] SNP markers associated with region 3 include SEQ ID NO: 162 through SEQ ID NO: 174. All of these SNP makers for region 3 map to a region on linkage group L. Table 4 lists sequences for PCR amplification primers, indicated as SEQ ID NO: 39 through SEQ ID NO: 64, and probes indicated as SEQ ID NO: 252 through SEQ ID NO: 277.

[0120] SNP markers associated with region 4 include SEQ ID NO: 175 through SEQ ID NO: 180. All of these SNP makers for region 4 map to a region on linkage group I. Table 4 lists sequences for PCR amplification primers, indicated as SEQ ID NO: 65 through SEQ ID NO: 76 and probes indicated as SEQ ID NO: 278 through SEQ ID NO: 289.

[0121] SNP markers associated with region 5 include SEQ ID NO: 181 through SEQ ID NO: 189. All of these SNP makers for region 5 map to a region on linkage group L. Table 4 lists sequences for PCR amplification primers, indicated as SEQ ID NO: 77 through SEQ ID NO: 94, and probes indicated as SEQ ID NO: 290 through SEQ ID NO: 307.

[0122] SNP markers associated with region 6 include SEQ ID NO: 190 through SEQ ID NO: 196 of these SNP makers for region 6 map to a region on linkage group D1b. Table 4 lists sequences for PCR amplification primers, indicated as SEQ ID NO: 95 through SEQ ID NO: 108, and probes indicated as SEQ ID NO: 308 through SEQ ID NO: 321.

[0123] SNP markers associated with region 7 include SEQ ID NO: 197 through SEQ ID NO: 203. Table 4 lists sequences for PCR amplification primers, indicated as SEQ ID NO: 109 through SEQ ID NO: 122, and probes indicated as SEQ ID NO: 322 through SEQ ID NO: 333.

[0124] SNP markers associated with region 8 include SEQ ID NO: 204 through SEQ ID NO: 213 of these SNP makers map. Table 4 lists sequences for PCR amplification primers, indicated as SEQ ID NO: 123 through SEQ ID NO: 142 and probes indicated as SEQ ID NO: 336 through SEQ ID NO: 355.

Example 2: Identifying allelic combinations of genomic regions associated with plant maturity in early maturity group soybeans

[0125] Genomic regions 1 and 2 are used to predict the plant maturity of progeny plant resulting from a cross between early maturity and mid-maturity parents (III - V). In particular, the allelic combinations of genomic regions 1 and 2 are correlated with a delay in plant maturity. To determine the correlation between allelic combinations of region 1 and 2 and delay in plant maturity, three populations are developed from crossing an early maturity parent (maturity group 00) with a mid-maturity parent (maturity group III or IV) (Table 6). Populations 1-3 are used to determine the association of the composition of genomic regions 1 and 2 with delay in plant maturity.

Table 6: Maturity group phenotype of parents in soybean populations

Population	Maturity Group of Female Parent	Maturity Group of Female Parent
1	00.9	3.1
2	00.9	3.4
3	00.9	4.1
4	5.9	4.7
5	5.9	5.1
6	5.8	4.7
7	4.1	00.9
8	3.1	00.9
9	3.4	00.9

[0126] The three populations segregate widely for maturity and are polymorphic at genomic regions 1 and 2. F<sub>3</sub> seed are obtained by selecting one pod per F<sub>2</sub> plant (modified single seed descent). The F<sub>3</sub> populations are planted in Guelph,

ON and 1,214 F<sub>3</sub> individuals from all three populations are phenotyped for genomic regions 1 and 2 with the SNP markers NS0128378 (genomic region 1) and NS0118907 (genomic region 2). Individual plants in the F<sub>3</sub> populations are also genotyped for maturity by counting the number of days after August 31<sup>st</sup> until plant matures; plants are considered mature when 95% of the pods were brown. The procedure is repeated with 1055 of the individual plants where each plant row is grown in Chile and phenotyped for maturity by counting the number of days after March 1<sup>st</sup> until plant matures; plants are considered mature when 95% of the pods are brown. The procedure is repeated with experimental breeding lines developed from 88 of the 1055 individual plants. Table 8 compares the days to maturity of individual plants across all three populations and the genotype of the individuals at genomic regions 1 and 2. The markers associated with 1 and 2 explain 64% of the variation in plant maturity in year 1 and 94% of the variation in plant maturity in year 2.

Table 7: The association of days to maturity with composition of regions 1 and 2. Presence (1) or absence (0) of dominant allele indicated. Homozygous allele states are 0,0 and 1,1. Heterozygous allele state is 0,1.

Allelic Combination	Region 1	Region 2	Days to Maturity (D after August 31 <sup>st</sup> )	
			Year 1	Year 2
1	0,0	0,0	19.2	9.5
2	0,0	0,1	25.7	13.5
3	0,0	1,1	33.6	15.5
4	0,1	0,0	26.2	16.4
5	0,1	0,1	40.3	ND
6	0,1	1,1	49.1	19.5
7	1,1	0,0	34.2	17.11
8	1,1	0,1	49.3	22.7
9	1,1	1,1	53.5	23.9
Correlation:			64%	94%

Example 3: Identifying allelic combinations of genomic regions associated with plant maturity in late maturity group soybeans

[0127] Genomic regions 1, 2, and 3 are used to predict the plant maturity of progeny plant resulting from a cross between late maturity and mid-maturity parents. In particular, some of the allelic combinations of genomic regions 1, 2 and 3 are correlated with a delay in plant maturity (Table 8 and 9). To determine the correlation between allelic combinations of region 1, 2 and 3 and delay in plant maturity, three F<sub>3</sub> populations are developed from crossing a late maturity group V with a late maturity group IV. The populations 4-6 following crosses are used to determine the association of the composition of genomic regions 1, 2 and 3 with delay in plant maturity.

[0128] The three segregate widely for maturity and are polymorphic at genomic regions 1, 2, and 3. F<sub>3</sub> seed are obtained by selecting one seed per F<sub>2</sub> plant (single seed descent). 5,984 F<sub>3</sub> individuals from all three population are genotyped with the SNP markers NS0099529 (genomic region 1), NS0118907 (genomic region 2), and NS0115535 (genomic region 3) and seeds with the same marker haplotype are bulked. F<sub>3</sub> seeds are planted.

Table 8: Summary of days to flowering for soybean lines containing various compositions of genomic regions 1, 2, and 3 for plant maturity. Presence (1) or absence (0) of dominant allele indicated. Homozygous allele states are 0,0 and 1,1. Heterozygous allele state is 0,1. ND = no data.

Allelic Combination	Region 1	Region 2	Region 3	Days to flowering (DAP)		
				Pop. 4	Pop. 5	Pop. 6
10	1,1	0,0	1,1	57	57	57
11	1,1	1,0	1,1	58	57	58
12	1,1	1,1	0,0	58	59	55
14	1,1	0,0	0,0	ND	ND	54

(continued)

Allelic Combination	Region 1	Region 2	Region 3	Days to flowering (DAP)		
				Pop. 4	Pop. 5	Pop. 6
15	0,1	0,1	0,1	59	57	56
16	0,0	1,1	1,1	43	36	41
17	0,0	0,0	1,1	44	38	45
18	0,0	1,1	0,0	44	39	44
19	0,0	0,0	0,0	44	38	43

[0129] The individuals are also phenotyped for maturity by counting the number of days after August 31<sup>st</sup> until plant matures; plants are considered mature when 95% of the pods were brown. Genomic region 3 influences the time of maturity (Tables 8 and 9).

Table 9: Summary of days to plant maturity for soybean lines containing various compositions of genomic regions 1, 2, and 3 for plant maturity. ND = no data.

Allelic Combination	Days to Maturity (D after Aug)		
	Pop. 4	Pop 5	Pop 6
10	59	58	58
11	54	58	58
12	59	57	59
14	ND	ND	58
15	54	54	53
16	41	35	37
17	37	35	38
18	44	44	43
19	38	42	43

#### Example 4: Discovery of molecular markers associated with genomic regions affecting plant growth habit

[0130] Plant growth habit is an important characteristic for late maturity group growing regions. To identify genomic regions associated with plant growth habit, three F<sub>3</sub> populations are developed from crossing a late maturity group V (determinate growth habit) with a late maturity group IV (indeterminate growth habit). Populations 4-6 are used to determine the association of the genomic region 3 with plant habit (Table 6). Seven hundred and seventy-four soybean lines are screened with the markers associated with genomic region 3. The three populations segregated widely for maturity and are polymorphic at genomic region 3. F<sub>3</sub> seed are obtained by selecting one seed per F<sub>2</sub> plant (single seed descent). 5,984 F<sub>3</sub> individuals from all three population were phenotyped with the SNP NS0115535 (genomic region 3) and seeds with the same marker haplotype are bulked. F<sub>3</sub> seeds are planted. A single marker, NS00115535, is determined to be most predictive and able to separate determinant group V varieties from indeterminant group IV and earlier varieties.

#### Example 5: Genomic regions associated with growth habit and maturity independent of yield

[0131] Plant maturity and yield are closely associated in soybean. An increase of one day in maturity may be equivalent to a -0.7 bu/A increase in yield. The correlation of plant maturity and yield confounds the evaluation of potential QTLs and candidate genes associated with yield. Identification of genomic regions associated with plant maturity allows breeders to genetically fix plant maturity within a soybean plant and elucidate traits associated with yield.

[0132] Three soybean populations are generated from crossing a maturity group 0 with a maturity group III or IV. Populations 7-9 are used (Table 5). The progeny seed planted in Chile and then harvested 52 seeds from those progeny plants are selected in Chile and the plants are grown in Ontario in 2006. Eighty-four progeny are screened with markers

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associated maturity regions 1 and 2 and evaluated for maturity days and yield (Table 10-12). Markers associated with regions 1 and 2 select for maturity and are independent of yield. For example, Progeny 0430 has significantly higher yield than Progeny 0083 (Table 11). The higher yield of Progeny 0430 is not attributed to differences in plant maturity due similar days to maturity and allelic states of maturity genomic regions 1 and 2.

5

Table 10: Summary of yield, maturity and the allelic combination for maturity regions 1 and 2.

<b>Pedigree</b>	<b>Progeny ID No.</b>	<b>Best Est. Yield (Bu/A)</b>	<b>Maturity Days</b>	<b>Allelic combination</b>
10	Population 8	0117	30.93	5.50
	Population 8	0140	29.18	6.50
	Population 8	0234	32.84	6.50
	Population 8	0043	34.67	6.50
	Population 8	0267	36.80	7.00
15	Population 8	0276	40.67	7.50
	Population 8	0243	42.88	9.50
	Population 8	0198	39.56	10.50
	Population 8	0325	33.42	11.00
	Population 8	0011	39.92	11.50
20	Population 8	0390	41.22	11.50
	Population 8	0418	44.05	11.50
	Population 8	0119	41.62	9.50
	Population 8	0069	37.68	10.00
	Population 8	0274	38.90	10.00
25	Population 8	0165	43.03	10.00
	Population 8	0219	39.67	12.50
	Population 8	0373	49.22	13.00
	Population 8	0089	50.41	17.00
	Population 8	0186	43.74	18.00
30	Population 8	0395	43.20	9.50
	Population 8	0426	41.12	10.00
	Population 8	0256	43.83	10.00
	Population 8	0216	45.47	10.50
	Population 8	0367	47.94	11.50
35	Population 8	0266	42.86	14.00
	Population 8	0285	42.04	16.00
	Population 8	0277	50.47	16.00
	Population 8	0188	45.62	17.50
	Population 8	0143	44.47	13.50
40	Population 8	0101	41.22	14.50
	Population 8	0366	41.79	16.50
	Population 8	0340	47.41	11.50
	Population 8	0359	46.10	14.50
	Population 8	0184	46.24	14.50
45	Population 8	0158	43.08	16.00
	Population 8	0401	50.95	16.00
	Population 8	0255	47.26	17.00
	<b>Overall Mean Non-Check</b>	42.78	12.00	
	<b>Mean</b>	42.60	12.38	
55	<b>Check Mean</b>	44.08	9.25	
	<b># Locs</b>	3	2	
	<b># Reps</b>	3	2	
	<b>CV</b>	9.978	15.094	

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(continued)

<b>Pedigree</b>	<b>Progeny ID No.</b>	<b>Best Est. Yield (Bu/A)</b>	<b>Maturity Days</b>	<b>Allelic combination</b>
5	<b>LSD(.05)</b>	6.989	3.640	
	<b>F-Statistic</b>	4.525	7.670	
	<b>P-Value</b>	0.000	0.000	
	<b>Repeatability</b>	0.781	0.870	
	<b>Root MSE</b>	4.269	1.811	

10

Table 11: Summary of yield, maturity and the allelic combination for maturity regions 1 and 2.

<b>Pedigree</b>	<b>Progeny ID No.</b>	<b>Best Est. Yield (Bu/A)</b>	<b>Maturity (D)</b>	<b>Allelic Combination</b>
15	Population 9	0381	38.46	11.00
	Population 9	0473	40.89	12.50
	Population 9	0371	36.86	9.00
	Population 9	0380	31.86	10.00
	Population 9	0263	43.01	11.00
20	Population 9	0396	38.97	12.00
	Population 8	0083	29.01	15.00
	Population 8	0430	42.65	15.00
	Population 9	0299	39.96	16.00
	Population 8	0076	42.95	22.00
25	Population 9	0142	32.31	11.50
	Population 9	0487	27.86	14.00
	Population 8	0240	43.66	15.50
	Population 9	0317	46.74	16.50
	Population 8	0392	38.21	18.50
30	Population 9	0206	45.77	19.00
	Population 9	0254	44.06	19.50
	Population 8	0280	48.22	26.50
	Population 9	0262	41.41	17.50
	Population 9	0173	43.17	23.50
35	Population 9	0032	33.65	13.50
	Population 9	0166	40.72	11.50
	Population 9	0188	42.19	16.50
	Population 9	0117	47.98	19.00
	Population 8	0229	45.34	20.00
40	Population 9	0437	43.25	20.50
	Population 9	0077	34.05	10.50
	Population 9	0078	47.66	17.00
	Population 9	0187	37.18	27.00
	Population 8	0230	47.26	20.50
45	Population 9	0368	46.49	21.50
	Population 9	0505	34.06	23.50
	<b>Overall Mean</b>	39.96	15.69	
	<b>Non-Check Mean</b>	40.38	16.57	
	<b>Check Mean</b>	37.07	9.50	
50	<b># Locs</b>	3	2	
	<b># Reps</b>	3	2	
	<b>CV</b>	15.453	13.984	
	<b>LSD(.05)</b>	10.105	4.434	

(continued)

Pedigree	Progeny ID No.	Best Est. Yield (Bu/A)	Maturity (D)	Allelic Combination
<b>F-Statistic</b>		2.546	10.862	
<b>P-Value</b>		0.000	0.000	
<b>Repeatability</b>		0.609	0.908	
<b>Root MSE</b>		6.176	2.194	

Table 12: Summary of yield, maturity and the allelic combination for maturity regions 1 and 2.

Pedigree	Progeny ID No.	Best Est. Yield (Bu/A)	Maturity (D)	Allelic Combination
Population 7	0121	35.25	8.50	1
Population 7	0107	30.98	10.50	1
Population 7	0251	36.59	10.50	1
Population 7	0377	34.51	11.00	1
Population 7	0375	34.34	11.50	1
Population 7	0326	30.51	13.00	1
Population 7	0216	42.26	10.50	2
Population 7	0312	36.15	18.00	2
Population 7	0298	41.40	19.00	2
Population 7	0205	39.41	13.00	3
Population 7	0139	38.59	14.50	3
Population 7	0365	38.14	13.00	4
Population 7	0004	39.79	12.50	5
Population 7	0361	47.75	24.00	8
<b>Overall Mean</b>		39.37	12.55	
<b>Non-Check Mean</b>		37.79	13.57	
<b>Check Mean</b>		44.10	9.50	
<b># Locs</b>		3	2	
<b># Reps</b>		3	2	
<b>CV</b>		16.518	11.343	
<b>LSD(.05)</b>		10.749	2.979	
<b>F-Statistic</b>		3.074	16.491	
<b>P-Value</b>		0.002	0.000	
<b>Repeatability</b>		0.675	0.939	
<b>Root MSE</b>		6.503	1.423	

Example 6: Utilization of molecular markers associated with plant maturity to select geographic region for planting seed

[0133] Soybean genotypes are grown to narrow zones of latitude to optimize yield due to photoperiod sensitivity. Northern soybean varieties, in contrast to Southern varieties, initiate flowering with longer days. Northern varieties planted south of their adaptation zone exhibit accelerated flowering, limited plant growth and reduced yield. Southern soybean varieties planted north of their adaptation zone have delayed flowering with a potential for frost damage that may reduce yield. When the parents differ in plant maturity greater than 10 day, the progeny of the cross segregate widely for plant maturity. Molecular markers associated with plant maturity genomic regions allows breeders to cross with parents that differ in maturity greater than 10 days, select seed of the cross to grow in the appropriate maturity zone.

[0134] A BC<sub>2</sub>F<sub>1</sub> soybean population is generated by crossing MG III.5 with MG 000 and the seed is selected for the appropriate maturity zone growing region using the molecular markers associated with plant maturity. Ninety-three BC<sub>2</sub>F<sub>1</sub> plants are screened with 106 SNP markers to evaluate the genetic similarity to the recurrent MG III.5 parent (Table 13). Additionally, the SNP markers included markers associated with the maturity genomic regions 1, 2, 3, 4, and 5. Each individual is heterozygous for at least one maturity genomic region. Individual Progeny:0107 is heterozygous for 1, 2, 3, 4, and 5 and may be used to select for individual varieties adapted to each maturity group zone. Individuals selected

to move forward to the next generation based on adaptation to specific maturity group regions using the allelic combination for the genomic maturity regions.

5 Table 13: Summary of heterozygosity for maturity genomic regions with the F2 generation of MG III.5 parent/(MG III.5 parent\*2/MG 000 parent). Individuals within the population are selected for a geographic maturity group region with SNP markers associated maturity genomic regions.

	Plant	Similarity to MGIII.5 parent (%)	Heterozygous for genomic maturity region:				
			1	2	3	4	5
10	MG III.5 parent	98.7					
15	MG 000 parent	2.6					
20	Progeny: 0050	86.2	x			x	x
25	Progeny: 0107	85.8			x	x	
30	Progeny: 0050	84.9	x	x			
35	Progeny: 0093	84.9	x		x		
40	Progeny: 0050	82.8		x	x	x	x
45	Progeny: 0096	82.8			x	x	
50	Progeny: 0107	82.3		x			
55	Progeny: 0096	81.9	x			x	
	Progeny: 0107	81.5	x	x	x	x	x
	Progeny: 0066	81.9				x	
	Progeny: 0096	81.9	x		x	x	
	Progeny: 0093	82.8	x	x			
	Progeny: 0050	81.9	x		x	x	
	Progeny: 0050	81.9	x		x		
	Progeny: 0096	81.0	x		x	x	
	Progeny: 0046	80.6		x	x	x	x
	Progeny: 0050	80.2	x	x	x		
	Progeny: 0107	80.2	x		x	x	
	Progeny: 0093	80.2	x			x	
	Progeny: 0096	80.2		x			

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(continued)

	Plant	Similarity to MGIII.5 parent (%)	Heterozygous for genomic maturity region:				
			1	2	3	4	5
5	Progeny: 0093	79.7	x			x	
10	Progeny: 0063	79.7			x	x	
15	Progeny: 0093	79.3		x	x		x
20	Progeny: 0096	78.9	x		x		
25	Progeny: 0012	78.9	x			x	x
30	Progeny: 0085	78.4	x		x	x	
35	Progeny: 0096	78.0	x				
40	Progeny: 0107	77.6	x		x		
45	Progeny: 0063	74.6		x	x		x
50	Progeny: 0063	74.1	x			x	
55	Progeny: 0012	61.2	x	x	x	x	
	Progeny: 0036	61.2	x	x	x	x	
	Progeny: 0012	61.2	x	x	x	x	
	Progeny: 0093	61.2	x		x	x	x
	Progeny: 0012	61.2	x		x		x
	Progeny: 0050	61.2	x		x		
	Progeny: 0036	61.2	x		x		
	Progeny: 0063	61.2	x			x	x
	Progeny: 0050	61.2	x			x	
	Progeny: 0012	61.2	x			x	
	Progeny: 0107	61.2	x				
	Progeny: 0012	61.2	x				
	Progeny: 0012	60.8	x		x	x	
	Progeny: 0012	60.8	x		x	x	
	Progeny: 0012	60.8	x		x	x	

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(continued)

	Plant	Similarity to MGIII.5 parent (%)	Heterozygous for genomic maturity region:				
			1	2	3	4	5
5	Progeny: 0050	60.8	x		x		
	Progeny: 0012	60.8	x			x	
10	Progeny: 0036	60.8	x			x	
	Progeny: 0012	60.8	x				
15	Progeny: 0012	60.8	x				
	Progeny: 0036	60.8		x	x	x	
20	Progeny: 0012	60.8			x	x	x
	Progeny: 0012	60.3	x	x			
	Progeny: 0093	59.9	x	x	x	x	
25	Progeny: 0096	59.9	x		x	x	x
	Progeny: 0012	59.9	x		x		
30	Progeny: 0050	59.9		x	x		x
	Progeny: 0085	59.9		x	x		x
	Progeny: 0050	59.5	x	x			
35	Progeny: 0096	59.5	x	x		x	
	Progeny: 0036	59.5	x	x		x	
40	Progeny: 0096	59.5	x		x	x	x
	Progeny: 0063	59.5	x		x		
	Progeny: 0036	59.5	x		x		
45	Progeny: 0096	59.5		x			x
	Progeny: 0093	58.6	x	x		x	
50	Progeny: 0050	58.6	x				x
	Progeny: 0050	58.6	x				
55	Progeny: 0093	58.6		x	x	x	
	Progeny: 0093	58.2	x	x			

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(continued)

	Plant	Similarity to MGIII.5 parent (%)	Heterozygous for genomic maturity region:				
			1	2	3	4	5
5	Progeny: 0012	58.2	x	x		x	
10	Progeny: 0012	58.2	x		x	x	x
15	Progeny: 0050	58.2	x		x	x	
15	Progeny: 0012	58.2	x		x	x	
15	Progeny: 0012	58.2	x		x		
20	Progeny: 0143	58.2	x		x		
20	Progeny: 0096	58.2	x			x	
20	Progeny: 0050	58.2	x			x	
25	Progeny: 0012	57.8	x	x		x	
25	Progeny: 0050	57.8	x	x		x	
25	Progeny: 0012	57.8	x		x		
30	Progeny: 0093	57.8	x			x	
30	Progeny: 0093	57.8	x			x	x
35	Progeny: 0012	57.8		x		x	x
35	Progeny: 0012	57.8			x	x	
40	Progeny: 0096	57.3	x		x	x	
40	Progeny: 0050	56.9	x	x		x	
45	Progeny: 0093	56.9	x		x	x	
45	Progeny: 0050	56.9	x		x	x	
50	Progeny: 0050	56.9	x		x		x
55	Progeny: 0096	55.6	x	x		x	x

55 Example 7: Estimating effect of genomic regions associated with maturity

[0135] Each allele of each individual maturity genomic region is associated with a value that can either increase or

decrease the relative maturity of a given line. The relative maturity of a given line are predicted by using an additive or epistatic model. The example in Table 14 demonstrates predicting relative maturity based on the allelic combination of the maturity genomic regions. The maturity group of a soybean seed is predicted by the composition of maturity genomic region alleles.

5

Table 14: An example of predicting relative maturity based on additive model

Maturity genomic	Δ Days	Direction
10	1	10
	2	-5
	3	-3
	4	2
	5	6
	6	4
	7	-5
Sum		9
Constant		3
Maturity Days		12
Maturity Group		1.2

Example 8: Utilization of molecular markers associated with plant maturity to facilitate crosses with exotic germplasm

25 [0136] The genetic base of cultivated soybean is narrow compared to other field crops. Eighty to ninety percent of cultivated soybean gene pool are traced to 12 plant introductions in northern United States and seven plant introductions in southern United States. Due to the narrow genetic base, soybean is more likely to be impacted by disease and insect attacks. Exotic germplasm helps expand the genetic base of soybean. In addition, exotic germplasm possesses such key traits as disease resistance, insect resistance, nematode resistance, and tolerance to environmental stress. At present, many exotic species are inaccessible in part due to limitations with crossing soybean plants from extremely different maturity groups. Traditionally, breeders must produce and maintain large numbers of progeny plants from crosses between exotic and cultivated germplasm, in order for breeders to select for a small number soybean plants of the desired maturity group. It is often cost prohibitive to maintain the large number of plants required.

30 [0137] Molecular markers associated with plant maturity facilitate the used of exotic germplasm. Breeders create crosses between exotic and cultivated germplasm. The progeny seed is assayed for plant maturity without expending the resources required to plant and grow large numbers of progeny.

Example 9: Utilization of molecular markers associated with plant maturity to facilitate introgression of a transgene

40 [0138] After a transgene is introduced into a variety, it may readily be transferred to other varieties by crossing. Most soybean variety development crosses are made between parents within 10 maturity days of each other. When parents differ in plant maturity greater than 10 days, the progeny of the cross segregate widely for plant maturity. In order for breeders to obtain and select for soybean plants of the desire maturity group, they must produce and maintain a large number of progeny plants, the practice of which is cost prohibitive. If a transgene is present in a maturity group III variety needs to be transferred to maturity group 0, a direct cross between a maturity group III variety and a maturity group 0 variety is not typically performed. Instead, the transgene is transferred through a series of intermediate crosses between varieties close in plant maturity. Molecular markers associated with plant maturity genomic regions allows breeders to cross parents that differ in maturity greater than 10 days, then select seed of the cross based on the presence of the transgene and the plant maturity phenotype.

45 [0139] If a variety possesses a desirable trait, it may readily be transferred to other varieties by crossing. Most soybean variety development crosses are made between parents within 10 maturity days of each other. When the parents differ in plant maturity greater than 10 days, the progeny of the cross segregate widely for plant maturity. In order for breeders to obtain and select for soybean plants of the desire maturity group, they must produce and maintain a large number of progeny plants, the practice of which is cost prohibitive. If a trait is present in a maturity group III variety needs to be transferred to maturity group 0, a direct cross between a maturity group III variety and a maturity group 0 variety is

50 Example 10: Utilization of molecular markers associated with plant maturity to facilitate introgression of a trait

55 [0139] If a variety possesses a desirable trait, it may readily be transferred to other varieties by crossing. Most soybean variety development crosses are made between parents within 10 maturity days of each other. When the parents differ in plant maturity greater than 10 days, the progeny of the cross segregate widely for plant maturity. In order for breeders to obtain and select for soybean plants of the desire maturity group, they must produce and maintain a large number of progeny plants, the practice of which is cost prohibitive. If a trait is present in a maturity group III variety needs to be transferred to maturity group 0, a direct cross between a maturity group III variety and a maturity group 0 variety is

typically not performed. Instead, the trait is transferred through a series of intermediate crosses between varieties close in plant maturity. Molecular markers associated with plant maturity genomic regions allow breeders to cross with parents that differ in maturity by greater than 10 days and to select seed of the cross based on the presence of the trait and the plant maturity phenotype.

5

Example 11: Utilization of molecular markers associated with plant maturity to select environments to optimize expression of traits

[0140] Soybeans cultivated in different environments often perform differently. For instance, a soybean variety may produce seeds with a particular fatty acid profile in one environment and a different fatty acid profile in another environment. A number of environmental factors can influence the expression of traits, including soil type, soil conditions, temperature, photoperiod, geography and cultural practices. Variation in performance of genotypes across different environments is often referred to genotype x environment interactions.

[0141] Soybean seed oil levels are highly impacted by environment. Oil concentration increases with decreasing latitude, therefore, soybeans in maturity groups 00-I generally have lower oil levels than later maturing soybeans (Figure 1). Molecular markers associated with plant maturity assist breeders in selecting soybean genotypes and produce plants that are better adapted to a maturity group region to produce higher oil.

[0142] Soybean seed fatty acid composition is highly impacted by the latitude of cultivation. The present invention provides molecular markers associated with plant maturity which are useful for assisting plant breeders to select favorable soybean maturity genotypes to optimize the expression of particular traits in specific geographies, such as fatty acid synthesis, wherein the trait is conventional or transgenic. As used herein, conventional traits include those obtained by mutagenesis. For example, the profile of fatty transgenic soybean plants engineered to produce stearidonic acid (SDA) have a positive correlation with latitude for SDA production and have a negative correlation with latitude for oleic acid, stearic acid, palmitic acid and  $\alpha$ -linolenic acid production (Table 15). The percent of SDA increases with increasing latitude (Figs. 2-3).

Table 15: Correlation of longitude and latitude on fatty acids for mature soybean seed

<b>Fatty Acid</b>	<b>Latitude</b>			<b>Longitude</b>		
	<b>R</b>	<b>P value</b>	<b>N</b>	<b>R</b>	<b>P value</b>	<b>N</b>
<b>stearidonic acid</b>	0.6625*	3.12E-10	71	-0.3748	0.001281263	71
<b><math>\gamma</math>-linolenic acid</b>	0.1097	0.362504877	71	-0.0798	0.508051934	71
<b>oleic acid</b>	-0.4081 *	0.000411819	71	0.167	0.16389379	71
<b>linoleic acid</b>	-0.1581	0.187769857	71	0.0837	0.48752276	71
<b><math>\alpha</math>-linolenic acid</b>	-0.2403*	0.043495686	71	0.1901	0.112261464	71
<b>palmitic acid</b>	-0.7305*	4.82E-13	71	0.4592	5.62E-05	71
<b>stearic acid</b>	-0.258*	0.029810388	71	-0.1498	0.212583113	71

\*signifcant at 0.05 level

[0143] Latitude is closely related with maturity groups and growing regions. Soybeans are classified into 13 maturity groups (000, 00, 0, I-X) according to the range in latitude in which the plants are adapted and most productive. Group 000 are the earliest maturing and cultivated at the higher latitudes and Group X are the latest maturing and cultivated in lower latitudes. Molecular markers associated with plant maturity will assist breeders in selecting soybean genotypes that are adapted to latitudes known to be associated with preferred SDA production in the plants. As a result, the soybean breeders more efficiently produce plants that are better adapted to the environment and produce higher levels of SDA or other similar traits.

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&lt;223&gt; Synthetic Primer

&lt;400&gt; 142

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&lt;210&gt; 143

&lt;211&gt; 1040

&lt;212&gt; DNA

&lt;213&gt; Glycine max

10

&lt;400&gt; 143

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 acaacacat g gcat gat t gg agccgtt at at gat cct ac gcat gaaaat gt tt caact a 120  
 cacat ccgt t cccgt caag aat gggagag gat ccgt at g cgat gaact g aact t gaat t 180  
 gatt cat t a at at agt gag agagaaaaaa gtt aaacca t caaat ggt t gat t gct tt 240  
 agt ttt at at ttccctttta caaatt aacc ct att gtt aa cagattt aatt t ggtt aat ga 300  
 at at tttt tttt tctttt at tctttt aat ttcaat caaa caattttt att tttt actttt 360  
 tttt ctattt ct gtcattt a ttttcat tttt ct cacgat ca aacagaggat tagt ct aaaa 420  
 aaat att taaa t aat gctt ga ttttattt gga act aattt ct t aattt cat ga ccggaat att 480  
 cacat gaattt aattt gaaaaa t gttt aagat t ggtt agattt ggattt aattt actt gacttt 540  
 cttaattt gtc tttt at gaa tttt gact aac ct aattt ct tttt attt attt tttt g cgaagaaaga 600  
 agt attt attt g t at ccgt gt g t gt at at at a aaat aaagt c attt caat cgg t cctt aattt a 660  
 cacaagat ac at gt caaat a t gcaa at gaa gt aact ct tttt gat ct gaaaa aaaaaaaaaa 720  
 aaaaaaacaat ccagt tttcc ct t gttt gaaaaa aagagctcca aat agcttca ggtt gaagca 780  
 aaaaat aaaaat attt gaagaaa aggtt gaagc taaacat aaa cct caaaaac t ggtt gt acgt 840  
 cattt aacat g ggt gaccccg aagt t gcccac gt act ccaag cgt gt gcgg t aacaacgt a 900  
 cgagagt at c gaat ct gcct ct gctttt ct tcaattt caa cagaacccat cacacacaca 960  
 cagaccccat caaccaaaaaa caaagaacaa t gat t ct gag attt gcagca gct gcagg t c 1020  
 gact ct agga aagacccggg 1040

&lt;210&gt; 144

&lt;211&gt; 821

&lt;212&gt; DNA

&lt;213&gt; Glycine max

45

&lt;220&gt;

&lt;221&gt; unsure

&lt;222&gt; (1)..(821)

&lt;223&gt; unsure at all n locations; n = a, t, c, or g

50

&lt;400&gt; 144

55

## EP 2 134 870 B1

at tccat aac gg t t gcaac t ctt gaagat cgt gact ct g gt cgt gt cac t cct gcgt at 60  
 cgcgcct ggg agcaacaaga tt agt t gtt c ct ct cat ggc tt caat ccac cgt t t ct gct 120  
 5 cccat t ct t c gaaatt t cat cggct gcact agt t t gt ggc tt ct ct agga caaaaat ccac 180  
 aact at t t t c at gct cat ac aaat gcaaag gcacggccac tt cgt acaga gct gcat caa 240  
 ct cact ct t g aagg t cgt ac t a t t ct gat t a t t gact g agat t cagaa t ctt gt t gat 300  
 10 t ctt t t act g ct at t ggt ga tccaat t t ct at t t gcaac at gtt gacat t a t t a t gaa 360

gaat gt gt ac cagaaaact a t gagt cct ct gtt cgcaca t caat aat ag at ct gaacct 420  
 ct cact at t g at gaaat caa aact gtt ctt ct cgg t cat g aggct cagat t gacaat t c 480  
 15 aggaagaagg cagt ggt t c ggt t aat gtt gct t ccacat ccact gt gtc tt ct gt gact 540  
 aat ccat ct c at gct aat t t t ggagg t t c agaat cagaa t cagat cag t a t aaaaaca 600  
 gaggacgt ag cagt at t cag t gtt acat ct gt cagaat t t ggt cat gat gt t gccaact 660  
 gct ggcacag gccct caact t cct at gtc t gct ctt at cct at gt t gg cacaat t cc 720  
 20 caccat gcct cagct t t att ccaat t t ctt t ggagct gct ct gcatt t cc ct t at ct g 780  
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<210> 145  
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 g t t t c a t a t a t a c t t t t t a a c a a c a t a a g t t a a t t t t c a t a t a g t t t t t t a t t t a a t t 120  
 t t a t a a a t t t t g a a t a a a a c c a a a a a t a t g t a a g t c g t c g t a c a t a a g a c g c g t t 180  
 40 a a a c g t c a g t a c t t a a t a a t a t a g t g t a a g a a a c t c a a c t g g g g a a g t g c a t a a 240  
 a a a a a a a a a a a a g t a t a a a t a c a a g a a a a a a t g a a g a a a g t g t g t a c t t a t g t g c t a a t 300  
 t a g c a a g a t c g t g g a a c a a a a g c c a a a t g a c t g g t a c t t t c t c g t t a a t t t c t t c a a 360  
 45 t t t t c a t t g t t c g t t a a a t a c t a g t g g c a t g t c c g t c a a a g t c a a a a g c c a c a t t g 420  
 a t g a a a t t g t g t g t a g a a t a a t t a a t a c t t g c a g a c a a t c t c c a a t t c t c c a a t 480  
 t t t t c t t t t t t c t c t a c c c a a g a g a c t t c c t t c a a c t c a g a t a c t c t t t g a t t c t c t t 540  
 50 c a g g a a a a c a t c a a c t a a t t a a a a t c t a a t t t g t c t t g a t a c t c t t g t c c g g a a t 600  
 t c a c c a c c c c c a c c t t c t c a a t t g t t g c t t c t c a c c t c t t t t t c t c a g a 660  
 t t t c a t t g g t t g a t c c t t t c t c a a t t c t t c t c g g g t t t g a t t g t t g t t t t t a t c t 720  
 g a c t t g t g t t c t a a a a t c c a t g a a c c g t a t g t g a t t c c a g t g t c t t t t c t t t t c c a 780  
 g a t t c c c a g a g a a a a a a g a a a a a a t c c t t t g t t g t g t g a g a g a c t g t a a g g a t c a a t t 840  
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<210> 146  
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 tt gaggctt t ggccgat ct ct aagaccaa acatt gttt gaat cat aat t t ctc tt gt 180  
 t ggat caagt t caat ct gca agaaacat gg agact gat cc tagt aat cgg gat gt caaga 240  
 15                 gat t gaaagt t t ct gat aat at ggt ggt gg acaaacagct ggt agat t cc aaccat gggc 300  
 aacagt t gt c at at gggat gat aat gt gg t caaagat gg gt ggt caggt aat aat t cca 360  
 20                 t gccat cat c agat cct aat at gct aagct t t caacaaa gccact t gat ggacagt aca 420  
 caaat gcat c t t ct caagag gaggt t ggtt at ggt aaaaa aatt gct ct t aat gtt gct g 480  
 acagt aacaa agcagcct ct gtt aaaagt g at t t ct ct ggt aaat cct caaat ggcac 540  
 25                 cat cat ggt t t gagcgat at ggaact t t aaaaat ggt aa gat gt t gcca at gt acaat g 600  
 cacagaaaat gact gct gct aagat aat gg accagccctt cat t gt agca aaccaat t ca 660  
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 30                 agt aat gct a gt gaaagt cc aat gcct gct t t agct gcaa at aagcat gc agact ct cag 780  
 tt at cgacac ct gct gt t ga acct gact t a ct t att at ga gaccgaagaa gcgaaaaagt 840  
 gcccacat ct g aact cat acc at ggcataaa gaact gtt ac agggt t ct ga aaggct t cga 900  
 35                 gat at caggt ggt t gccaaa act aagt gat t t aat gt gct t a t t t t cgg t gt t gct att 960  
 gt t ggt gt ag t aaaagat cc cat gt ct cca gtt gat at t g t g t g t t ca at t g t t t ga 1020  
 aagaaaaacgg t g t g t t cca tagt gt cagt at gact at t t taat at t gtt tt at 1080  
 40                 caat at at ca agt at t gtt t t cct at aac t t aaaaat t c t t act at gt g gcagt gt ggc 1140  
 agaatt agac t gggct caaa gt gcaaggcag att gat t gaa aagggt t gtt t aat aat aaaa 1200  
 t cagt ct acg cat gaat ct a t aatt ct at a att t at gagt t cact t t act ct gt at aatt 1260  
 45                 at aat t at ag gt t gaagaca gt gt ggaggt agt t gaagat tt gccagcag t ggt gaagt c 1320  
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 <212> DNA  
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55                 <400> 147

## EP 2 134 870 B1

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 5 agagtccat a agtttgttga ttacttgat a caatctaat a gagtatttt accggcccatt 180  
 ttttttctt gggctaaagt gatgtaacat ctacaactg ttgaggagat aaaacatttt 240  
 caaggagttt gatttgttggatatctagagc aatttgtaggg tttatgtatcatgatgc 300  
 10 ttcttaatca ttcaaattgt ttgtgccttt tcatgttata gctttgtgaa gaggagttac 360  
 tcaaggaaga agcgctttt a gtaaaaaaac aacttatttc cttagttt attaatgact 420  
 15 tgtatgcaga ttggacaaca cttagggat ggctacttgc ataaagaaga atttaagata 480  
 gtttatgttg ctccaatgaa ggtatgttga tgctttgtt tttcttaca tttcttattt 540  
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 20 tgcctttcc aggctttg 618

<210> 148  
 <211> 1066  
 25 <212> DNA  
 <213> Glycine max

<400> 148

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 gactggatata cctgtgacaa ggcttgccca aaacgataaa gaaaggatgtat tttggcttgc 180  
 35 tggacatgtt caccagagag ttgtgggca agaccaagca gttaatgctgttggcttgc 240  
 tggctgata tcaagagctg ggcttggaa acctcagcaa ccaactggtt cttcttggtt 300  
 ctgggtcca actgggttg gcaagactga gcttcaaag gcacttgcgtt agcaactctt 360  
 40 cgatgacgaa aatcaattgg tggaaatgtatcatgtctgaa tacatggaac aacactctgt 420  
 ttgcgggttg attggtgcac caccagggtg tggatgttgcattttcaca tttcagttt 480  
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 50 tttatcatgacc tccaaacctt gtcgcagagca tctcctcactt ggactttcag gaaaatcttc 780  
 aatgcaagtat gcccgtgata gatgtatgca agaggtatgt ctcttgacac catttggtt 840  
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 55 ttgtaacagg tggaggaggca ttttaggcca gagttgttga accggctcga tggaaatgtt 960  
 gtatttgatc ctcttccatca cgagcaacta aggaaggatca caaggttaca aatgaaggac 1020  
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<210> 149  
 <211> 1052  
 <212> DNA  
 <213> Glycine max

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 gaccgct t gt at gagat gga ggaaaaagac att gggcat taattcgta t gcgcct gga 180  
 ggaagggt at gcaact t t a ct agaat gat ttt cgaagat ttccat caga ggt t ggt t cg 240  
 15 gat gt t gaag aaat gct gat taaatgtttt cttatcccttc ccctttttag tt ggt caagc 300  
 aacacctagg gt at tttcca t cactt cagt t at cagcaac t gtagtcca att accagaa 360  
 ctgtgtt gaa ggt at t cat gat gaagatt ttttttcca gact gct cag tt gacatttt 420  
 20 tt cattgatt t cat cacat c aaaaagcctt gat accaaat tct gcat cac cact cattat 480

ttt caggttg at ct ggt cat t acgcctgtt tt cattt gga aagat cgt tt t cat ggt act 540  
 25 gct caacgtt ggtggat ttt ggt agaggt g aat aaat ttt cat gt gat ga tt ggt cacat 600  
 t gt aaatcc tt ggt tttt g tt aaaaactc t gat ct ct tg tt at aaaagg agaaat t t at 660  
 caagat gaag agaaagactt t ccaaagagaa aggaggat ga ggaat cctcc t aaacaaagg 720  
 30 aacaaaacag aaaacaacta ggaagaaaaga gat aat caga gaaacaaatc ttcccagt tg 780  
 ct cgat at aa ct ttcagt ga aat gct aaa gaaacccct tt aagcaaa tagat act ga 840  
 gcacct gat c tt at accaaa t cat gt gacg t gct aaagaa acct cttt a aaaat act ag 900  
 aacagct t gt agcat at gt a gcagattt at aaaaaaaaaat agct tttt a ct t ct gt caa 960  
 35 aacctt gaaa accaat cat c gat aattt gtt ttt gagactt aggacacacc caacatt aac 1020  
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 <213> Glycine max

45 <400> 150

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55

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 5 tttggatgtt aaat at accg ttttagttt ttcaatgatg aaaat aagat aattt gat gat 180  
 taat aggttt tacttttgg agcat agt tt atatttcta tattt gat gca tagt act tag 240  
 tagcct acca caacaat atg aggct caaa tat ggt gatt tgccat gat cc cacaat gaaa 300  
 10 t agaat gt aa cttttattt tttttaaaac at agct at ag aaagt aactt ttttttattg 360  
 aagt at gaac aaaccat tgg tt aacaat gc at at at tttt at caact aaa agt gcacaaa 420  
 tttgt acggg aagt cagt gt cagccat gct tttgaggt aa t gt aact act gagcccaa 480  
 15 gcaaattttt g aggt aat gt a cagt acacgc cattt at agt a caat gttaaa tttgct aata 540  
 ct gt attttaa tt gcat acat at gt aaaagt at gt cgat ga aat ctttgt accactt gta 600  
 ccatccgcgc ct tttt gttt gttt gaccactc attt gat gatt tacct gcat ttttattt 660  
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<210> 151  
 25 <211> 681  
 <212> DNA  
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 35 ct acagt gac gact gtt acg aacagaggac aaagct agct at gct aaact acact aat gg 180

tt acctt cgt aatttttctt tcccttctt at ttcat ttttctt g ccatattttt at aat gatttca 240  
 40 acaaaaagat a at at at ggca ttccaaatgg ccat aacaga aaggaaaat a t cctt aat aac 300  
 agatgagat gaagt tttgtt at aacagaaa ggttattttgg ggcaat aaca gaattt gat gg 360  
 agt gat ggt ggaat at cctt gaagt tttgtt g ccat gct gt tt at cctt aca ct tttt gat cat 420  
 45 agcagcgtt g ct at caacga cgcagagaga aaggggcttt gaattt aat ac tt attt cctt gg 480  
 t cat gaagag gaacgcaaaa agt at gcgaa acacaggtac t aatttccagc tt tttt cttt aac 540  
 aat aaaaaca t at gtttttga at gt cctt at tttt gttt ccacagg tggattt aaga gt ccattt aaaa 600  
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<210> 152  
 55 <211> 993  
 <212> DNA  
 <213> Glycine max

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 aaatt caccg aagtt cata a gat att gt ct tattgttatt t gat cct gaa acat gct agc 180  
 aggatt aata a aaaagaat aa aaat gtt acc agct gcact a gt at agt ttt gat cct gt ca 240  
 10 tcctttctag caat ggt tcc att cctt gaa tacact tcat ct gaat gacc aattttattc 300  
 ttggcacctt catttttttca aat ggaat ca at gtt ggt gg agct cacacc act ggaat ac 360  
 actt caccgg aat gaccaat tttattctt gAACCTTcat tctttcaat ggaat caat g 420  
 15 ttggtg gggc t cacaacact tgaat acact t caccggaaat gaccaatttt attcttggca 480  
 ccttcattttttcaat gca at caat gtt g at agagct ca caacacttga agt cagct cc 540  
 at gat ct gct cagactttgt tcccttgcata caatttgcattt cctcagt agt t gtc t ggc 600  
 20 at at ct tcat aagt agagag tttgacagaa t cgct gaaag aaact cttt aatttttggc 660  
 gttatttgggc tttcttaactt agaaacatct gattcaacca ttgacataga aaat ctttgt 720  
 at cggaccag gttggataaa aaaatttcttccctttgacc aaattttgtt agagt agt ct 780  
 25 ttggttgtcc tccatctttt cagtttctgt ctgccactgc tactttggct act ggaagag 840  
 cctttaaagg tatttaagttt caatttcatcc gtttgcctcg at gtt gaaatt tggagagacc 900  
 ctctcaagct ccagaacaga atttggagcc tgccttttcccccagatc cttgggt gga 960  
 30 tttttccccca aagactatct cttaacttgaag gaa 993

&lt;210&gt; 153

&lt;211&gt; 435

&lt;212&gt; DNA

35 &lt;213&gt; Glycine max

&lt;400&gt; 153

40 aggcatttggc agat gagaag act gat gcccaaaaagcaat t gagagt aca ccccaagt cga 60  
 caccggatc t acttcttggc atttgggatttattttaaaga ct caccttttta gtttacaccaa 120  
 45 gttttaactcc agaaaaacca caaaaagatc taaaaaatgat at cat gagc ctctttgaga 180  
 aggtatgtgc cagttgttca at aggtttgt ttaaggctga gttacttctt t gatttata 240  
 tattatatat tggttagaaa tgcctttttaa aatatacaca ttctatattt gttgacatttcc 300  
 50 ct.ccttgcctt gatgtgat tttatccaag acaccaaaac aagtgaattt agttgtcgat 360  
 cgatctctat ccttagatgg gttttatgt tttggatgt gaat aagatt ttacctgacc 420  
 cagtaaattt gacat 435

55 &lt;210&gt; 154

&lt;211&gt; 362

&lt;212&gt; DNA

&lt;213&gt; Glycine max

&lt;400&gt; 154

5           aat acaatt a tt caat gaca at at gct ct a ttt at aaaag aaatt gagcc act acact ag     60  
 ccact aact c ct aggt gcct aggaaaacaa t at ccagcaa taacat aatt t att caaat a     120  
 ccccacat ca cct aat aaca at at caat aa cagaaact t a aaaccaat t a aat gaccac     180  
 gt cacct aac att cct t ccc gt aaact gaa t gat caat at t cagtttaaa caacat aagc     240  
 10          agt agaat at t at ct ct gaa act aatt att caaaact gcc cacaccaagc aattttt gta     300  
 gct t ct gaaa tacaggt gct tt gagaggt t tagt aagt at at cagcaact t gat ct t ac     360  
 tt   362

15

&lt;210&gt; 155

&lt;211&gt; 652

&lt;212&gt; DNA

&lt;213&gt; Glycine max

20

&lt;400&gt; 155

25          at t gt gatt t t cact ggt tt gt gagagt gc aaaagaatt g tt cagtt gaa t gt gcaaaat     60  
 tgct t ggat c agt t gaaat g cacct at gaa t tt gt at ttt t cttttt at gacaaaggc     120  
 at gt agaat a t gatt at att tt gt tt gaat agt gt ggggg agcatt act g tttttttt     180  
 ttt gaaaaaa aaaat ct gat gt ggt agt gg t gt ct gatt c acat gt ggaa aattt ct t at g     240  
 30          gatt t gggaaa gaat at t gat t gttt cctt t ct cacagt g ct ggt ggt ga aagcagt gga     300  
 tt cct t gcat t cagagt t ca gggct gt gga t aattt ggt t gt gt gcaat a ccaaccgt gt     360  
 cct taaagct tt ccagaat g ct cgagt t gg at cccat gt a agcatt cccc tt gat tata     420  
 35          taacctttat gcaa at gt ac attt aat at g at gct caat g ct caagggtt caaggct aat     480  
 aaact t gtt a act gt tttt gat t gtaattt gg tagagat gt c ct t aagcca tt gggct gat     540  
 ct t gat gcct tt at gt at t t gacattttt accaaaaaca taact aat at aggaacccaa     600  
 40          aaact t tagga tt cgattt agg gagaacctaa ggct gcccat taaaact t gat gc     652

40

&lt;210&gt; 156

&lt;211&gt; 1180

&lt;212&gt; DNA

45

&lt;213&gt; Glycine max

&lt;400&gt; 156

50

55

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aaaat aaaat cgagtggt aacccagaa ttatggtgc cagttagt ggcttgta 60  
 gcctggctgg ggttagtgtt gctcaaaaa aaccatctgg atatttcaac caaaaattt ag 120  
 5 tagcatgaca caaaaatgc tctaacaaag gcaaaaatgtt ggttagtgat a gttacatgca 180  
 aatgccggag aaactaacca aacaagagcc aagt aagaag agccatttt aaaaatagttt 240  
 cccaaaatga gaagtat aag ccattgaaag gatccagctt tatagagcca tctttcagcc 300  
 10 tccattttga caacagctgc tttaatatg ggggtcaact gccttcctt ttagttagtga 360  
 ttcgaaagat atttttgaag tagaagaaaa cccaaatgtt gaccagtcaa tgctccgaga 420  
 aaagctggtg caccataa tccaaaccacc aacattggc ttgaaacata tggtaactgga 480  
 15 gatgaagat a tcaaaggtag gagaatgaa acaaggaagg agaaaactaa ggcaaagacc 540  
 cacatgagga gaacactcaa acacgacaag gccaatgaag ctgcagtgac tcaaggacaa 600  
 aaacattatg tttagtgctt gacaaacact tcaatgcagt tcaactaatc ataatat 660  
 20 atcaataatc aatgaagagg ggttatatct ttttctcaat aactcaatcc atcaatat 720  
 aatgatctt ctaaaccact gtcatcaac tcccatatca t cagcgcgtc accaaatcat 780  
 atgat aagaa aaggtttac tgcgtcaac cattcatatg at aagaaaag ggtttactg 840  
 25 ctgccaacca tactgttgt tgccgttacc acccatcatg tttgatccac gcccagctgc 900  
 cgatccacca taaggagcac ctgaggat a atttggagct ccaactccat agccgctcc 960  
 acccccacca cgaggaccac cactgggccc accgggcat a ccacttgacc cat aaggtgc 1020  
 30 accccgcct t gaggaggat at ggcact tccataacca ctccctccct cttgacctgc 1080  
 acgattcggg t gattgtatc cccaccaga gttcccacca gaaggatatc ttccctggccc 1140  
 accagaagga cctcgaggct ttccatatg at ggctaggt 1180  
 35 <210> 157  
 <211> 628  
 <212> DNA  
 <213> Glycine max  
 40 <400> 157  
  
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 acgggtggaa atttttctga ttgtcttaat ttaagat taa aat acaaaaaa tacaatgct 120  
 45 gaattctctt gaaaaaaaaa tacaatact gaattgtagc aaatcaaact ttttttcta 180  
 cat aaaaaaaaaa aacattttt tccctaaaaa tgcctttgt ggttgaagat ggttaacaac 240  
 catttat t cagttatgt attcaatag taaatagtaa tattcat tta acctaataat 300  
 50 attcatat a a tcaaaacttt acacaagat a ct agat taaa atctagtgtg atcat t gta 360  
 ataaaaagaa t aat cgaagc attacactat tttctgtcaa aaaagaaaac aattgaaccg 420  
 tttcgagcaa atcaa atcat caacatcat a tcaagttt aatcaaagt a gatctttct 480  
 55

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cgt at cat gt gat tttttt a t gt gt aaaaa t at gt caaat t aagacaatt tttttt aaga 540  
ccct aaat ca at aaaaaaaaaa tt at cgaat c gt gt t gggt c aaat tt at tt at t aggaaaa 600  
5 aatt caattt aact t aaatt acccaaat 628

<210> 158

<211> 774

<212> DNA

10 <213> Glycine max

<400> 158

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aacagaat gc tacaacat tt caagaggacc caaacgt aga tt at aaggag aat aat gaat 120  
cct cct at tt aaaacagaaaa gaaact cct a tcct at ct aa aacagaaaaga acccaat gag 180  
20 ccaaagt ggc t caaaaat gc aat aaagt at tccaat at tt t cgcatacaa at gatt gat t 240  
ctt gaagca gccatt aacc aagaaccat c at agagacaa tcct at cct a t gacgact gt 300  
aaagggaaaag aggt gct tt gaaaat acac gcattt cat t acaaccaa at gcaact ag 360  
25 at aact acat at act gcaca at gcgat aaa at t aacact ctttgtt cct tt caaaacct 420  
ttaaggcat g taaagagaaaa agct ccaacc t at gat t gga gaaact catt gt tggtt agg 480  
aaccggaaaaa caatt cagca ggt gt accac aaaagt ggcc tacct at agt at t at cagct 540  
30 t at ttt agca t gttt at acc t agat gt ct c tatttc tta tgaact tcaa t agtt caact 600  
accat tt gat gaat gt gt cc at gat cat at cat aact t at at cacgcaaa ct t cagaggt 660  
tattt at ctt tttgtt ctc at t gt at t ct acaccaat ga ggt aaaacaa gcgagcccc 720  
35 aacgcat gat gaaacat aat catccattgt t gct act t gt cagat cacct ct t g 774

<210> 159

<211> 637

<212> DNA

40 <213> Glycine max

<400> 159

45

50

55

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ctt agt gagc t at gt ct aac tt caat gct t at att gcaag cct cgt ct aa ct t cacaaa 120  
5 tt gat t gct a aatt gt tt ac agt at aagt a tgacaaaatt gct t ct gtt t at gagat ac 180  
gt cccccccc cccccccct act cat t att at aat gaagg gaacagct ga aaat aatt t a 240  
tagt aaggaa att agtt gat tttttttt acat tt gttt gt t gt cgact gcaaccgaga 300  
10 aat gacaat a att gt gt cct t gt t ggcaag gact t ct ttt ct ggcagct g gcagagt t gc 360  
agat gaaggt t cgg t ct gat gt gagtt ct g aat act ggct caat gct aag t gt gcat at c 420  
ct gacaggca att gt tt gat t gggcat ga t gaggt t gcg ccgt ccat t g t at ggt gt t g 480  
gagat ccatt t gccat ggat gct gat gat c aatt aaagaa gaaacgggag gct gaggt aa 540  
15 cttt ct tttc tt ct t cagta at at gt at t c cccct ct ccc cttt gt ggg ttt gaact t g 600  
ctt tccat at cat ggat at c at agact at a gtt acat 637  
20

<210> 160

<211> 1040

<212> DNA

25 <213> Glycine max

<220>

<221> unsure

<222> (1)..(1040)

30 <223> unsure at all n locations; n = a, t, c, or g

<400> 160

35

40

45

50

55

## EP 2 134 870 B1

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 cctcaatcccttgcatttc tcttcagccttttccctggtc caaagtccatacttctgtg 120  
 tgaaaatcgattttagtgc agatgtgg tcatccctc cacaagggtctgcctagtgt 180  
 tctgactagg aggccataat ttgattgaaa atggctcggt ctgtgaattc agtccatgg 240  
 caatgaaacc tacaagaact gcaagtgc aaatgtatataat ggaatattt aggatacata 300  
 acaagagtaa aaaatagttc aagcagactt gttagagaaa acaattgtaa ggcaatgtca 360  
 ttcatagatg catgtcccaat atatggtatc agcccttgtc agacagaata aaacatttta 420  
 caagccctta acacaaattt gagtttgaa tggataaaaaaaatgtttt acaaaacagg 480  
 tttagttataa ggagcagaag taaaagaaat cccatccaag aaataatgc tataacatat 540  
 aaacatat atat acacccag gcccnncnnc ncaannncanc acacacacac acacacacac 600  
 acacatat atat atacac acacacgtaa agtggattgc aatataaag tggggcatg 660  
 ttcccttcca tttcaatcat atcatgttaa tggaaactaat aaaaacttca aagcatgatg 720  
 aaaaaatgaa aagggttagg gagtttataa aggaaacttt gcaaaacata ccatgaatgt 780  
 acgtacgagg ttgtttccctc tgaagagaag agaacactca caaaaccgtc gacagtataa 840  
 aataaccgac aatcaactta gttggttca cccacaaatc aaacccatcc taaataaaaat 900  
 cttagctta agtctgaatg caacagatcc gagactgcgc aaagatacaa actttaaaca 960  
 cgaatttcaa aatctttgaa gaaaggggaa ggaactggaa acaggtaga ctgaaataaa 1020  
 gagagacaag tttgacaacc 1040

&lt;210&gt; 161

&lt;211&gt; 845

&lt;212&gt; DNA

35 &lt;213&gt; Glycine max

&lt;400&gt; 161

tgcattgcctg cagcttgcgtg ccaaactttg ctacatttgg tatgatcag acagaagaga 60  
 atcatgttag ctctgtact atgtagaatt gtatggttat tagtgggttt tgcataagat 120  
 taacagtgaa ctgcaaaatg gagtttttag agggatgcattgtatataat atttacgtaa 180  
 caagtgtggc ttcccagttt tccatgcattgtatcagc tttaagtgaca tggcgaaa 240  
 acaatactgatttcatgcta atgatcagat tttccctcgtg cagtggacaa tgcataaaag 300

50

55

act gat ccca aagct caggc ctt gaagact gct at ggct g t gat t gt gaa at caggat gc 360  
 caagt gtt a agaaaaaaac t aaat ggacc aggacatt ac t gt gacat cg att acat tt g 420  
 5 gcaggccaaa gacat t gct g t aaat tt gaa act gt t t ggg at at t gct gt aaat tt gga 480  
 ct gat at t ga acgt t t gatt ct gaaact gt t t ggacaga t at t gct gt a aatt t cggcc 540  
 ttt gagggaaa aat gt t t gt t gct gcaact gttttt gat t ct gaaat at t gtt gat at t 600  
 10 gct gt aaatt ct t aaact gt gtt gt ggct t gtt acgt t g t gt at t gat c aggt t t gaga 660  
 aaaaaacata at gaat caaa gaaat t t gtc aat acat gcc aaaaacat tt gcgaat gcag 720  
 taagt ct ggg t aaat cat gg t t cat aacc accacgt t ag t aact gt gt a aat ggcaggg 780  
 15 act agaacac act aaat tt g ttt t gt acaa ggat t aaaaa ct t acaaggg gt caaaaat t 840  
 ct aaa 845

<210> 162  
 20 <211> 631  
 <212> DNA  
 <213> Glycine max

<400> 162  
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 tt gt t t att a agggtaat ga gagtt cgaga t gtt agt gtt att gt t gt t g t taaaat gag 120  
 30 ttttttataa ct taaaat ga gt ct taaaat gaat ttttgc at aagt taaa aat ct tttt aa 180  
 aaat ct at ga gagttttt at aaatttattt attt agct t a ttttcat tttt atttttttc 240  
 tcttaaaagt gttt agaat aaatttcat tcaat aagt aa ttatttattt gttcgat at t 300  
 gttattt gtt a ttat cat att ttgtttttt ttttggaaag ttgaat at ca taaaact gat t 360  
 taaaagaga ggcctt ggtt gt aaaaaacc at aaactt ac gt cat aggt g tgcgaat at g 420  
 at aact aaaa aactt t c gag gagtgacttt tgacggtaaa at tggaaag aaaacaacat 480  
 40 act agagaaa tt cat caaca tactt at ct tatttataat tt cactttgt tacaat acat 540  
 tgggtttttt ttat aatttt tattttttt ttat cgaat c tttcaatttt at gt gccc tt 600  
 tactgtttt ac cat aaaaat at cccccct ac t 631

<210> 163  
 45 <211> 439  
 <212> DNA  
 <213> Glycine max

<400> 163

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5 tcactaggt aaaaaat tta gaaaaaaaaa ctgataaaaa cagcaaaggg acacaattca 180  
at t gaaat at ggaaactgt a acacattat t aagt t cat a cttatttgc aagt ttgaca 240  
aat t gaat at ggagagct ag cat agaaatg at atcat tca ttaaaataga aat aaataaa 300  
10 taaat agcaa ggacaagt tg at cttaaat tttaaacagt acaacaat aa gcactaagt a 360  
gaccaagaca ccat gat agc gat aat atca ccagt t caga attagagt at cagt cat tga 420  
at at gaaaaa tgaat gtca 439

15 <210> 164  
<211> 543  
<212> DNA  
20 <213> Glycine max  
<400> 164

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ctcaagaaga agat aagagg aaagcaaggg cacttaggtg ggttcatcat tcccttcat a 120  
at tttgtttct cccaaattcat attgtttaca agaatcaagc at gcat ggtg tcttatttagt 180  
30 tatttaggtct ttctgttttg ggttcccagg tttgcgaatc cgtcgtcaac ttctatagct 240  
aatgttaatg gcgaggcaaa gat t gagccg gt aagaccct tttggtcaact ttcaatgctt 300  
tgcgtcatac aaagat gaaa aaaaaatgt a tttttgtgtt gactgtt gtt ctgtt gttt 360  
tcaaactaga aggctgctat tgcaggcaat gctggaggag ggacctgaat gacaggtcgc 420  
35 gtcttaatt tgtaggaatt tttcttgtaa gtcaattatc tttgcctt gcttattacc 480  
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ttt 543

40 <210> 165  
<211> 369  
<212> DNA  
45 <213> Glycine max  
<400> 165

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55

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	tatttaattt cttttattta atggtcggcc tatataataaa ttgtagtaga gcttctcaa	120
5	ctatgcctt tactacagag gaaaatcttc aaaaggaacc tcgtatcatg gaactttagga	180
	atcaagt aag aatacaatcc tatgattagt atgctttt cttttcaatt tattgtctgac	240
	tactgacttt ttcttcttcc tcccatggaa acagtgtaga ataatcgga caactgagtt	300
10	agctgctgct aaggagaaaac taaatgagct tgagaagcag aaagaagaca t gttgaaat	360
	gaattccgc	369
	<210> 166	
15	<211> 821	
	<212> DNA	
	<213> Glycine max	
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20	<221> unsure	
	<222> (1)..(821)	
	<223> unsure at all n locations; n = a, t, c, or g	
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30	ctaatgctaa attttaggg tttatatc cacattcttc gggttcgggc aagaagagag	180
	agaagacgca attttgaccc tcaatttac agaccttta tcatcacgc gattcttctt	240
35	ggttaactccg ccttcctcaa tccataatct gctttataa ttatattt tagttttat	300
	tttgattttg gcttgctgca agctaatttac cgcttttcag ttcaatattc cccgttatt	360
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40	aactgcacct tttcccttg tattttttt gaaatgctt cttagtttta atgactatac	480
	tgagcttttta cttcagttt ttctgtttt ctgcacgcct tttatgtat tttcacctt	540
	tgaggtctct ttgaaatttt tttatgtc gatttgtgca atgattatg gcaaaaaatc	600
45	aaaaacaaaaa atgacttcaa ttccatgtt ttccgcgtt gcttattaga gagagt agga	660
	aaggtaaga gggctgaaa atgaaatgca t gaaatgat tttatgtg atggatgt	720
	gaacanagca ttatgcttcc tcttacttg ggaggaatga acattattt tggaaaactc	780
50	atattagaat aacctgcccc taatttacac ttttttggga g	821
	<210> 167	
	<211> 848	
	<212> DNA	
55	<213> Glycine max	
	<400> 167	

EP 2 134 870 B1

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5	t act gt ccat tt ggt tt gt g gcat ct aggcc at ct ggct ct t ct gct gt t a at gt t agt cc	120
	t gt caact gt aaagt agaaa ggagct cacc agt caggcct t ct ccaattc t t gt at gaat	180
	tt gt ccct gc aaact ct gt a t gcct gatt a t gat att agc at gat gact a t gat at act a	240
10	gt cat agct t aacaatt aga aatt aat att t aagt att t aact aat gct t cattttttt	300
	taat att t ac acaagcat gt t ggaccagct t gtt attttc ctaatttcc t ggt at cct	360
	taattccact at gacat acc ctt gcat acc gt ggaggact taacat ctt t ggacat ctt	420
15	tattatttga t gtcgtatt tctt gtaa attatttgg ttaatttaatt ttttgaattt	480
	actttatagt cat gaattt a tact gcat ct taaatctgt t cacaactca ttgattggtt	540
	at gccgttt caacgaagaa t ggagt t gat cccat gggc gaaat gt gga gaaacctaga	600
20	actgt ggaag at ggaat aga taaagctaaa ccat ggcagc t gtcgaaat t gt ggt gct	660
	gt ccaat gt c ggttagtac aacgcctgac agt acagatt cttccagcaa ggtttgttga	720
	ttttatgaaa ttgagat tgc cattttctat t gtaactgt gtactgt gaa ct gcttaac	780
25	tctaaagtca t acat agggt t gttcgactt ttatatacaa actct ggtgc t ggtctttt g	840
	gcacttgg	848

<210> 168

<211> 825

<212> DNA

30 <213> Glycine max

<400> 168

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	tttaaatttg aatcgcaatt ttgaaaggta atgctgagca aacaaaagaa atacctttat	120
	ttctccattt ttccaatttt aattattttt catattcaat accttattgt attttttctg	180
40	agtttttctc acaaatttgg actgcaagt gtttgttagt tggttggaa tattttttt	240
	tcatggacct agttttgtt ggtgcagaga gattggct aatattgaatt ttttttattg	300
	gttaattttgt attgttattt aactattcag agatttattt gttaaaattt gtatgttgtt	360
45	cattatcattt ggttgaagtt tactatttag tagttttttt aatattaaac tacaggttct	420
	gtaaatggag gaaattacaa agaattggca gcatgaccag caaatagatc atcatggta	480
	attaaatcatg ttgatgggt a cttatattt gactcaccaa atttttgc	540
50	acaggtaaat t gatctatca taaaaaaagat ttgcaggtaa gagtcacgc atctccgtt	600
	tggtccttga gagagacatg gtttcatcct ccttgaataattttagacc cttaattttt	660
	tgttaattttt cgtttttttagt gtttttttgcatttttactc attttgtctcc attttactga	720
55	ttaactaataa attttttttt cttattcaag gtacacittt attttttat gatagattag	780
	acaatttaaa ct agtctaat gcctaaccatt ggctgcaggt cgact	825

<210> 169

<211> 953  
 <212> DNA  
 <213> Glycine max

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<212> DNA

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<400> 171

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	gcaaccaact cctt caca t c acat aact aa aaaaat aat ac tcat gt aacc aaaaaat agt	660
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	ctttttgtt ggat gagaga gaaagagaaa aaaaatt aatt ttctttct agactt at ct	240
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40	aagaagagca attcatat aa aaaaagcattt gatat agcaa acagaat at c ttccctt gata	420
	cct ggt aat a tgcacattag aagt agt aaa tacactt ac at accttctg acggctctgg	480
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	aaat cagt tg gatt caccaa ct tggctt acagcat gt a agggct ggaa aattt gaaga	300
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	gat agat gt t gt gagggagt tagt acacat gact aaagaa agt ct t gat g gaggaat t at	480
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	aat cat tttt c aat acagagg tttttt at g ggt at at at a tagt t gat ga aaat ct t cgc	840
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	ggtt gaagt a actt at gggc tt cct gaaat t cgacaagaa tggat t tca t ggaagaat a	240
40	t gcaaaaact at cagcacca agt agaacaat at aaaacaat at t agat gaa t cat ccacaa	300
	cat at t at gc agat gaat at ttt acat att t gct aat at a aat caaat gt caaat at t ac	360
	at ct at gaaa gt t ggt at cc tttccat tttt cat cact aga tacaat gggc ct t gcaat a	420
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t cct caacaa aagaggt agc agt at ct t ct ct t ccattt aa tt aat gt t t c acacact aat 420  
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10	acaaaagataatatggca ttccaaatgg ccataacaga aaggaaaataccctaataac	300
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	agcagcgttg ctatcaacga cgcaaagaga aaggggcttt gaattaatac ttattccctgg	480
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	aataaaaaca tatgttttga atgtccattat tggatttaga gtccattaaa	600
	agtggttcc caacacatga tggagaacac cctatatttc ataaagatac taccattagg	660
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	ctataaatgtttttaaat taaatccatagtatgatctgtgc atttatggatgttttacgat	780
55	tatcttttgg tggggtgcaat gatagtatatgttttgcataatgttgaat aacat	840
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 <211> 863  
 <212> DNA  
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5    <400> 185

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15	gacatggatg	atattggta	gaggtgaggc	tttggctg	ataatatgtg	atggtaaaca	240
	ttgtcttacc	attattgaca	ttatattaa	tatgacaatt	tagttgtca	tggacaaatg	300
20	gatattgtat	cataatcaga	cttatgtaa	tgcaggatga	tgaagaagga	ataactcatt	360
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25	attractcc	ccaaccacct	tcaagtgtat	tgtctacttc	atttatgca	gacttacac	480
	catctagagg	gaccacacca	gttaccaca	ctttgggac	cccttctagg	aatagaattc	540
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	aagatgttat	tatgccta	aaatctgcacatt	gcactcctt	tctctcagag	gctaactctg	720
	cctgtatag	tgaaaggacc	atgagcatga	gcgaggatcg	ttcatccatt	agagagaaaat	780
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35    <210> 186  
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40    <400> 186

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	tatagatatact	acactaatac	tggagcttga	ttatattac	tatataaaa	gcacttggtt	180
	aatttagaat	attttacaaa	atttgttctt	cttttcctcc	ttttccctt	taatttctac	240
50	aaaacaactt	cactgaacct	gcccaattgg	agggtgctgc	taatagtaa	ttagattatga	300
	attgtttgtat	aaaaggcatg	gatgtacca	tcaaacttgc	tctactttat	tgcagtatgg	360
	tttgtgaagg	aagtgaatgc	ggtttcaatt	tgaacttttt	ttggctgtag	actggatcct	420
55	tcttcttca	ttctgtttta	tgtgactagt	atttgtttt	ctattgcatt	tgtcaaaaaaa	480
	atcagttttt	ctatgtttac	cattattatg	caggtgctat	tgtcatgcag	gagatattga	540
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<210> 187  
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<212> DNA  
<213> Glycine max

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	t ggct ggcag	aatt acct t a	at caat gct g	t ct t gacagc	ttt gccct t g	ttt t at at gt	180
15	ct ttt t cag	ggcccct t ca	gcagt cat ca	agaggct cac	t act at ccaa	agacaat t t c	240
	t t t ggggt gg	aaact t ggaa	gaaaaaaaga	t agct t ggat	ct cat ggcag	caagt gt gt g	300
20	ct cct agaga	aaagggaggg	t t gggat ca	aagat at caa	ggct t t t aat	agagct ct t c	360
	t cat caagt g	gaaat ggt t g	at gt t ccagc	aaccagat ca	t ct at ggagc	agaat cct ca	420
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	ct t ggt ggt c	t gact t aaga	t caatt at t c	aacat agt ag	cat ggct gct	gt t aat aagc	540
30	agt t t ct t t g	gaaact gggc	aggggt gat c	aaat t t t att	t t gggaaagac	t cat ggg t gg	600
	gagat ggaac t	at t ct t aga	gacaaat act	cagaatt at a	t ccaa at at ca	t ct caaaaac	660
35	t acagacagt	ggcaagcat g	gggatt t t t g	gagaaact gg	ct gggagt gg	aaat t ct cct	720
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40	caggcat gca g						791

<210> 188  
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<212> DNA  
<213> Glycine max

<400> 188

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45

50

55

## EP 2 134 870 B1

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 5 aagagt tt a act agat gct gat gaacagc accaacagcc aggagggagc t cacagat  
 gcgt tt ctt gagaaggct a t cacagat ac aaat aaatt a gat ggat gt t cact gt aaaa 180  
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 10 t ggt gt agcc tt cat t aatt gcat cat gt g cct ct gcaca cat cct gt cc aaggct t cgt 360  
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 ccct at aatt cat ttttta at ggct tcca tt tctt cagt ggat aaaagg ggacct ttt a 480  
 15 gt gaaaggcg gt gacat tgc t cct cagt ga tt tctgt aag at cacc t ct ggaccaacca 540  
 t acatt gcat agaagt gact att ttc tct c taat aggat c aat aggaggg tttgt cact t 600  
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 20 at ggagt at c att tccatt gacccaaggg ct t ct acacc at cct t gccc at aggaagt a 720  
 at agcat ttc caat gatt ca act gt at at c t gcat gaat a aagcat aat a t aaaaat at t 780  
 t cct t cat aa at gcagcaga taaaat t gag gaaat att aa t gat ggt cat t gcat accca 840  
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30 <210> 189  
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 <212> DNA  
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35 <400> 189

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 ccaa at ca t cat gcccaa gaagccacca t gccaagt tt tagcagcat a acacat ct ag 240  
 t cccgt aaag gggggagaaaa aagt aaagaa tt agt aaatt cat t ct at gc aact agat gc 300  
 45 at at aacccc t at t gt agt c ct tgc tttt agt tttt at c ct at tttaca at ggct aagt 360  
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 <223> unsure at all n locations; n = a, t, c, or g

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&lt;400&gt; 190

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	t t cagct aa cat gttttg taggt gt aaa tt gt catt at t cttt at ct tt gt aagggg	180
	t at cat agca aaat acagaa t acat agt gc tgct tgc t c t c t act t t gat gagt	240
15	t cct gct gc t ggt act ggc att att aaca t aaat ct agt gtt cttttt tt tttt at	300
	t t tttt aacaa act acagagt aact t gact a t gaaatt ct gc gt aagaagat t at gat gat a	360
	cat aaact aa ct aaaagt ct gaaat aacaa aaat gaacca gt t gccat t g gat cat cacc	420
20	t ccaaggcaca agaggt aaat a agaact t gat t cat ccaacc aagacacaga gccccat ct c	480
	t ct cct ct ag agt gt aat gt cct cggt agc t t cgct gaag at tttttt att gaacaagt aa	540
	t aaacgagtt cagtggt at ggt gcgaat c cagccat agt aaaacgcgt ct ccact t ct	600
25	t cagaagct c gt gacgt t cc act ct t cct gccct t caca t gct att aag tt gacaact t	660
	ct cgagccaa acaat gct gc t ccacatt aa t cctt t cttt gt gct ccct c ggcagagcaa	720
	cat caattt ga tt canaaat a gccaagt agt agt t cat cgt ct caacgaaa cggggat aa	780
30	t gggannngt gt ggt at gt g att t ct t gct c aact agt gt c acaat c	826

<210> 191  
 <211> 969  
 <212> DNA  
 <213> Glycine max

35

&lt;400&gt; 191

40

45

50

55

## EP 2 134 870 B1

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 5 tt cattt gta aat at tttt ggct gat gga aatt cat gt g att gt catt a aact t cttt c 180  
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 10 t gtt cat att gt ct t aaatt gt tttttt g tagct gaagg agt ct gaaga aaagat t ggg 300  
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 aat at gt ggc gt t ct gaact ggcaaaagct agagagcat g at gt gat ct t agaagcagct 420  
 15 gt agt aagag cagaagaaaa ggt t agggt t gcagaagcaa at gct gaaac t aggat aagg 480  
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 aat gt act aa aagcccaact t caaaggct t agcgt ct t at tt tttt t cttt gctttt 600  
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 aaaaat at at at caat acct agt gaaacag ggaaat ggaa ggagact tttt gat ggt t at 720  
 20 tt gt ct tttt accagt t t at t gat t t gaa t at gt at at c agct acgaaa t gt ggagct t 780  
 cat aaaacca aagt t gacat agcagat att tt tttt t ca acaggcaaca catt gat aca 840  
 act caagt tt tt gagaagac agagt cat gc t cagat acaa agcat gt t ga ccccact gaa 900  
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 25 aagct ggcg 969

30 <210> 192  
 <211> 1269  
 <212> DNA  
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35 <400> 192

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 tt ggat ccag caccat t ca agat gaat at gcaccagt ga t attt gct ca agaaacagt t 240  
 45 t cccat at t g t at ct at t ga cat gat gt ca gggaggt gt gt ct t gct t a gct t gct gt c 300  
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 at cact aaac agt at at aat tt act tt gaa cagt gaaaat gt gaaagt t a aat gt ggggt 600  
 aaacagaat c aact acat ac aagagttt a ct cat gaaca t caat aaaat gact ct gagc 660  
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 55 ct cat t at ct gt ct ggt t aa aagaaaat at tt t gtt gcat t gtt gcacag gaggaccat g 780

## EP 2 134 870 B1

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 5 t agat gct ac accggagcag cgt accgaag ct act gt ggg gt aat cct ac at t t aact at 960  
 ct act ggt t a aaat at gcat t t cat t t gt g ct ct gat cca cct cccct aa gaagaaaaca 1020  
 at t acaaagt gt aat gt gag t gt t gt t acc t at t t at t gg caggt at ct g ggt gcat ct t 1080  
 10 at gat gt t cc at cact ggt g gacaagct t c tgcaccaact t gccagt aaa caaaccat t g 1140  
 tt gt aaat gt t t at gacaca act aat gcat ct gaccaat cacaat gt at ggt acagat g 1200  
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 15 at agat gca 1269

<210> 193  
 <211> 1246  
 <212> DNA  
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<400> 193

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 30 aaaaagacat aaat aaaagg aaat t at cca aacaccaa ac ct agt t t caa ggt t aaat t g 240  
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 cagt aaggaa at gt t t gatt t gatt at t t t at t t cat t t t act gaa aacgaaaaat 480  
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 40 aaat aat cag aaaat gat aa cagaaacct c at t t cggat a aat aaaat t acggt aacaa 600  
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 t t t cagt at t t t t at t t ca t gaaaacaaa aaacaagaag t t aaacaaa cat gt t t ca 720  
 45 gaatt ct t t c tttt gaaaat gaaaacaatt t t caaaaaat aaaaaaaaaa t gaaaat aca 780  
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 acagt t t att gat aact gaa accaat gt t t acaagg t t g accagagaaa gct agct ct a 1020  
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 55 t ct ccact at gaagaacct t t t ct t at t c tccacaagga acgct ccacc t ggc当地 1140  
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<210> 194  
 <211> 671  
 <212> DNA  
 <213> Glycine max

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	aat cct ct at ggt aaat gca ct t caagat g ccaat aat ag t gaaaaagt g gagcct agaa	180
	ct agt ggcaa gaaagggatt gt at gcggag tt gaagt agc aat gt ct aaa gaaact at t g	240
15	aat gt cagaa ggaagat aaa acgaaggt at t ct t ct aat g tt at t gt t aa tttttt ct	300
	t gat gt t gt a tt att gt t ct taact t gt ga at t gt gat ca cg t t aagct t cat t t t at t t	360
	t catt act tt caat ttt ct t agt at gt acc accaat gat g aatt aaagat ttt gat ccaa	420
20	t gat gat cct ttt gat at at tt agt tt agt aagt t caaat at t t tagct agcatt gcaa	480
	t gt t t acac agt t acccac cct t t cccgt aaccaat aaa aaaaagt gt g at cacat t gt	540
	t gt act t gt g t at gt ct at a t ccaaagt ct tt at at ggac acct t aatt a t gt ggggt t c	600
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**EP 2 134 870 B1**

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gtt ccaggc at gtt aat ga tct gaatt t g aacaaat caa gggccat aac tt agccagt a 180  
agt acaagt c tagaagaaag acagatt aga t gt ct ggat c caggt gt ac t agt agt t ac 240  
agcaat aaat gcaagat ttt ccttaact ac tcat gat aca gct ccattt gaagccaagg 300  
10 gcact t gt at accaaat gt a aat ggat gaa act aaagt aa ct agt aat aa acacagat t a 360  
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15 tt aaaaat ct a acaacagt ac aagacat gga cgctttatcc ct cct aat at tctttt aat 540  
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20 taagat taaa agggctt t ct ggct gttt a acaaaccatt tttctgcat tgaat aacag 720  
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tcc t a g a a c a gccc t a a a a agaaaaagag aaccat gcaa aaccaacaaa gaacat ttt a 840  
25 aaaat aaagt caaccatt at t gat gaaa tattcttt a caaatt aat a agt cat t t g a 900  
  
gagt t gagac act aaat tca cact cagt aa t t ct at tttt tttt agataa caagaggaag 960  
30 agaagat tga cat atttaca tt ct t acat g aaat act t ac at t gtt ct at aaagaat ttt 1020  
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<211> 694  
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<213> Glycine max  
  
40 <400> 196

45

50

55

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	caatctcatt cattccatgg ctttcagttt ttccacttt taatttttaatctctgtgag	120
	gcaatttaat cctgctgcat tcacatcagt tcttgtatat tctatgtaa ccaacctgtg	180
	ctgataattt gaaagaaacc aactgatcag aagtctttatgccataagt aaaacattgt	240
10	ttaaccagtgt ttacagggtt ccacatttt ttttccaac gttcatttgt ttgcaggcag	300
	aggaaaaagat gcgtgttatg catgatagga agtgtcgcaa gctgaagcgt ttggatgat	360
	gggggtgctgattttcataaa gttgattcaa ctcgaaacttt ggtaggaat ctgtccacaa	420
	aaattagaat ggcaattcag gtggttgat aagattctat gactataacaaagat aaggg	480
15	atgaagaact gtggccacag ctgaaggaat taatccaggg gtatgtgatgtt aaaaacta	540
	accatcttg ttatgttc aagtctaaa tgcctctctgtt aatgatgg gcaacactgt	600
	atgt aaggat ggatggaaaa ataatcccatatctatgct ccataatcca atccttcaat	660
20	tcccccaat ccattgaata ttgaatttttggaaaa	694

<210> 197

<211> 693

<212> DNA

25 <213> Glycine max

<400> 197

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	ttatccaat atagaatcat cgttaatgtg ggttggttga ctggttcaa ttggtagcct	180
	taactaaatg gtccaaagt acaaaacccct ggcagaggcc tggcatgc attgagcttg	240
35	cttggaaatgg agtaacctct cctccccaa acttctctca caaaaaattaaagaaat	300
	gaatcttcag gagttctagg acaacccttc catcatgcta aaaaactat ttttgaat	360
	ttgttcaaaa tagtatacc aataccat gaagctactc aggacagtgg taaaagtat	420
40	ctaaatacaatgttggaaatg cagataaagt cagagaggtg ctgaagcttg atctggagat	480
	gaaggatcta gcaaaggact tggatgctga gcagtcttttctgtcttggagaggata	540
	caactatgca acttgcctt gagggagctt tgaaagtaaa ggaagtggct ctatgcata	600
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50	<210> 198	
	<211> 738	
	<212> DNA	
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55	<400> 198	

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 5 t gaat t caac tttt acat ga ggagt cagag ccaat gcccg tttt at agact gagtt gcat a 180  
 cagt t acagc acct at gact acttt at t gt tt aaaaat tt agt ct cct ct agct caacct 240  
 aaat aat at c ct caat at aa agat gacatt gacagt aaat acaaaaagag accaaacaga 300  
 10 aaagt caaga tt gcat acgt gat at act ct aaaaagggt tg acaacggtt c t gt ggct t gg 360  
 cat agcat gt tctt gt aat c at caaccagc tt aagagt ac act t ct ccac aat cgt ggt g 420  
 gt at gcaacg gagaaggct c tgcaat aggt aact t act ca t cagccat ca gcat agacca 480  
 15 at aat ctt aa tt agat ctt g tcaacgagt a aacaccatt g tact aat t gt act aat acaa 540  
 aagcaacaga aat t ct t gt a aacct t caat caaat t caat t caacagaca gagtt aaaca 600  
 aaacccagac taacaaacaa cagtt cact t t caat aat ag tt aacat t ca gacagat at a 660  
 20 acat ggaaat aaacaagata acacaccact tttt tttt g tttt cccacaa gt at cat cct 720  
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25 <210> 199  
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 <212> DNA  
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30 <400> 199

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 35 gcctttt gat cctttt aat g agt caggcct t ggggt ggaca tttt gat agt g cagatt aagg 180  
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 ggaaggcct t caaaggagag gaat gacaca agat ct t agt t gggacaat g ct gct cagca 300  
 40 gt at gaggag gt gct t ct t g ct gccaaat a ccaat ggt ga act tttt ggca tttt at t ccat 360  
 ct aagaagac tt gt aaaat g gagct gct aa tt cat gtt ga at act t ccag t gt act gat t 420  
 gt t gt gtt ag ggaaaagaac t gt gcaagt t gt t aaat t t at aggt t ac agt t agagcc 480  
 45 tttttt at gg gaagt gggaa ggccaaat tt t ggt gct gga tt at gt aact gt aat at agt 540  
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50 <210> 200  
 <211> 915  
 <212> DNA  
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55 <400> 200

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 5 aagtcgagct cacgatgcag aggattt gga ggacaatcca ccaggaaat a cagcatat ga 180  
 ttgtatggaa accagt agac aaaat agtcc attgtgttca tctatgtccc catgtgcagt 240  
 tgaagggttgt ctgtct aagg ggtttgcagg actactcaac ttgttccat gtgtagttg 300  
 10 gt agggagag aagggttcc tttgtgaa gctccctct tatagttaaa gaaat acata 360  
 gaagggttctt ttgctagtig atatctgttg atgtatcat tttgaatggg ctacaattac 420  
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 15 gagtgttaaa taactatcca ttgccttga taaaatgaat agaaatgttg tggtttgcct 540  
 ctatggaaa tttgtccat aattcccat tttcaatat gtatacttga aattgaaaac 600  
 taaggacat a tggtaacca gtatattaaa ttttagaact ttgat tgaat tttaaaaaaaa 660  
 20 attattccccg ggtttaccct atgaaaaaga aaaaatggaa aaactgtaaa tggat tttat 720  
 ttat tggat ttat ttttactggctc tttctaaca caacttttag ggacat aaat 780  
 ct aagt acca aattaccttc ccttatttt ttaaggccct gaacaacctc tgaat aaat a 840  
 25 aaagaaaatt ttttgc cca aatttttga accctcatta aaaaaaaaaa tgtaaatgag 900  
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30 <210> 201  
 <211> 668  
 <212> DNA  
 <213> Glycine max

35 <400> 201

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 aaaaaatgg ttaaacccgt atttctttag gagatattag gtgaattt gga t gattaaagg 120  
 40 aaaatggaa gaaaaatat cacaagttaa attcctgcta ataaaattaa tattttaaaca 180  
 actaacat ttt gctcatat aa aaaaccccaa tattttaa aatttaatct aaagcatttt 240  
 taaagttaact aaaaacatat ttat aagaa ttaaaatagt t gtaat tttat tttat 300  
 45 attattat ttttactttat aa aacatat aat ttat catta tatcaattt gcaat ttttta 360  
 tttatctt gattt cacat aatc ttatattaaac ttctt gtttt ctttttattt ctatgtaa 420  
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 50 aacgat tttt gtagttat tttt atatcaaaca ct aaatcaat tcatcagttt tctgggtcaa 540  
 accaacaat ttgatctggc atcttataac acaat tttt atggaaaaca catctaatgt 600  
 gattaaacaa ggacatcacg caacttggca gttaccactt ct tgcctt gctccaaat 660  
 55 ttttattt 668

<210> 202  
 <211> 941

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<212> DNA

<213> Glycine max

<400> 202

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10 ctgtgtcgct gtcaaataa gatgacaagg acaaggcat gagaagt aag agaaaactcaa 180  
gtgatattga gagtgagtat gtgaagtggc ttgattcatg gcctccgagg tcagtgattt 240  
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15 gatttggaaagc gacaaaaagg ccattcattt gggtgcttag aggtgcata ggaagagagg 360  
agatggagaa gtggctgttg gaagatgggt ttgaagagag ggtgaaaggg agagggttt 420  
tgatcaaggg ttgggtgccca caagtgttgc tcttatcaca tagagcaata ggacgttca 480  
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25 aggtgaccag agaaaaatgtt ctggattctat tttgaaaggta atggagaat ggccaaaaaa 720  
aaaaaaaaata taggaaaggg cttaaagtat tccgcattt ggcaggaaa gcaaaaaaaa 780  
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30 ccaagttaaacc ccagggttct atttttgtt ttcaacacca attgcatttc tcaagggtca 900  
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<210> 203

<211> 652

<212> DNA

<213> Glycine max

<400> 203

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45 aaaaaatitc ttttttcaga ttggtcaaag ctaaagaaga tagaggat tttttatcc 180  
ggcaacgaat tttagggacc acttccctcg tctttgtta acatgacatc tcttcggag 240  
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50 tcacttgaat attttggttt tacagaaaac caatttgaag ttccgtttc tttctcaaca 360  
tttgc当地atc attcaaagat caagtgtatc gacggtgtag gaaacagat catattggac 420  
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55 tcaacaactg aaactaagtc tcttccactc cccaaatttc ttctataccaa aacagttaa 540  
atcagcctag acttcagtag ttggaaatgtt gaaaggact ttccatttttggaa 600

aacaacacaa aaat gact ga agct ct gt tt agaaat t gct ctt cact gg tg 652

<210> 204

<211> 699

<212> DNA

<213> Glycine max

<400> 204

10

t gcat gcct g cagcaat ct c agct aaacaa gacagg t t c agcgaaaaac aaat t ccat 60

t gct cacaac acacagt t cg gcaaaaaagc t t catt aaac t caaccat gc cat caccat 120

15

t catt ct gt g tagct t gtt ttt cat t ca t gaaat t ct cagcat t t ca aacccact c 180

t t gccc ccc ccaaacc c act taaaat t cccaccaaa ccact ccaa cccacat ccc 240

catt t ct cag aact ccat t t cact ct acc t at cacgct t cgccgt cata a aagt t t caaa 300

20

t t gggcgca t t cccggcga cccagcaacc gccgcaact c ct t t ggaag aagct cct cc 360

gt gat cgcaa ggt aaact ct aat cagat t c ccaacgaccc t t t ct ct gtt t cggcaat g 420

gcgt t gaaga gagt ggt gtt ggt gat cagg ggt t gacaa t gt ggt t gaa gt t gaaaaac 480

25

caaagt ct aa gct t t gcgt gagt ct gt t t gt ggaat aa gt t ggagaat t gggct gacc 540

agt acaagag ggat gt t gag t att ggggt g taggat ct gg t cct at at t c act gt t at g 600

aagat t ccat t ggaggt gt c aagagggt gg t t gtt at gt agaccagat t ct gaaaagaa 660

30

gcaaggt aaa cat ggct agg gagat ggaga gt gggaaat a 699

<210> 205

<211> 578

<212> DNA

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<213> G yci ne max

<400> 205

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aaacgt gt t g aaat gaat at t t gggat ct t aacaact t ca ct aagat ct g gaat t ccaa 60

cct gt gagt c t t act t caa ccct t act t t t c a c t ct gc t a a t a t t g t t a a c t t a t ct 120

t cct ccagt t t acct t t t gt t ggt gcacat t t t t a g t g c t t g a t t t a c a a a t c c t t t 180

45

aacct at gca gccaact aag gat aacct cg gt at a t t a c a c t a c t t g t t t c a c a t c t g 240

c t a c a t t c t t a t g a a a g a t g a c c a t c g a a a t t t g t t g c t g g c a c c a a c a g c c a t c a g g 300

t a a a t t t a a t t g g t a g a t t c t t c t a t t a a a g a t g a a g a a a t a t a t t t t a t 360

50

c t a a t a a t g c a t g g a a t a t t c c t c t t g a t a t c t t g a a a a a a c a t g t g c t g g a g a c t a t 420

t g a g a a g g a t t a a t a a g t t t c t c t t a a c c t a g a a t t a c a g a g a a t g a t a t t t 480

t g a g a a t t t t t t t g c a a a a g a a c a c t c a a g t a c a t g t c c t t c a a g t c a c a t g a g a t c t 540

55

t t c c c a t t t t a t t a a c a a t a t a a a t g a g t a a t a t g a 578

<210> 206

<211> 754

&lt;212&gt; DNA

&lt;213&gt; Glycine max

&lt;400&gt; 206

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       tt ct t t gaca aaaat t cat c cacaagt gt t t gt aagt aat t gat aacat c t t ggt ccaa 120  
 10                  acat cagagc gagacagct g aat ct t gt ca gcagccccag aagaaat acc aacagaacca 180  
       ggt cgaacct gaaaaagcca at cat aat ct acaaat at t a aaaaat at aa agcaat t cat 240  
       cat aat aggt aaat at acca acact gaagc at ccct acct gatt gaggt a agt aacct t g 300  
 15                  at aaaccagg t ggccct aag caat ggaaca tt att t cct ga taagaacct c t aaaagt gat 360  
       gt cct t t t at aaccat gagg aacat gat ca gccaaagagc gt aat cgct t gt gct gct ga 420  
       gat aaacct t gt aat aaaaat t t t aact gag tt act aaaga t acat t gaaa caccaagt aa 480  
 20                  t gtt caaaaat gt agt at t t a aggat t t ac agccacaaca acct gt cagc at caacaaca 540  
       aaaaacct aa ggccct att t c ggggt ggct g agt t t t ct gt t t ccgt t t t aaaaat gct at 600  
       t t caaaaat gg aaggt gt t cg gct aaaaat gg tt gcgagtt g gat t t t at c t gtt t t t aaaa 660  
 25                  acagt t gt ca ccccat t t t a taaacaat aa aaat aggt t t tatgt t t t at t t t aagg 720  
       tt act t ct at ccct acct ca acgat ct acg ct gc 754

&lt;210&gt; 207

&lt;211&gt; 798

&lt;212&gt; DNA

&lt;213&gt; Glycine max

&lt;220&gt;

&lt;221&gt; unsure

&lt;222&gt; (1)..(798)

&lt;223&gt; unsure at all n locations; n = a, t, c, or g

&lt;400&gt; 207

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5 tt at ggagt g att aact gt g caagaaaat a att gt at gtt taact gcct g cat acat cgc 180  
t at gct aat t ct gt cct t ca caaat ct ttt cacaact t gt tt gt gcat ga ct ct cgaaa 240  
agacat gct a acat gcat at ggt gaagat a agaaaat t aaa agaaaat t gg aaaaggaaag 300  
10 at gat at agc aat t aaata tttttaaga tagat gt at g att gct at at cagaaaagg 360  
tt agt aaaac tagat t gat g t ggt t gt cct ggt t ct ct t t gat agt aaaa ttttggatt 420  
t cct t gaca cat at ggt gt at ct t t gat g gt at gt agt a at accct gga att cagccct 480  
15 aaatt aaagc attttt t at t ct ct t gga aggt agat at ct t ccaagga gt t accct tc 540  
cagat gct cc aagat t t gga aaggggggt t gggggggcggt at gaat t gga caggt at gaa 600  
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20 tt at acct ac gct t gcgt ac gt ggcaagt t tatttcttgt a cgcctt ggc ct t gt agcac 720  
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25

<210> 208  
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<400> 208

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## EP 2 134 870 B1

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 5 aaaaatgggc tcaaggtaaa tgtgcata gaggat t tga gaggggttat cacaagtatt 180  
 aaaaaggaaaa gggaggaaga t gttgat t tga agaagt aacg aaagttagtg tgggttcaa 240  
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 10 tt cagaccctt ggggc ttag cttaatagaac aagagaacat aat ttcctt taagaaaaagg 360  
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 15 attgtattct ttat tttcttgc t cgggttttga tg tttctatt taat tttact ggtggtgaga 540  
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 20 gt agt ggtat act caaaaac t gcaaaat t agctt acag ttcatctgca ttttttggtg 720  
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 25 ttgaatcaga aaactgtgct accaat tttc t cagccatgt atgatgaa gacagat t g 900  
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 30 tagttatcc aagatttctat gataaaatgt aaaggctt t c agctat t tga t caaaaactt 1080  
 t gatggttgc tctctt cgtat tt 1102

&lt;210&gt; 209

&lt;211&gt; 697

&lt;212&gt; DNA

&lt;213&gt; Glycine max

&lt;400&gt; 209

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 gaatataaaa aacaattatc t aattcatca agatcaataa aagt aatcat attagttaat 120  
 45 ttatcataaa ttttaattat attccatgac t atccatgat t acttttcaa aagat aat t 180  
 ttcaaccaat aagcttacct t gttttat t g atagctcaat aaagacgcca cttttatggg 240  
 aaaaatgaatg t aatcattag ggataaaaaa t aattttagc aagaaaaat t acctttt g 300  
 50 ttctgatata t gggatggaa gtggatggaa gt agt at aat gatactcatg ccattggat 360  
 atctatatttg cacatgctaa tttcaagctat tggaaaatg t ggaaaagag atgaaat t at 420  
 aggcatgcaa ct ttaatcaa ttactat t t ctagggtgac aatgtgtt a gctctaaccc 480

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5	gt ggt at at t gaattttgat ttat tttgtc aat cagatta gatt cgaacc aact at ttag	660
	t at aaaaaat actttcaaa gcat tcaaa ttttcaa	697
10	<210> 210	
	<211> 934	
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20	ctgtgaatg ttgttgagtt attccaagat ttgtgtgctg cttttttttt ttttttttt	180
	ggagt ggtta ggtgtaacct ttgtacttat ttttggatgc agacaaggag aaacttggct	240
	aaactggagc tctaccgaaa gtttacaaac acgctcgccg tgcgtgtgt gctgtccatt	300
25	gcgtggattt gctttgaggt agtttttggaa aaaaatattt tttttttttt tttttttttt	360
	at ttaggaga tttgctctat ctgtaggagt ttccaaacat tttcatttat gtat ttttat	420
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30	actgatccat tttttttttt gttttttttt gttttttttt gttttttttt gttttttttt	540
	ctttcatatg ctctcttgggt ggtgatgc at cttttttttt tttttttttt tttttttttt	600
	aggtacttgt tttttttttt gttttttttt gttttttttt gttttttttt gttttttttt	660
35	ttttttttttt gttttttttt gttttttttt gttttttttt gttttttttt gttttttttt	720
	actagttttt gttttttttt gttttttttt gttttttttt gttttttttt gttttttttt	780
	ggatattgtc cgccacttag ggggggggtat ttttgcgtgt aaatatcac agcaagtgtc	840
40	ttttttttttt aaagaaaaac taacccctt actttttttt ttat tttttttt gttttttttt	900
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	<213> Glycine max	
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5 catt act t gt aat aaaagt g gct agt tt at tttt gaacat at gt att aat aat agt t gca 180  
cat gt gt gag at gat gat ac at gt gcat cc tgaact ct t g gaaagg t gct aaaat gagaa 240  
act at ct ttt taaat caggc t act tt agt t ggt t gaatt c tgaat aaagt cct ct aat tt 300  
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gggaaaaaga tt ccccagct ttt aaat gga cccagtt gt tcaa at cc cat gcaacat 420  
*Dna 62*

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agatt ct aga tatt cat t gc caaat aaaca acgaact tt g acagaagct g aaccagaaaa 600  
20 gact aacaat aact gct t aa aat aaaat ca t gcccagacac tgaat t at gt ggt cct at t g 660  
att aact gaa agct cct t cc gct t at at cg gaccat cact tt cccct t ca ct cccacaac 720  
cat ggcattt c caat ccat ct ct ggaaaagg tgacaccaac tccagct caa aatt cct aag 780  
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5 tttt acat ct at at caaaca at acacat t g aaat agcaac aat gact at t t caaccat tt 180  
ttt at aagaa aaaat t t ac agat aaaagg gagaaaaaaa aaat t ggaaa ggacaaggta 240  
10 tttcccaa ac caaggggagg t gt gggat ac t gaat ct t ca gct t aagccc aat aat t gag 300  
ct gaat agga t aaggat at g t t aagagt at t at aat aaag ggct t t agaa gt t agt t gt a 360  
t gt acat t t a taaaat t t t t cacaat ct t t ct t t a t caaaat t g agt gt gat ct 420  
15 tttcait t ct a aat gacagca tt act ggaat att t t aat a t t t t t t gg aaaat agat t 480  
tt gaaaaact t at at gggct t gggccacat gcct t at gt acat aacct a gt aat at aaa 540  
t at gagagt c act acacatt ggagt ggagt ggat cccatt at agt t t att gacacaccc 600  
20 t t t at ct t c t ct ct ct a ct ct acat gg t at caagagc caggt aggt t t ggt gccat 660  
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5 tcttcctgca ttgtcataca acattatgtt gatttacgtg taatgtttgg tttcggttca 240  
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aaacaagaaa caaagtgaaa acagaaaaatg ttttctcaaa ccaaacggtg agtgcac 660  
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20 attctatcaa aattctagat aattttatata ctatcaacag agaccctttt naatacctgc 780  
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## Claims

- 20 1. A method of screening and selecting a soybean plant or soybean seed for maturity group association comprising:
  - (a) assaying genomic nucleic acids of said soybean plant or soybean seed for the presence of a genomic maturity marker genetically linked to a genomic region, wherein said genomic region is associated with a plant maturity group, said genomic maturity marker is within 10 cM, or within 10,000 kilobases of any of SEQ ID NOs: 25 143-213;
  - (b) determining whether said genomic maturity marker is homozygous or heterozygous;
  - (c) and selecting said soybean plant or seed based on said determination.
- 30 2. The method of claim 1, wherein said genomic maturity marker is within 1 cM or 5 cM, or within 5 kilobases, 10 kilobases, 20 kilobases, 30 kilobases, 100 kilobases, 500 kilobases or 1,000 kilobases of any of SEQ ID NOs: 143-213.
- 35 3. The method of claim 1 or 2, wherein parents of said soybean plant or seed differ in soybean plant maturity by over 10 days.
- 40 4. The method of any one of Claims 1 to 3, wherein said genomic maturity marker is associated with maturity genomic region 1 located on linkage group C2 identifiable using SEQ ID NO: 143-155, or complements thereof.
- 45 5. The method of Claim 4, wherein said maturity genomic region 1 can be monitored by assaying for an allele of 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, or more markers of SEQ ID NO:143 through SEQ ID NO: 155, or complements thereof.
6. The method of any one of Claims 1 to 3, wherein said genomic maturity marker is associated with maturity genomic region 2 located on linkage group O identifiable using SEQ ID NO: 156-161, or complements thereof.
- 45 7. The method of Claim 6, wherein said maturity genomic region 2 can be monitored by assaying for an allele of 1, 2, 3, 4, 5, 6, or more markers of SEQ ID NO: 156 through SEQ ID NO: 161, or complements thereof.
8. The method of any one of Claims 1 to 3, wherein said genomic maturity marker is associated with maturity genomic region 3 located on linkage group L identifiable using SEQ ID NO: 162-174.
- 50 9. The method of Claim 8, wherein said maturity genomic region 3 can be monitored by assaying for an allele of 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, or more markers of SEQ ID NO:162 through SEQ ID NO: 174, or complements thereof.
- 55 10. The method of Claim 3, wherein said any of SEQ ID NOs: 143-213 is selected from SEQ ID NOs: 149, 151, 158, 169, 178, 187, 192, 202, 204, and complements thereof.
11. The method of any one of Claims 1 to 3, further comprising associating said soybean plant or said seed with an indeterminate or determinate growth habit if said genomic maturity marker is associated with maturity genomic

region 3 located on linkage group L identifiable using SEQ ID NO: 162 through SEQ ID NO: 174, or complements thereof.

5       **12.** The method of Claim 11, wherein said maturity genomic region 3 is **characterized by** a G at position 433 in marker SEQ ID NO: 169, or complements thereof.

10      **13.** The method of any one of Claims 1 to 12, further comprising obtaining DNA from said soybean plant or seed using a non-destructive method.

15      **14.** The method of Claim 1, wherein said maturity group is between 0.0-III.9 or between 00.0 - III.0.

15      **15.** The method of Claim 1, wherein said soybean plant reaches maturity at least 5 days before a soybean plant that is homozygous dominant within maturity genomic region 1 located on linkage group C2 identifiable using SEQ ID NO: 142-153, or complements thereof, homozygous dominant within maturity genomic region 2 located on linkage group O identifiable using SEQ ID NO: 156-161, or complements thereof, and is grown under the same environmental conditions.

**16.** The method of Claim 1, wherein

20      (i) said selected soybean plant or seed is homozygous recessive at maturity genomic region 1 located on linkage group C2 identifiable using SEQ ID NO: 142-153, or complements thereof, and homozygous recessive at maturity genomic region 2 located on linkage group O identifiable using SEQ ID NO: 156-161, or complements thereof, and said associated maturity group is between 0.5 - II.0; or  
 25      (ii) said selected soybean plant or seed is homozygous recessive at maturity genomic region 1 and heterozygous dominant at maturity genomic region 2 and said associated maturity group is between I.5 - II.9.

**17.** The method of Claim 5, wherein said allele is associated with SEQ ID NO:151, or complements thereof, and wherein said soybean plant or a soybean plant grown from said soybean seed has a maturity group of III-VI.

30      **18.** The method of Claim 7, wherein said allele is associated with SEQ ID NOs: 156 or 158, or complements thereof.

### Patentansprüche

35      **1.** Verfahren zum Screening und Auswählen einer Sojapflanze oder eines Sojasamens für die Zuordnung zu einer Reifegruppe, umfassend:

40      (a) Testen genomicscher Nucleinsäuren der Sojapflanze oder des Sojasamens auf die Anwesenheit eines genomicschen Reifemarkers, der genetisch mit einem Genombereich verknüpft ist, wobei der Genombereich mit einer Pflanzenreifegruppe assoziiert ist, wobei sich der genomicsche Reifemarker innerhalb von 10 cM oder innerhalb von 10 000 Kilobasen von einer von SEQ ID Nr. 143-213 befindet;  
 45      (b) Bestimmen, ob der genomicsche Reifemarker homozygot oder heterozygot ist;  
 50      (c) und Auswählen der Sojapflanze oder des Sojasamens auf der Grundlage dieser Bestimmung.

**2.** Verfahren gemäß Anspruch 1, wobei sich der genomicsche Reifemarker innerhalb von 1 cM oder 5 cM oder innerhalb von 5 Kilobasen, 10 Kilobasen, 20 Kilobasen, 30 Kilobasen, 100 Kilobasen, 500 Kilobasen oder 1000 Kilobasen von einer von SEQ ID Nr. 143-213 befindet.

**3.** Verfahren gemäß Anspruch 1 oder 2, wobei sich Elternpflanzen der Sojapflanze oder des Sojasamens in der Sojapflanzenreife um über 10 Tage voneinander unterscheiden.

**4.** Verfahren gemäß einem der Ansprüche 1 bis 3, wobei der genomicsche Reifemarker mit Genomreifebereich 1 assoziiert ist, der sich auf der Kopplungsgruppe C2 befindet, die unter Verwendung von SEQ ID Nr. 143-155 oder Komplementen davon identifizierbar ist.

**5.** Verfahren gemäß Anspruch 4, wobei der Genomreifebereich 1 überwacht werden kann, indem man auf ein Allel von 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13 oder mehr Markern von SEQ ID Nr. 143 bis SEQ ID Nr. 155 oder Komplementen davon testet.

6. Verfahren gemäß einem der Ansprüche 1 bis 3, wobei der genomische Reifemarker mit Genomreifebereich 2 assoziiert ist, der sich auf der Kopplungsgruppe O befindet, die unter Verwendung von SEQ ID Nr. 156-161 oder Komplementen davon identifizierbar ist.
- 5 7. Verfahren gemäß Anspruch 6, wobei der Genomreifebereich 2 überwacht werden kann, indem man auf ein Allel von 1, 2, 3, 4, 5, 6 oder mehr Markern von SEQ ID Nr. 156 bis SEQ ID Nr. 161 oder Komplementen davon testet.
- 10 8. Verfahren gemäß einem der Ansprüche 1 bis 3, wobei der genomische Reifemarker mit Genomreifebereich 3 assoziiert ist, der sich auf der Kopplungsgruppe L befindet, die unter Verwendung von SEQ ID Nr. 162-174 identifizierbar ist.
- 15 9. Verfahren gemäß Anspruch 8, wobei der Genomreifebereich 3 überwacht werden kann, indem man auf ein Allel von 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13 oder mehr Markern von SEQ ID Nr. 162 bis SEQ ID Nr. 174 oder Komplementen davon testet.
- 10 10. Verfahren gemäß Anspruch 3, wobei die eine von SEQ ID Nr. 143-213 aus SEQ ID Nr. 149, 151, 158, 169, 178, 187, 192, 202, 204 und Komplementen davon ausgewählt ist.
- 20 11. Verfahren gemäß einem der Ansprüche 1 bis 3, weiterhin umfassend das Zuordnen der Sojapflanze oder des Sojasamens zu einer unbegrenzten oder begrenzten Wuchsform, wenn der genomische Reifemarker mit Genomreifebereich 3 assoziiert ist, der sich auf der Kopplungsgruppe L befindet, die unter Verwendung von SEQ ID Nr. 162 bis SEQ ID Nr. 174 oder Komplementen davon identifizierbar ist.
- 25 12. Verfahren gemäß Anspruch 11, wobei der Genomreifebereich 3 durch ein G auf Position 433 im Marker SEQ ID Nr. 169 oder Komplementen davon gekennzeichnet ist.
13. Verfahren gemäß einem der Ansprüche 1 bis 12, weiterhin umfassend das Gewinnen von DNA aus der Sojapflanze oder dem Sojasamen mit Hilfe eines zerstörungsfreien Verfahrens.
- 30 14. Verfahren gemäß Anspruch 1, wobei die Reifegruppe zwischen 0.0 und III.9 oder zwischen 00.0 und III.0 liegt.
15. Verfahren gemäß Anspruch 1, wobei die Sojapflanze ihre Reife wenigstens 5 Tage vor einer Sojapflanze erreicht, die innerhalb des Genomreifebereichs 1, der sich auf der Kopplungsgruppe C2 befindet, die unter Verwendung von SEQ ID Nr. 142-153 oder Komplementen davon identifizierbar ist, homozygot dominant ist, innerhalb des Genomreifebereichs 2, der sich auf der Kopplungsgruppe O befindet, die unter Verwendung von SEQ ID Nr. 156-161 oder Komplementen davon identifizierbar ist, homozygot dominant ist und unter denselben Umgebungsbedingungen gewachsen ist.
- 40 16. Verfahren gemäß Anspruch 1, wobei
- (i) die ausgewählte Sojapflanze oder der ausgewählte Sojasamen im Genomreifebereich 1, der sich auf der Kopplungsgruppe C2 befindet, die unter Verwendung von SEQ ID Nr. 142-153 oder Komplementen davon identifizierbar ist, homozygot rezessiv ist und im Genomreifebereich 2, der sich auf der Kopplungsgruppe O befindet, die unter Verwendung von SEQ ID Nr. 156-161 oder Komplementen davon identifizierbar ist, homozygot rezessiv ist und die zugeordnete Reifegruppe zwischen 0.5 und II.0 liegt; oder
- 45 (ii) die ausgewählte Sojapflanze oder der ausgewählte Sojasamen im Genomreifebereich 1 homozygot rezessiv ist und im Genomreifebereich 2 heterozygot dominant ist und die zugeordnete Reifegruppe zwischen 1.5 und II.9 liegt.
- 50 17. Verfahren gemäß Anspruch 5, wobei das Allel mit SEQ ID Nr. 151 oder Komplementen davon assoziiert ist und wobei die Sojapflanze oder eine aus dem Sojasamen gewachsene Sojapflanze eine Reifegruppe von III-VI aufweist.
18. Verfahren gemäß Anspruch 7, wobei das Allel mit SEQ ID Nr. 156 oder 158 oder Komplementen davon assoziiert ist.

55

**Revendications**

1. Procédé de criblage et de sélection d'une plante de soja ou d'une graine de soja pour l'association du groupe de

maturité comprenant le fait :

- 5                   (a) d'analyser des acides nucléiques génomiques de ladite plante de soja ou graine de soja pour détecter la présence d'un marqueur génomique de maturité génétiquement lié à une région génomique, où ladite région génomique est associée à un groupe de maturité de plante, ledit marqueur génomique de maturité se situe à environ 10 cM, ou à environ 10000 kilobases de l'une des séquences SEQ ID NOs: 143 à 213 ;  
   (b) de déterminer si ledit marqueur génomique de maturité est homozygote ou hétérozygote ;  
   (c) et de sélectionner ladite plante ou graine de soja sur la base de ladite détermination.

10               **2.** Procédé de la revendication 1, dans lequel ledit marqueur génomique de maturité se situe à environ 1 cM ou 5 cM, ou à environ 5 kilobases, 10 kilobases, 20 kilobases, 30 kilobases, 100 kilobases, 500 kilobases ou 1000 kilobases de l'une des séquences SEQ ID NOs: 143 à 213.

15               **3.** Procédé de la revendication 1 ou 2, dans lequel les lignées parentales de ladite plante ou graine de soja diffèrent en terme de maturité de plante de soja de plus de 10 jours.

20               **4.** Procédé de l'une quelconque des revendications 1 à 3, dans lequel ledit marqueur génomique de maturité est associé à une région génomique de maturité 1 située sur un groupe de liaison C2 identifiable en utilisant les séquences SEQ ID NO: 143 à 155, ou leurs séquences complémentaires.

25               **5.** Procédé de la revendication 4, dans lequel ladite région génomique de maturité 1 peut être contrôlée par une analyse permettant de détecter un allèle de 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13 marqueur(s) ou plus de la séquence SEQ ID NO: 143 jusqu'à la séquence SEQ ID NO: 155, ou de leurs séquences complémentaires.

30               **6.** Procédé de l'une quelconque des revendications 1 à 3, dans lequel ledit marqueur génomique de maturité est associé à une région génomique de maturité 2 située sur un groupe de liaison O identifiable en utilisant les séquences SEQ ID NO: 156 à 161, ou leurs séquences complémentaires.

35               **7.** Procédé de la revendication 6, dans lequel ladite région génomique de maturité 2 peut être contrôlée par une analyse permettant de détecter un allèle de 1, 2, 3, 4, 5, 6 marqueur(s) ou plus de la séquence SEQ ID NO: 156 jusqu'à la séquence SEQ ID NO: 161, ou de leurs séquences complémentaires.

40               **8.** Procédé de l'une quelconque des revendications 1 à 3, dans lequel ledit marqueur génomique de maturité est associé à une région génomique de maturité 3 située sur un groupe de liaison L identifiable en utilisant les séquences SEQ ID NO: 162 à 174.

45               **9.** Procédé de la revendication 8, dans lequel ladite région génomique de maturité 3 peut être contrôlée par une analyse permettant de détecter un allèle de 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13 marqueur(s) ou plus de la séquence SEQ ID NO: 162 jusqu'à la SEQ ID NO: 174, ou de leurs séquences complémentaires.

50               **10.** Procédé de la revendication 3, dans lequel ladite l'une des séquences SEQ ID NOs: 143 à 213 est sélectionnée parmi les séquences SEQ ID NOs: 149, 151, 158, 169, 178, 187, 192, 202, 204, et leurs séquences complémentaires.

55               **11.** Procédé de l'une quelconque des revendications 1 à 3, comprenant en outre l'association de ladite plante de soja ou de ladite graine de soja à un port déterminé ou indéterminé si ledit marqueur génomique de maturité est associé à une région génomique de maturité 3 située sur un groupe de liaison L identifiable en utilisant les séquences allant de la SEQ ID NO: 162 à la SEQ ID NO : 174, ou leurs séquences complémentaires.

60               **12.** Procédé de la revendication 11, dans lequel ladite région génomique de maturité 3 est **caractérisée par** la présence de G à la position 433 dans la séquence SEQ ID NO: 169 du marqueur, ou ses séquences complémentaires.

65               **13.** Procédé de l'une quelconque des revendications 1 à 12, comprenant en outre le fait d'obtenir un ADN à partir de ladite plante ou graine de soja en utilisant un procédé non destructif.

70               **14.** Procédé de la revendication 1, dans lequel ledit groupe de maturité est compris entre 0.0 et III.9 ou entre 00.0 et III.O.

75               **15.** Procédé de la revendication 1, dans lequel ladite plante de soja atteint la maturité au moins 5 jours avant une plante de soja qui est homozygote dominante dans une région génomique de maturité 1 située sur un groupe de liaison

C2 identifiable en utilisant les séquences SEQ ID NO: 142 à 153, ou leurs séquences complémentaires, homozygote dominante dans une région génomique de maturité 2 située sur un groupe de liaison O identifiable en utilisant les séquences SEQ ID NO: 156 à 161, ou leurs séquences complémentaires, et qui est cultivée dans les mêmes conditions environnementales.

5

**16. Procédé de la revendication 1, dans lequel**

(i) ladite plante ou graine de soja sélectionnée est homozygote récessive au niveau d'une région génomique de maturité 1 située sur un groupe de liaison C2 identifiable en utilisant les séquences SEQ ID NO: 142 à 153, ou leurs séquences complémentaires, et homozygote récessive au niveau d'une région génomique de maturité 2 située sur un groupe de liaison O identifiable en utilisant les séquences SEQ ID NO: 156 à 161, ou leurs séquences complémentaires, et ledit groupe de maturité associé est compris entre 0.5 et II.0 ; ou  
10 (ii) ladite plante ou graine de soja sélectionnée est homozygote récessive au niveau d'une région génomique de maturité 1 et hétérozygote dominante au niveau d'une région génomique de maturité 2 et ledit groupe de maturité associé est compris entre I.5 et II.9.

15

**17. Procédé de la revendication 5, dans lequel ledit allèle est associé à la séquence SEQ ID NO: 151, ou à ses séquences complémentaires, et dans lequel ladite plante de soja ou une plante de soja cultivée à partir de ladite graine de soja a un groupe de maturité de III à VI.**

20

**18. Procédé de la revendication 7, dans lequel ledit allèle est associé aux séquences SEQ ID NO: 156 ou 158, ou à leurs séquences complémentaires.**

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30

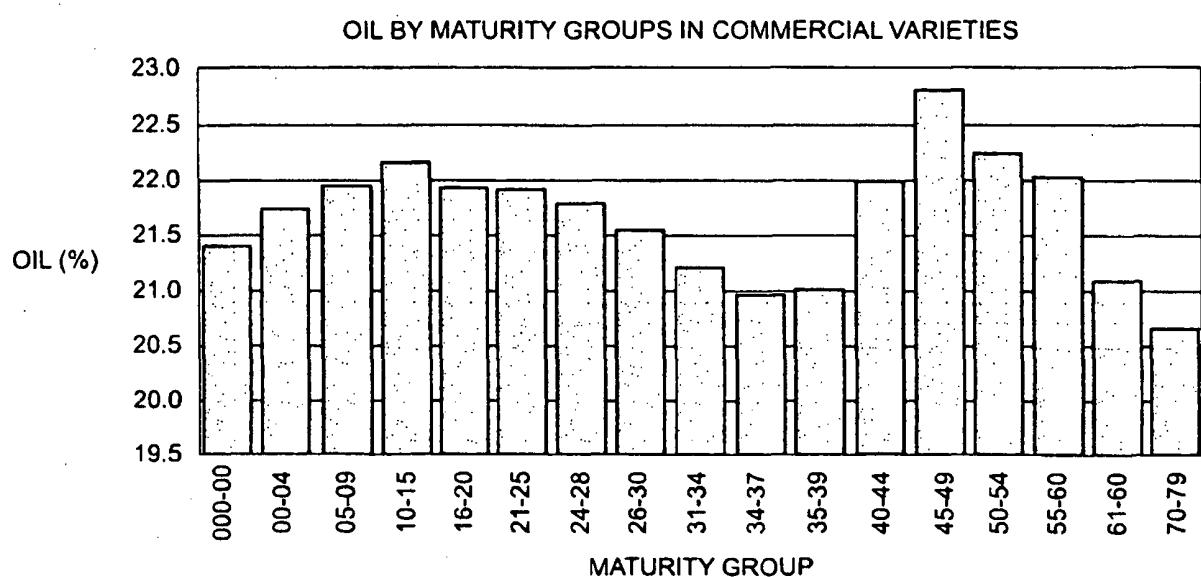
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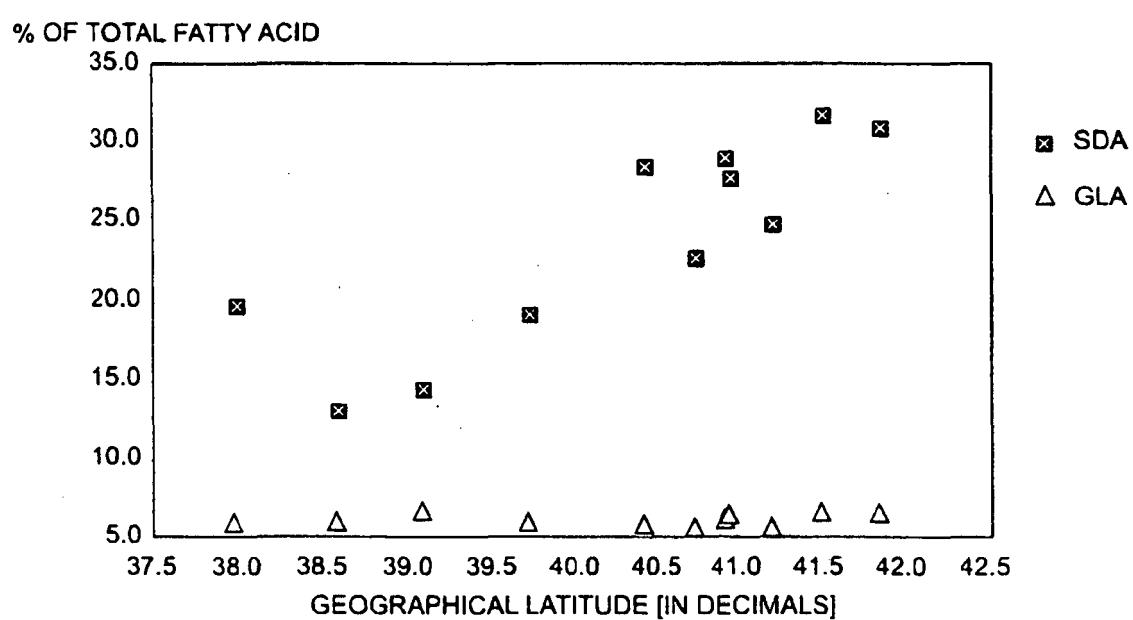
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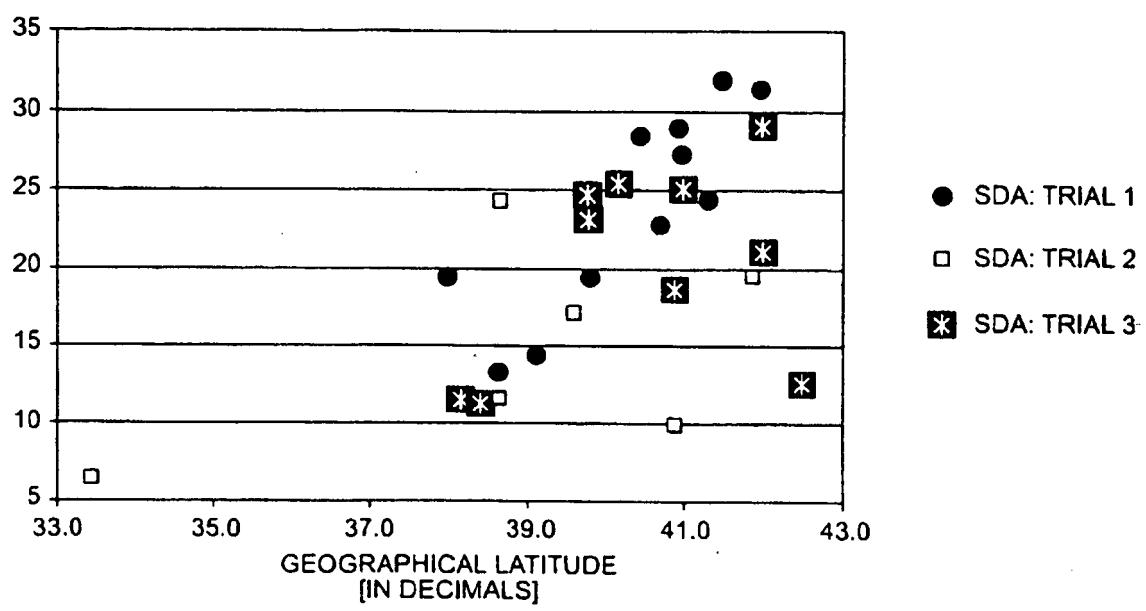


**FIG. 1**



**FIG. 2**

% OF TOTAL FATTY ACIDS



**FIG. 3**

## REFERENCES CITED IN THE DESCRIPTION

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